

The Effect of Repeated Vinyl Chloride Exposure on Rat Hepatic Metabolizing Enzymes¹

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The Effect of Repeated Vinyl Chloride Exposure on Rat Hepatic Metabolizing Enzymes. DU, J. T., TSENG, M. T., AND TAMBURRO, C. H. (1982). *Toxicol. Appl. Pharmacol.* 62, 1-10. Sprague-Dawley rats were exposed to 2.8% vinyl chloride for 2 (70 hr), 4 (140 hr), and 6 (210 hr) weeks to determine the sequential biochemical changes related to the oxidation and detoxification ability of hepatic tissue. Glutathione-S-transferase(s) activity using 1,2-epoxy-(p-nitrophenoxy)propane and p-nitrobenzyl chloride as substrates was elevated 17 to 24, 28, and 35 to 42% after 2, 4, and 6 weeks of exposure, respectively, suggesting enzyme(s) induction. Reduced glutathione, the major substrate required to conjugate the toxic metabolites of vinyl chloride, was also consistently elevated. Similarly, the activity of glutathione reductase, the enzyme necessary for the regeneration of reduced glutathione from its oxidized form, was also increased following vinyl chloride exposure. Albumin, the major protein involved with vinyl chloride metabolism, was reduced after vinyl chloride exposure, confirming reports of others that vinyl chloride metabolites destroy P-450. No abnormalities of standard clinical biochemical blood tests of liver function were found during 6 weeks of vinyl chloride exposure. The only consistent ultrastructural modification was the dilation of endoplasmic reticulum. The biochemical and ultrastructural alterations could reflect early hepatocellular adaptation to vinyl chloride exposure.

Vinyl chloride, at high concentrations, has been shown to be carcinogenic in both laboratory animals (Maltoni and Lefemine, 1975; Viola *et al.*, 1971) and man (Creech and Johnson, 1974). Present data support the metabolism of vinyl chloride by hepatic microsomal mixed-function oxidase system into toxic intermediates, chloroethylene oxide (Bolt *et al.*, 1975; Hefner *et al.*, 1975; Kappus *et al.*, 1976) and chloroacetaldehyde

(Gross and Freiberg, 1969). These two intermediates are considered to be the ultimate carcinogens (Barbin *et al.*, 1975; Jaeger *et al.*, 1974b; Van Duuren, 1975), to be mutagenic in bacterial systems (Elmore *et al.*, 1976; Greim *et al.*, 1975; Malaveille *et al.*, 1975; McCann *et al.*, 1975), to act as an alkylating agent by reacting with adenosine (Barbin *et al.*, 1975) and cytidine (Laib and Bolt, 1978), and to bind with protein (Bolt *et al.*, 1976; Kappus *et al.*, 1976; Watanabe *et al.*, 1978). Detoxification of these metabolites occurs mainly by conjugation with glutathione and is catalyzed by hepatic glutathione transferases; the conjugates are excreted in the urine as substituted cysteine derivatives (Watanabe *et al.*, 1976b,c; Green and Hathway, 1975, 1977). Chloroacetaldehyde can be further oxidized to chloro-

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sequential biochemical changes related to the hepatic oxidation and detoxification of vinyl chloride following prolonged exposure are reported here.

Animals and experimental design. Eight- to ten-week-old Sprague-Dawley male rats (~300 g), supplied by Laboratory Supply of Indianapolis, Indiana, were randomized prior to the experiment into three groups: a vinyl chloride-exposed group and the air-exposed group housed in identical chambers and a second control group housed in the University's Central Animal Care Center. The exposure level was 28,000 ppm vinyl chloride, 7 hr/day, 5 days/week for 2, 4, and 6 weeks. The exposure chambers were 4400-liter airtight vats. Vinyl chloride (~300 to 340 g) was added to the vat to give a time-weighted average concentration of $28,000 \pm 1000$ ppm. The chamber air was changed daily and the vinyl chloride concentration was determined by gas chromatography. The air was constantly circulated by a stirrer. The rats' respirations had negligible effect on the composition of the chamber's atmosphere because of the chamber's large volume. Animals were fed on standard laboratory chow pellets *ad libitum*.

killed approximately four after each day.

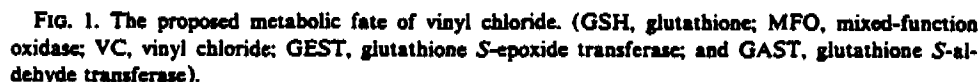
NADPH, glutathione, and glucose were obtained from Sigma Chemicals, St. Louis, Missouri; 1,2-epoxy-3-(*p*-nitrophenyl)propane was purchased from Eastman Kodak Company, New York; *p*-nitrobenzyl chloride was from Matheson, Coleman and Bell, East Rutherford, New Jersey; benzphetamine was from Schering Company, Kenilworth, New Jersey; and benzphetamine was from Schering Company, Kenilworth, New Jersey. Benzphetamine was used throughout.

Sample preparation and bioassays. Homogenates and subcellular fractions were prepared as described previously (Du et al., 1987). The homogenate was prepared from a single organ preparation. The remaining livable cells were stored at -450 concentrations were determined. The day following treatment, the glutathione (GSH) concentration, S-transferase, glutathione reductase, and glutathione S-transferase activities were determined. The fractionated frozen liver. For tissue, the livers from control and treated rats were frozen in an identical manner. Cytochromes P-450 (Sato, 1964), nonprotein sulphhydryl (Lindsay, 1968), and glutathione (Carlberg and Mannervik, 1978) the microsomal or cytosol fraction were prepared as described previously (Du et al., 1987).

Glutathione-S-transferase was assayed using the 100,000 $\times$ g supernatant. 3-(*p*-nitrophenoxy)propane and 3-(*p*-nitrophenyl)propane were the substrates for glutathione-S-transferase and glutathione-S-alkyltransferase, respectively. Enzyme activity was assayed by others (Habig *et al.*, 1974). All assays were linear function of time and timed for at least 2 min. 3-(*p*-nitrophenoxy)propane and 3-(*p*-nitrophenyl)propane were prepared in absolute ethanol. The concentration in the incubation mixture was 1 mM. Glutathione function oxidase activity was assayed in the same fraction by measuring the reduction of NADPH-dependent dehydrochloride of benzphetamine (Lu *et al.*, 1974). The activity was determined by the method of Lu *et al.* (1974). The serum clinical liver tests including aspartate aminotransferase, alanine aminotransferase, bilirubin, cholesterol, and triglyceride were determined by Technicon sequential analyzer (SMAC) system.

Light and electron microsc
were removed under ether an
cubes, placed immediately in
troxide (pH 7.4), and fixed

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killed approximately 20 hr after exposure at 1:00 pm each day.

NADPH, glutathione, and glutathione disulfide were obtained from Sigma Chemical Company, St. Louis, Missouri; 1,2-epoxy-3-(*p*-nitrophenoxy)propane was purchased from Eastman Kodak Company, Rochester, New York; *p*-nitrobenzyl chloride was obtained from Matheson, Coleman and Bell, East Rutherford, New Jersey; benzphetamine was donated by the Upjohn Company, Kalamazoo, Michigan. Double-distilled water was used throughout.

Sample preparation and biochemical determination. Homogenates and subcellular fractions were prepared as described previously (Du *et al.*, 1979). Each sample was prepared from a single organ and kept at 4°C during preparation. The remaining liver was frozen rapidly in liquid nitrogen and stored at -70°C. Cytochromes *P*-450 concentrations were determined in the frozen microsomal fractions the day following sacrifice. The glutathione (GSH) concentration, as well as glutathione-*S*-transferase, glutathione reductase, and mixed-function oxidase activities were determined in the freshly fractionated frozen liver. For the assays using frozen tissue, the livers from control and experimental rats were frozen in an identical manner for the same length of time. Cytochromes *P*-450 concentration (Omura and Sato, 1964), nonprotein sulfhydryl content (Sedlak and Lindsay, 1968), and glutathione reductase activity (Carlberg and Mannervik, 1975) were determined in the microsomal or cytosol fractions by methods described previously (Du *et al.*, 1979).

Glutathione-*S*-transferase activity was determined using the 100,000 × *g* supernatant fraction. 1,2-Epoxy-3-(*p*-nitrophenoxy)propane and *p*-nitrobenzyl chloride were the substrates for glutathione-*S*-epoxide transferase and glutathione-*S*-aralkyl transferase (GAST), respectively. Enzyme activity was determined as described by others (Habig *et al.*, 1974; Kaplowitz *et al.*, 1975). All assays were linear functions of protein concentration and timed for at least 2 min. Solutions of 1,2-epoxy-3-(*p*-nitrophenoxy)propane and *p*-nitrobenzyl chloride were prepared in absolute ethanol; the final ethanol concentration in the incubation mixture was 0.5%. Mixed-function oxidase activity was estimated in the microsomal fraction by measuring NADPH disappearance in the NADPH-dependent demethylation reaction of benzphetamine (Lu *et al.*, 1972). The protein content was determined by the method of Lowry *et al.* (1951). The serum clinical liver tests including aspartate aminotransferase, alanine aminotransferase, alkaline phosphatase, bilirubin, cholesterol, and triglyceride were determined by Technicon sequential multiple analyzer computer (SMAC) system.

Light and electron microscopy. Small strips of liver were removed under ether anesthesia, sliced into small cubes, placed immediately in ice-cold 1% osmium tetroxide (pH 7.4), and fixed for 2 hr at 4°C. Subse-

quently, samples were washed overnight in phosphate buffer, dehydrated in ascending alcohol, and embedded in Epon. Tissue blocks were polymerized at 60°C for 2 days. Thin sections were cut with a diamond knife and stained with uranyl acetate and lead citrate before examination on a Philips 300 electron microscope. For ultrastructural analysis, three rats randomly selected from controls and groups exposed for 2, 4, and 6 weeks to vinyl chloride were studied.

For light microscopy, a block of tissue was fixed in buffered formalin and processed routinely for paraffin embedding. Sections 6 μm thick were stained with hematoxylin and eosin.

Statistical analysis. Analysis of variance was performed for the various groups at the different time periods and multiple comparisons were performed based on the results of the analysis of variance.

RESULTS

The protein content (mg protein/g liver) in the subcellular fractions in both control and vinyl chloride-exposed groups was the same throughout the exposure (data not shown); the enzymatic results, therefore, are expressed as micromoles of substrate converted per minute per milligram of protein.

There were no statistical differences between the normal and air-exposed groups in the glutathione and cytochromes *P*-450 contents or in any of the enzyme activities. The nonprotein sulfhydryl content (Table 1) was significantly elevated from 26 to 54% at 2, 4, and 6 weeks in the vinyl chloride-exposed group compared to both control groups. Although the nonprotein sulfhydryl content increased with exposure, the increases were not statistically significant. Glutathione reductase activity (Table 1) in the exposed group was increased by 53 to 77% at all three time periods. The increase in glutathione reductase was the same at 2 and 4 weeks of exposure but showed a further significant increase after 6 weeks of exposure. Glutathione-*S*-epoxide transferase (GEST, Table 1) and glutathione-*S*-aralkyl transferase (GAST, Table 1) activities were significantly higher than controls after 6 weeks of exposure, 37 and 45%, respectively. The cytochromes *P*-450 content, on the other hand,

TABLE I

SEQUENTIAL CHANGES IN HEPATIC NONPROTEIN SULFHYDRYL, CYTOCHROMES P-450 CONTENT AND ACTIVITIES OF GLUTATHIONE REDUCTASE AND GLUTATHIONE-S-TRANSFERASES (EPOXIDE AND ARLKYL) IN RATS EXPOSED TO VINYL CHLORIDE*

Treatment		Time (weeks)			
		0	2	4	6
Nonprotein sulfhydryl ($\mu\text{mol/g liver}$)	Normal control	7.9 ± 0.3	7.8 ± 0.4^b	7.1 ± 0.4^b	6.9 ± 0.4^b
	Vinyl chloride-exposed	—	$9.4 \pm 0.2^{b,c}$	$10.2 \pm 0.6^{b,c}$	$11.4 \pm 0.6^{b,c}$
	Air control	—	7.1 ± 0.3^c	6.9 ± 0.4^c	7.9 ± 0.3^c
Glutathione reductase ($100 \times \mu\text{mol/min/mg protein}$)	Normal control	5.0 ± 0.2	4.3 ± 0.4^b	4.2 ± 0.4^b	4.8 ± 0.3^b
	Vinyl chloride-exposed	—	$6.7 \pm 0.6^{b,c}$	$6.3 \pm 0.5^{b,c}$	$8.9 \pm 0.7^{b,c,d}$
	Air control	—	4.5 ± 0.2^c	3.7 ± 0.3^c	5.3 ± 0.4^c
GEST ($100 \times \mu\text{mol/min/mg protein}$)	Normal control	9.1 ± 1.6	7.8 ± 0.4	7.5 ± 1.0	7.7 ± 1.0^b
	Vinyl chloride-exposed	—	9.1 ± 1.7	9.7 ± 0.7^c	$11.0 \pm 1.3^{b,c}$
	Air control	—	8.1 ± 1.2	6.3 ± 0.7^c	8.4 ± 0.9^c
GAST ($10 \times \mu\text{mol/min/mg protein}$)	Normal control	2.4 ± 0.4	2.1 ± 0.3	1.9 ± 0.2	2.4 ± 0.3^b
	Vinyl chloride-exposed	—	2.6 ± 0.4	2.4 ± 0.3	$3.2 \pm 0.1^{b,c}$
	Air control	—	2.2 ± 0.2	1.9 ± 0.2	2.1 ± 0.2^c
Cytochrome P-450 (nmol/g liver)	Normal control	17.0 ± 3.7	17.1 ± 1.7	19.7 ± 1.3^b	15.5 ± 1.4^b
	Vinyl chloride-exposed	—	13.2 ± 1.1	15.3 ± 1.3^b	$10.6 \pm 1.0^{b,c}$
	Air control	—	15.5 ± 1.3	19.6 ± 2.7	14.3 ± 1.8^c

* Rats were exposed to 28,000 ppm of vinyl chloride; normal controls and the air controls were exposed to air only. Each number represents the mean and the SEM from a group of six rats.

^b Normal vs vinyl chloride-exposed, $p < 0.05$.

^c Air control vs vinyl chloride-exposed, $p < 0.05$.

^d Vinyl chloride (6 weeks) exposed vs vinyl chloride (2 and 4 weeks) exposed, $p < 0.05$.

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was significantly lower: 6 weeks of exposure to 1). No differences were mixed-function oxidase serum clinical liver te exposure, the two cont: weight but the vinyl cl did not (Table 2). Afte the normal control gr: inal care center had more weight than eit vinyl chloride-exposed of exposure, however, posed group failed to mal control group gain control group (Table: Morphological exam: bedral hepatocytes a plates interposed by va livers of the control r toarchitecture was in chloride exposure. H: rats contained a pro cleus, numerous ovoid of rough c: plasm: some aggregates of s: ticulum (SER), and v

Duration (week)	Body W.	
	Norm	Tr
2	Norm	VC-e
	VC-e	Air c
4	Norm	VC-e
	VC-e	Air c
6	Norm	VC-e
	VC-e	Air c

* Analysis of body weight the regression equation. (f
^a Normal control vs vin:
^b Air control vs vinyl ch
^c Air control vs normal

was significantly lower than controls after 6 weeks of exposure to vinyl chloride (Table 1). No differences were found in the hepatic mixed-function oxidase activity or in the serum clinical liver tests. After 2 weeks of exposure, the two control groups had gained weight but the vinyl chloride-exposed group did not (Table 2). After 4 weeks of exposure, the normal control group housed at the animal care center had gained significantly more weight than either the air-control or vinyl chloride-exposed group. After 6 weeks of exposure, however, the vinyl chloride-exposed group failed to gain weight; the normal control group gained more than the air-control group (Table 2).

Morphological examination revealed polyhedral hepatocytes arranged in irregular plates interposed by vascular sinusoids in the livers of the control rats. This general cytoarchitecture was maintained after vinyl chloride exposure. Hepatocytes in control rats contained a prominent spherical nucleus, numerous ovoid mitochondria, stacks of rough endoplasmic reticulum (RER), some aggregates of smooth endoplasmic reticulum (SER), and varying amounts of ly-

sosomes and glycogen particles (Fig. 2a). Few interstitial cells were scattered among the hepatocytes. These cells contained few cytoplasmic organelles and could be readily discerned at the light microscopic level by their hyperchromatic nuclei. The sinusoids were linked by fenestrated endothelium and some of the lining cells displayed phagocytic activity. After 2 to 6 weeks of vinyl chloride exposure, the principal organelle affected appeared to be the endoplasmic reticulum. Cisternae of the RER became dilated in a relatively small population of the hepatocytes in the 2-week treatment group. Four weeks after exposure to vinyl chloride, patches of dilated endoplasmic reticulum were prominently displayed in some hepatocytes (Fig. 2b). At this stage the SER was relatively unaffected. In the 6-week treatment group, vesiculation of SER and distention of RER were easily discernible in a large number of hepatocytes (Fig. 2c). However, other cell organelles showed no demonstrable change. These changes, though, are still beyond the resolving limit of the light microscope. The nonhepatocyte components showed minimum changes which

TABLE 2
BODY WEIGHTS OF RATS BEFORE AND AFTER VINYL CHLORIDE EXPOSURE^a

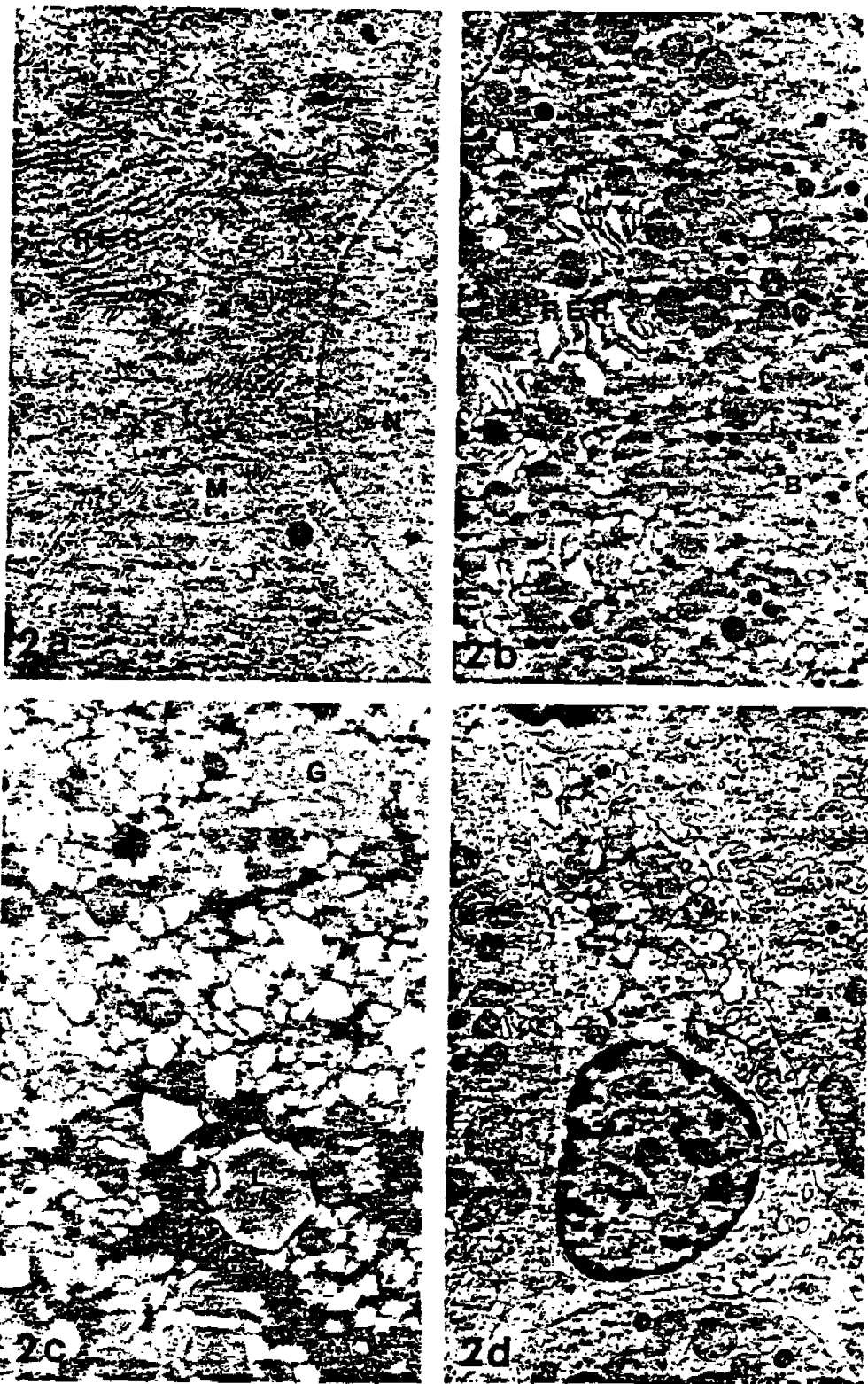
Duration (week)	Treatment	Initial weight (g)	Final weight (g)	Percentage gain
2	Normal control	400 ± 15	433 ± 16	8 ^b
	VC-exposed	405 ± 12	396 ± 14	-2 ^{b,c}
	Air control	396 ± 17	414 ± 18	5 ^c
4	Normal control	398 ± 8	450 ± 15	13 ^{b,d}
	VC-exposed	410 ± 16	421 ± 15	3 ^b
	Air control	395 ± 10	419 ± 5	6 ^d
6	Normal control	402 ± 14	486 ± 14	21 ^{b,d}
	VC-exposed	396 ± 9	398 ± 10	<1 ^{b,c}
	Air control	398 ± 14	449 ± 4	13 ^{c,d}

^a Analysis of body weight was by regression analysis followed by an analysis of variance on the residuals from the regression equation. (Residual = observed final weight - predicted final weight from regression equation.)

^b Normal control vs vinyl chloride exposed, $p < 0.05$.

^c Air control vs vinyl chloride exposed, $p < 0.05$.

^d Air control vs normal control, $p < 0.05$.



were characterized by
 mulation of lysosomes
 some of the sinusoids
 a greater tendency to
 the interstitial cells (

DISC

Glutathione conjugation
 pathway for the metabolism
 harmful electrophilic
 xenobiotics. Studies by W
 indicate this to be the
 activation of the vinyl

Glutathione *S*-transferase
 cytosol enzymes catalyze
 glutathione and electrophilic
 form less toxic and excreta-
 conjugates. Their activation
 exposure to xenobiotics
 could be a key determinant
 outcome of such exposure
 the longer arrow in the
 hepatic GST activation
 later increase in GST
 suggested that in
 exposures most of the
 diate is being added
 the later stages of
 chlorooxirane may
 yield more chlorooxirane
 react with other
 ferases or become
 chloroacetic acid
 sive chlorooxirane
 neously to form chloro-
 oxidized to chloro-
 turn may react with

FIG. 2. (a) Portals
 separated by many
 (b) Hepatocyte after
 two cells. Bile (B)
 chloride exposure.
 between SER and
 (GL) often accumu-
 in a vinyl chloride
 irregular in these

were characterized by an increased accumulation of lysosomal-like substances in some of the sinusoidal lining cells as well as a greater tendency to accumulate lipids in the interstitial cells (Fig. 2d).

DISCUSSION

Glutathione conjugation is an important pathway for the metabolism of potentially harmful electrophilic metabolites of xenobiotics. Studies by Watanabe *et al.* (1976b,c) indicate this to be the major route for inactivation of the vinyl chloride metabolites.

Glutathione-S-transferases are a group of cytosol enzymes catalyzing the reaction of glutathione and electrophilic compounds to form less toxic and more water-soluble conjugates. Their activity during chronic exposure to xenobiotics, like vinyl chloride, could be a key determinate in the ultimate outcome of such exposures as illustrated by the longer arrow in Fig. 1. The increase in hepatic GST activity at 4 weeks and the later increase in GAST activity at 6 weeks suggested that in the earlier stages of exposures most of the chlorooxirane intermediate is being adequately detoxified. During the later stages of chronic exposure, more chlorooxirane may become rearranged to yield more chloroacetaldehyde and, in turn, react with other available glutathione transferases or become further metabolized to chloroacetic acid. Alternatively, the excessive chlorooxirane could rearrange spontaneously to form chloroethanol and be further oxidized to chloroacetaldehyde, which in turn may react with glutathione, or be ox-

dized to monochloroacetic acid (Johnson, 1967). This would be consistent with the later increases in the aralkyl-transferases and the finding by Hefner *et al.* (1975) that monochloroacetic acid is found only in the urine of rats exposed for an extended time to higher levels (5000 ppm) of vinyl chloride. The increased use of alternate pathways, for chlorooxirane and chloroacetaldehyde detoxification, may reflect increased concentration of these active metabolites allowing greater opportunity for DNA injury.

A single exposure to vinyl chloride decreased hepatic nonprotein sulfhydryl compounds in rats (Watanabe, 1976a); similar decreases of glutathione concentrations were produced in rats by other xenobiotics such as 1,1-dichloroethylene (Jaeger *et al.*, 1974a; Reichert *et al.*, 1978) and acetaminophen (Mitchell *et al.*, 1973). In the present study, repeated exposure to vinyl chloride caused a significant increase of nonprotein sulfhydryl concentrations (Table 1) analogous to the elevation of glutathione concentrations seen after the administration of carcinogens to rats (Fiala *et al.*, 1976). In addition, the results showed that repeated exposure to vinyl chloride also caused an increase in hepatic glutathione-S-transferase activity (Table 1) similar to that seen after the administration of phenobarbital and 3-methylcholanthrene to rats (Mukhtar and Bresnick, 1976). These data suggest a mechanism for compensatory synthesis of hepatic glutathione and glutathione-S-transferases after repeated exposure to vinyl chloride.

The decreased concentration of cytochromes P-450 found in rats after repeated exposure to vinyl chloride (Table 1) is con-

FIG. 2. (a) Portion of a hepatocyte from control. Stacks of rough endoplasmic reticulum (RER) are separated by many ovoid mitochondria (M). Chromatin is finely dispersed in the nucleus (N). 9300X. (b) Hepatocyte after 2 weeks of vinyl chloride exposure. Dilatation of RER appeared widespread in these two cells. Bile (B) canaliculus appeared unaltered in these rats. 5300X. (c) Four weeks after vinyl chloride exposure. Golgi complex (G) appeared unaffected while cisternal dilation continued. Distinction between SER and RER is complicated by the detachment of ribosomes. Lipid droplets (L) and glycogen (GL) often accumulated. 9500X. (d) A fat-storing interstitial cell is surrounded by several hepatocytes in a vinyl chloride-treated animal. Unlike lipid stored in hepatocytes, the shape of lipids (L) appeared irregular in these cells. 8000X.

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sistent with work by Reynolds *et al.* (1975). This decrease in cytochromes P-450 content was also shown *in vitro* (Guengerich and Strickland, 1977; Ivanetich *et al.*, 1977) suggesting that a metabolite of vinyl chloride destroys the cytochrome. Mixed-function oxidase activity, with benzphetamine as substrate was unaltered.

With regard to the structural alterations produced by vinyl chloride, the present findings confirmed previous observations on the selective effect of this carcinogen in the endoplasmic reticulum (Du *et al.*, 1979). A gradual increase in the number of hepatocytes affected and the involvement of both smooth and rough ER were shown in this study. The endoplasmic reticulum is the primary site of protein synthesis and its dilation suggested the presence of a vinyl chloride-related effect. The lack of a concomitant Golgi hypertrophy is noteworthy since this organelle serves as the site of glycosylation and packaging of many exportable proteins. It may be inferred that the vinyl chloride effect is mainly an intracellular phenomenon. The relatively late involvement of SER could reflect a further attempt at detoxification by the hepatocyte. The tendency for the interstitial cells to accumulate lipid and the heightened phagocytic activity correlate with increases in collagen formation and may be evidence of low-grade cellular injury. This may prove to be the initial histological response to vinyl chloride exposure.

Ultrastructural responses to chronic inhalation of vinyl chloride were reported by Feron *et al.* (1979). Unlike our findings, the principal effects observed were swollen mitochondria and some proliferation of SER. The mitochondrial swelling presumably reflects the extensive vacuolization observed by light microscopy. The discrepancy probably resulted from differences in the dose and duration in vinyl chloride exposure since Feron *et al.* (1979) exposed rats to 5000 ppm vinyl chloride for 52 weeks. Such an extensive mitochondrial lesion could result in biochemical modifications such as a change in

ATPase or cytochrome oxidase levels. The only enzyme activity measured, glucose-6-phosphatase, was reduced.

The authors believed that the altered glutathione metabolism, as reflected by the increased nonprotein sulfhydryl content, and the increased activities of glutathione-S-transferases and glutathione reductase, in rat liver after repeated exposure to high doses of vinyl chloride represent an early hepatocellular adaptation to vinyl chloride exposure.

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Enzymatic Reticuloendothelial

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One of the effects (insecticide) is its effect on macrophage phagocytic activity (Malinin, 1973; Olefirowicz, 1973). It provokes a dose-related decrease in the clearance of colloidal carbon from the stream of the rat by the reticuloendothelial system (RES) (Piperno, 1971). On the basis of these observations could be the basis that carbaryl is an inhibitor of the activity of enzymes of the serine protease system (methyl carbamylation of the serine at the active site) (Piperno, 1971). In fact the hepatic cell-bound serine esterases are indispensable to the function of phagocytic cells.