

bcc: R. E. Keller - QUEENY.

February 27, 1967

Dr. W. J. Thomas
Research Division
Building No. 33
National Cash Register
Dayton 9, Ohio

Dear Dr. Thomas:

Attached is a photostat of the original paper of
Dr. Jensen in Sweden, relating to polychlorinated
biphenyls. I will be happy to have your ideas after
you read it.

As far as the section on toxicology is concerned, it
is true that chloracne and liver trouble can result
from liver damage. Whether or not this is at all relevant
to small quantities existing in human fat is, of course,
an entirely different question.

At any rate, I believe before we worry about the
toxicological part of the problem, we should settle
the analytical part.

Sincerely,

R. Emmet Kelly, M. D.
Medical Director

ENK/ln
att.

0431149

DEFENDANT'S
EXHIBIT

#9

PHGNCR 2000087

MONSFOX00000088

Mr Chairman, Ladies and gentlemen.

In honor to our British ^{guest} I will try to hold this lecture in English.

As the title of this lecture states, I am today going to tell about the discovery of some hitherto unobserved chlorinated hydrocarbons having up to eight chlorine in the molecule and found in residues and lyside. The chemical name of polychlorinated biphenyls (in the following called PCB). To get familiar with PCB I will start with the chemistry and toxicology.

Chemistry

1st

The main characteristic of PCB is 1. Their very high stability. As an example they can be boiled with nitric acid without being destroyed. 2. They are hardly metabolised in living organism. 3. If more than 4 chlorine are present they are non inflammable. It is clear that these three characteristics does it easy to understand that when they have entered the living organism they will have a low persistence. But it is difficult to explain how they find their way into the living organism. One thing seems to be clear, they don't come from agricultural use, but from a technical one and most probable it comes to the nature via wastes that are tried to be burnt up, because then we have them at once in the air, because of their non inflammability.

Toxicology

The PCB were introduced in 1929 and as early as 1936 Jones and Alden reported that 23 out of 24 men employed in manufacturing of PCB suffered from an acne form eruption of the skin. Acne did not appear until 6 to 8 months after the material was first used. In 1937 Brinkner reported that rats exposed to chlorinated biphenyls in concentration of approximately 1 mg/m^3 for 16 hours a day for 6 weeks showed damage of the liver. After that time the allowed concentration of PCB in air is 0.5 mg/m^3 . (For DP; the same value is $0.5 - 1 \text{ mg/m}^3$). The same authors finished their experiments in 1938, and related that these compounds have an injurious effect, manifested solely in the liver. Chlorinated biphenyls appeared to be the most injurious chlorinated compounds of all tested.

PHGNCR 2000083

MONSFOX00000089

Greenburg, Mayer and Smith 1959 reported that PCB and polychlorinated naphthalenes are blamed for the death of three young workers, and that pregnant women and persons who have at any time had any liver diseases are particularly susceptible.

Vedel, Baller and Denton gave 1942 animals PCB involving administration by inhalation, ingestion and skin absorption. Histological examination of the viscera showed important toxic effect only in the skin and liver, and the degeneration effects in the liver are essentially the same whatever was the method for the administration. Faribak (1955) found as an occupational poison in the electrical industry, mixed tetra and penta chlorobiphenyl causes folliculitis, eczema, pyoderma and other skin affections, and that its principal toxic effect is fatty degeneration of the liver.

Miller (1944) injected 69 mg PCB (4 and 5 chlorine) subcutaneously in 32 guinea pigs. Eight to ten days after injection, fat droplets were noted in the liver cells, and after 16 days they were present in moderate or very large numbers. Rabbits and rats were also tested in this investigation, as well as the PCB was administered both continuously, subcutaneously or ingested in the food. In the feeding experiment 8 guinea pigs received 2.8 ppm of 69 mg of the chlorinated biphenyl 1 week apart. Death occurred in 11 to 29 days.

Finally McLaughlin 1964 reported a method to test the chemical toxicity and teratogenic effect by injection into the yolk sac of fertile eggs prior to incubation. PCB was found between the eight compounds among 100 tested having the highest order of toxicity. No hatch was found at a level of 25 mg per egg. At a level of 50 mg per egg, one chick hatched out of 20 injected eggs, but died 2 days later. Some embryos which were examined after they died, showed weak deformities (often a short upper trunk) and growth retardation. Lead acetate resulted as an example in no hatch at a level of 1 mg per egg. Autopsy of the dead embryo have showed extensive brain damage. Mercuric chloride showed no hatch even at a level of 0.5 mg per egg.

As the analytical chemistry is a pronounced service science I have been in contact with many scientists from other fields during the work with residue analysis, and I have always found this contact very stimulating for my own work. This co-operation often demands that we are talking the same scientific language. Because of this need I will today try to give a lecture in low level analytical chemistry for biologists, illustrated by the residue analysis of polychlorinated biphenyls.

The lecture will be divided in the following three sub-divisions:

PHGNCR 2000089

MONSFOX00000090

1. Chemistry of PCB and their toxicology.
2. Analytical methods for residue analysis and proof of structures.
3. Behaviour of PCB in nature, differences in metabolizing rate of the PCB components, potentiation in an ecological serie, concentration levels, and examples of samples which have been proved to contain PCB.

A residue analysis can be divided in:

1. Extraction of the pesticides from the biological material, followed by a careful cleaning-up to take away interfering substances, most often fats.
2. Identification analysis by mean of gas chromatography. Thin-layer chromatography and mass spectrometry.
3. Quantitative analysis.

At an ecological laboratory in Riksmuseet in Stockholm 1-2 g of a sample is cut out of the biological material and transferred into a weighed and carefully cleaned test tube, and stored at -20° until analysis. Smaller samples have been used, min. 5 mg of body fat, and with dry materials such as hair, feathers, pine needles 100 mg are sufficient to reach the desired 10 ng/g level in residue analysis. In cases of water proofs 1 l. is used for reaching the 10 pg/g. level.

B.1(homog) In order to facilitate complete extraction of the fatty materials from the biological sample, the double amount of finely powdered anhydrous magnesium sulphate is added to the sampling tube, and the whole is homogenised with an insertable homogeniser. The resulting powder is transferred into a special Soxhlet extractor. After 4 hours of extraction the solvent is evaporated, leaving the fat in a small weighed test tube at the bottom of the extractor. This fat is dissolved in methylene chloride in such a way that 100 µl (0,1 ml) contain 20 mg of fat.

Sox.-tube)

The 100 µl solution is now transferred to a little object glass, 5 x 7 cm, covered with a silicagel layer 1 mm thick, in order to form a line 0,7 cm from one end of the slide. Inserting this thin-layer plate into a vessel the bottom of which is covered by a few mm of methylene chloride, the solvent will be sucked up in the dry layer of silicagel, and at least reach the upper end of the plate. The fact is that the fat has a greater affinity to the powder on the plate than the chlorinated hydrocarbons have, - and we get a separation. The fat being more polar than the chlorinated hydrocarbons will never go longer than 2 cm before the

0281422

PHGNCR 2000090

MONSFOX00000091

5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65
66
67
68
69
70
71
72
73
74
75
76
77
78
79
80
81
82
83
84
85
86
87
88
89
90
91
92
93
94
95
96
97
98
99
100
101
102
103
104
105
106
107
108
109
110
111
112
113
114
115
116
117
118
119
120
121
122
123
124
125
126
127
128
129
130
131
132
133
134
135
136
137
138
139
140
141
142
143
144
145
146
147
148
149
150
151
152
153
154
155
156
157
158
159
160
161
162
163
164
165
166
167
168
169
170
171
172
173
174
175
176
177
178
179
180
181
182
183
184
185
186
187
188
189
190
191
192
193
194
195
196
197
198
199
200
201
202
203
204
205
206
207
208
209
210
211
212
213
214
215
216
217
218
219
220
221
222
223
224
225
226
227
228
229
230
231
232
233
234
235
236
237
238
239
240
241
242
243
244
245
246
247
248
249
250
251
252
253
254
255
256
257
258
259
260
261
262
263
264
265
266
267
268
269
270
271
272
273
274
275
276
277
278
279
280
281
282
283
284
285
286
287
288
289
290
291
292
293
294
295
296
297
298
299
300
301
302
303
304
305
306
307
308
309
310
311
312
313
314
315
316
317
318
319
320
321
322
323
324
325
326
327
328
329
330
331
332
333
334
335
336
337
338
339
340
341
342
343
344
345
346
347
348
349
350
351
352
353
354
355
356
357
358
359
360
361
362
363
364
365
366
367
368
369
370
371
372
373
374
375
376
377
378
379
380
381
382
383
384
385
386
387
388
389
390
391
392
393
394
395
396
397
398
399
400
401
402
403
404
405
406
407
408
409
410
411
412
413
414
415
416
417
418
419
420
421
422
423
424
425
426
427
428
429
430
431
432
433
434
435
436
437
438
439
440
441
442
443
444
445
446
447
448
449
450
451
452
453
454
455
456
457
458
459
460
461
462
463
464
465
466
467
468
469
470
471
472
473
474
475
476
477
478
479
480
481
482
483
484
485
486
487
488
489
490
491
492
493
494
495
496
497
498
499
500
501
502
503
504
505
506
507
508
509
510
511
512
513
514
515
516
517
518
519
520
521
522
523
524
525
526
527
528
529
530
531
532
533
534
535
536
537
538
539
540
541
542
543
544
545
546
547
548
549
550
551
552
553
554
555
556
557
558
559
560
561
562
563
564
565
566
567
568
569
570
571
572
573
574
575
576
577
578
579
580
581
582
583
584
585
586
587
588
589
590
591
592
593
594
595
596
597
598
599
600
601
602
603
604
605
606
607
608
609
610
611
612
613
614
615
616
617
618
619
620
621
622
623
624
625
626
627
628
629
630
631
632
633
634
635
636
637
638
639
640
641
642
643
644
645
646
647
648
649
650
651
652
653
654
655
656
657
658
659
660
661
662
663
664
665
666
667
668
669
670
671
672
673
674
675
676
677
678
679
680
681
682
683
684
685
686
687
688
689
690
691
692
693
694
695
696
697
698
699
700
701
702
703
704
705
706
707
708
709
710
711
712
713
714
715
716
717
718
719
720
721
722
723
724
725
726
727
728
729
730
731
732
733
734
735
736
737
738
739
740
741
742
743
744
745
746
747
748
749
750
751
752
753
754
755
756
757
758
759
760
761
762
763
764
765
766
767
768
769
770
771
772
773
774
775
776
777
778
779
780
781
782
783
784
785
786
787
788
789
790
791
792
793
794
795
796
797
798
799
800
801
802
803
804
805
806
807
808
809
810
811
812
813
814
815
816
817
818
819
820
821
822
823
824
825
826
827
828
829
830
831
832
833
834
835
836
837
838
839
840
841
842
843
844
845
846
847
848
849
850
851
852
853
854
855
856
857
858
859
860
861
862
863
864
865
866
867
868
869
870
871
872
873
874
875
876
877
878
879
880
881
882
883
884
885
886
887
888
889
890
891
892
893
894
895
896
897
898
899
900
901
902
903
904
905
906
907
908
909
910
911
912
913
914
915
916
917
918
919
920
921
922
923
924
925
926
927
928
929
930
931
932
933
934
935
936
937
938
939
940
941
942
943
944
945
946
947
948
949
950
951
952
953
954
955
956
957
958
959
960
961
962
963
964
965
966
967
968
969
970
971
972
973
974
975
976
977
978
979
980
981
982
983
984
985
986
987
988
989
990
991
992
993
994
995
996
997
998
999
1000

solvent reaches the upper part of the glass.

The front of the fat appears quite visible against a lamp, and with the aid of a razor blade the zone above the fat is transferred to the elution tube and the chlorinated dioxides absorbed on the powder can now be eluted by one ml of ether. The concentration is sufficient for detection of the chlorinated hydrocarbons down to the 10^{-12} g level.

The most step in the analytical procedure concerns the separation of the different chlorinated hydrocarbons that the sample may contain. As a matter of fact, this is a troublesome task. It is easy to estimate what is not present, but more difficult to say exactly one is present. We suffer from the negative demonstration, as will be shown later. At first a few words about the separation of the components present in the sample and their visualization.

The separation is accomplished by means of a gas chromatograph fitted to a detector that transfers its impulses to a recorder.

The system is shortly described:

A spirally formed glass tube with an inner diameter of 2 mm and about 2 m in length is filled up by a support, covered with an thin layer of an oil. The tube is heated in the chromatograph to about 200° . Through the tube a stream of nitrogen continuously follows. When about 10 μ l ($1/100$ of 1 ml) of the purified sample is injected into the tube, the components of the sample will be evaporated and go forward through the column with the gas stream. As the constituents have different affinity to the column filling they will pass the column with different speed and it will take different time for them to reach the detector at the other end of the glass tube. If the temperature and the nitrogen flow are held constant this time, the retention time, has a specific value for a certain compound. This is true, but unfortunately it is also a fact that two components can have the same retention time. This is one of the bigger problems in gas chromatographic analysis of unknown samples, as will soon be obvious.

To make it possible to estimate the retention time it is necessary to visualise the chlorinated hydrocarbons. For that purpose more or less specific detectors are used. The detector most often used in pesticide analysis is the so called electron capture detector, which can detect down to one picogram ($= 10^{-12}$ g of lindan). Unfortunately this detector is not specific for chlorine, but gives answer also for oxygencontaining compounds. The response here is much lower but can be counterbalanced if the concentration of the oxygen containing is much higher.

0291423

The principle for the electron capture detector is shortly:

At the end of the gas chromatographic tube is placed a little tube containing a foil made of titanium trichloride. This is α -radiant. The α -

PHGNCR 2000091

MONSFOX00000092

particles are reacting with the nitrogen molecules coming from the column. Then we get $+ N_2 \rightarrow e^- + N_2^+$. Over the detector we have a tension of 90 volt and by mean of the electrons we will get a constant electrical current over the detector. This standing current is transferred to a one-mV recorder as a constant baseline. When now a chlorinated hydrocarbon leaves the column this compound has a high affinity to the electrons and this means that the amount of electrons will diminish, and they will diminish proportionally to the amount of chlorine. The electrical current will also diminish and this is noted as a peak on the recorder. The area of the peak will be proportional to the amount of substance in the sample. By mean of a standard injection it is now possible to compare the retention time and the area of an unknown component with the retention time and area of the known standard. As said before this detector is not specific for chlorine but anyhow very useful, because of its high sensitivity. The system described has, as we have seen, two disadvantages:

1. Two different compounds can have the same retention time and be detected as one peak.
2. A registered peak does not need to be chlorinated, because the detector is not specific.

If the sample is injected in two different columns with different chemical properties we have increased the chance for a good separation. If two compounds have the same retention time on one column they may not have it on another. When a result seems doubtful, - if the compound being responsible for a certain peak contains chlorine or not - it is possible to concentrate the sample and analyse it on a less sensitive detector such as the microcoulometric one, which is specific for chlorine. The compound is burned in a furnace and the generated chlorine titrated directly.

As is seen from the two last mentioned possibilities it is anyhow possible to get a rather high degree of certainty in residue analysis, but it is a rather time-consuming work. When using this method just described, we very often found that many chromatograms from residue analysis of most carefully purified samples still contain a large number of peaks. Many of these have retention times that do not agree with any known chlorinated pesticides, or their metabolites. This chromatogram can serve as an example. It was obtained by residue analysis of a sea-eagle found dead in the archipelago of Stockholm. In the range of the known peaks, there are so many unidentified that there also must be an obvious risk of the known peaks to be covered by unknown ones.

If this remark is found true, the reported results of many previous analyses

0281424

PHGNCR 2000092

MONSFOX00000093

titative anal. As must be brought into question. In the present investigation it is shown that most of the unknown peak of chromatograms at residue analysis of chlorinated pesticides are due to polychlorinated biphenyls.

I will show a chromatogram of human fat analysed on a so called SF 96 column, the most often used type in pesticide analyses. Early retention times were in agreement with DDE, DDT and DDEpp. Next slide shows the same sample analysed on a QP-1 column. Now the former 2 DDE peaks have divided into 4 peaks, and two of them are still in agreement with DDEpp and op., the two new were unknown.

Logically, these unknown components were at first thought to be metabolites of the insecticides. Against that spoke that neither treatment nor concentrated sulfuric acid in ether, this treatment made it rather sure that the compounds did not contain oxygen. In Sweden residues of organic mercury have been investigated rather intensively in the Swedish fauna. As these compounds give very high responses to the electron capture detector it was also investigated if the unknown peaks could have a mercuric origin.

It was found that the water-ecological series had high residues of both mercury (Westermarck, Johnels) and the unknown ones, when the same individuals were analysed. Anyhow, the pheasant suffering most from mercury poisoning only contained low levels of electron capturing compounds and these belonged to the normal insecticides. Therefore the unknown could hardly be mercurials or metabolites of them.

As the sample giving the chromatogram shown in fig. 10, could be estimated to contain DDE and DDE up to 17 g/kg in extractable fat, the amount of unknown compounds also were suggested to be in the same range, and then sufficiently high to do a run on the combined gas chromatograph-mass spectrometer. If this could be done successfully it would be possible to get very important information about the chemical nature of the unknown, for ex. the molecular weight numbers of chlorine etc. This method is up to now the method giving the highest degree of certainty in the low level analytical chemistry, amounts of 100 ng substance being enough.

As this method for identification of totally unknown residues surely will be very important in the future (when i.e. a biologist has found that fishes in a river die) it may be possible by means of this method to find out exactly what compounds are responsible for the death.

For this reason, I will go into some details with this method.

In the actual case we took the extract from 20 mg eagle and concentrated

0251425

PHGNCR 2000093

MONSFOX00000094

it as much as possible and made an injection on the gas chromatograph combined with the mass spectrometer. The result was the chromatogram shown on the next slide. Every time the recorder showed that a compound is leaving the column, the effluent is led to the mass spectrometer. Now just a few words about the mass spec.

The molecules leaving the column are bombarded with electrons at X. We have now got the molecule positive charged, but with the same mass as before. This M^+ is accelerated in a vacuum and will then get a kinetic energy.

where is the speed. Next comes the magnetic field that tries to bend the direction of the molecule. This will be big for a small molecule and less for

If we have a sieve in the other end we can directly read the molecular weight. Added to this parent molecule M^+ we will also get addition informations, because of the fact that M^+ may not be stable, a part of them will be broken down before they reach the sieve in the other end.

For ex. M^+_{DDT} $M^+_{DDT} - Cl_{35}$

Mass spectrograms from the different unknown peaks in the eagle sample are shown. The mass numbers equal to the molecular weights of the unknowns could be read to 426, 392, 358, 324. Astonishingly, the molecular differences were constantly 34 mass units. This difference shows a familiarity in origin of the unknown. Now the fact is that chlorine exists as a mixture of two isotopes with atom weights 35 and 37 in proportion 75:25. If the molecule has one chlorine, this will give two molecule peaks, one for Cl_{35} and one for Cl_{37} . If there are two chlorines we have the possibility of one with only Cl_{35} , one with both Cl_{35} and 37 and one with 2 Cl_{37} and therefore

0281426

PHGNCR 2000094

MONSFOX00000095

The relation of the peaks found on the different mass spec were:

Molecular weight	324	358	392	426
Chlorine content	5	6	7	8

An explanation of the familiarity of the compounds can be given if one substance is built from the former by substituting a hydrogen with chlorine

$M^+ \rightarrow Cl_2$ $RO1 \rightarrow HCl$
 $M^+ \rightarrow 34$

Then it is possible to calculate the molecular weight of the parent hydrocarbon PCB.

$M_{PCB} = M - x \cdot M_{Cl} + x \cdot M_H$, where M is the molecular weight of the component having x chlorine atoms. For ex. for $m = 426$ and $8 Cl$ we will get $M_{PCB} = 426 - 280 + 8 = 156$ and equal with the other molecule.

The most probable formula with carbon and hydrogen giving this molecular weight is $C_{12}H_{10}$ and this can only be satisfied when the parent-hydrocarbon is biphenyl, and the unknown being polychlorinated biphenyl.

This explanation was later fully verified by injection of a synthetic PCB on the mass spec.

Furthermore extensive gas chromatographic investigations proved that the PCB standard gave peaks with the same retention time as the unknown peaks from the sea eagle.

With the method just described I suppose that we have a new possibility to study the residues in the air because the pine needles can always be . We have had great difficulty in quantizing the PCB,

but when getting a little more time it will be possible. We have done a few calculations on a few species, and I suppose they are right within a factor 2. We have found the residue to be from

It has been my statement here to-day to present this method for studies of defiling of the nature, and with this method a new type of defiling agents has been found to be present in nature, and a few experiment have shown where they may be found.

Now this method is going to be used in the first hand to estimate how the situation is in nature as a whole, and in the other hand to find the leaks through which they find its way to nature. Some maybe are present here today to get news about the leaks, and to then I want to say come back in a year.

0281427

PHGNCR 2000095

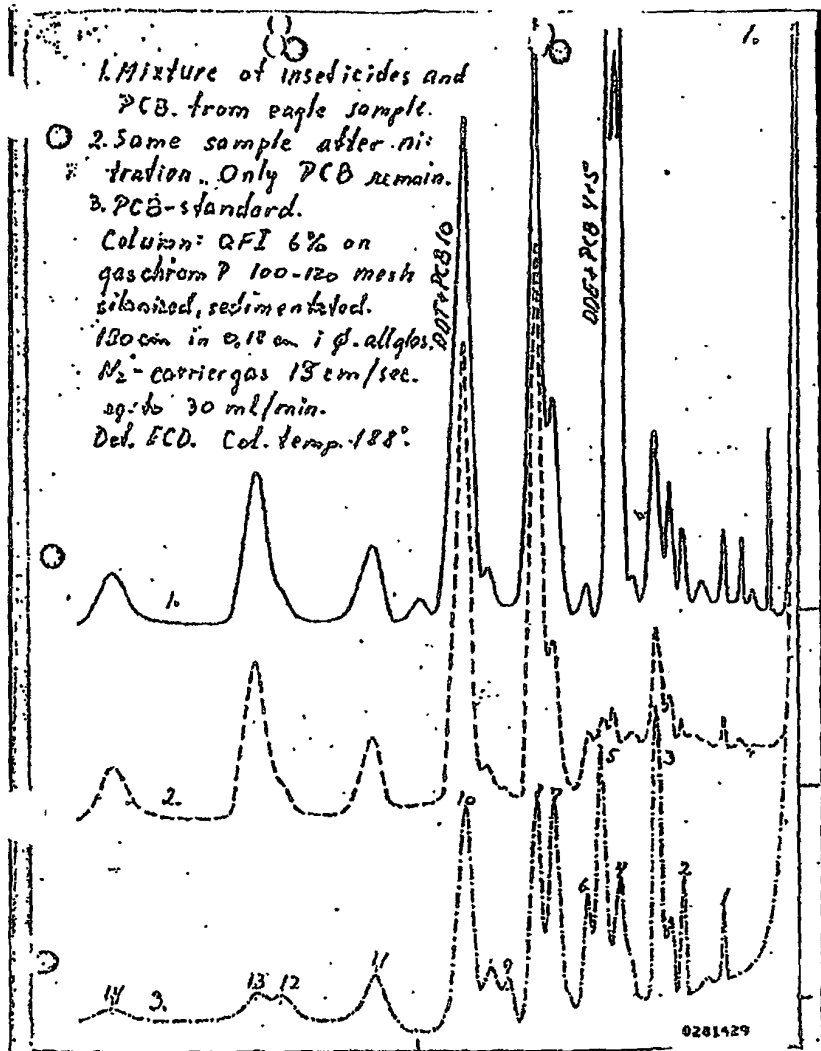
MONSFOX00000096

So much I think I can say again that the PCB hardly can come from agriculture. As support for this suggestion I can say that we have found PCB in eagle feathers from Hkamooet from 1944, where hardly any chlorinated pesticides were used in agriculture. One more thing that I find important to say is that in contrast to the mercury problem this does not seem to be a pure Swedish problem. I have just studied chromatograms taken from London air, and they clearly contain PCB, and Dr. Halden has told me that he also finds them in his fish samples. But finally in waiting at more results I should like to point out something. It is proved that PCB comes to nature, we don't know now where they are used, but they are very persistent to chemicals and to fire. I think the poison jury should try to state that a content of PCB shall always be found in an open declaration.

0281428

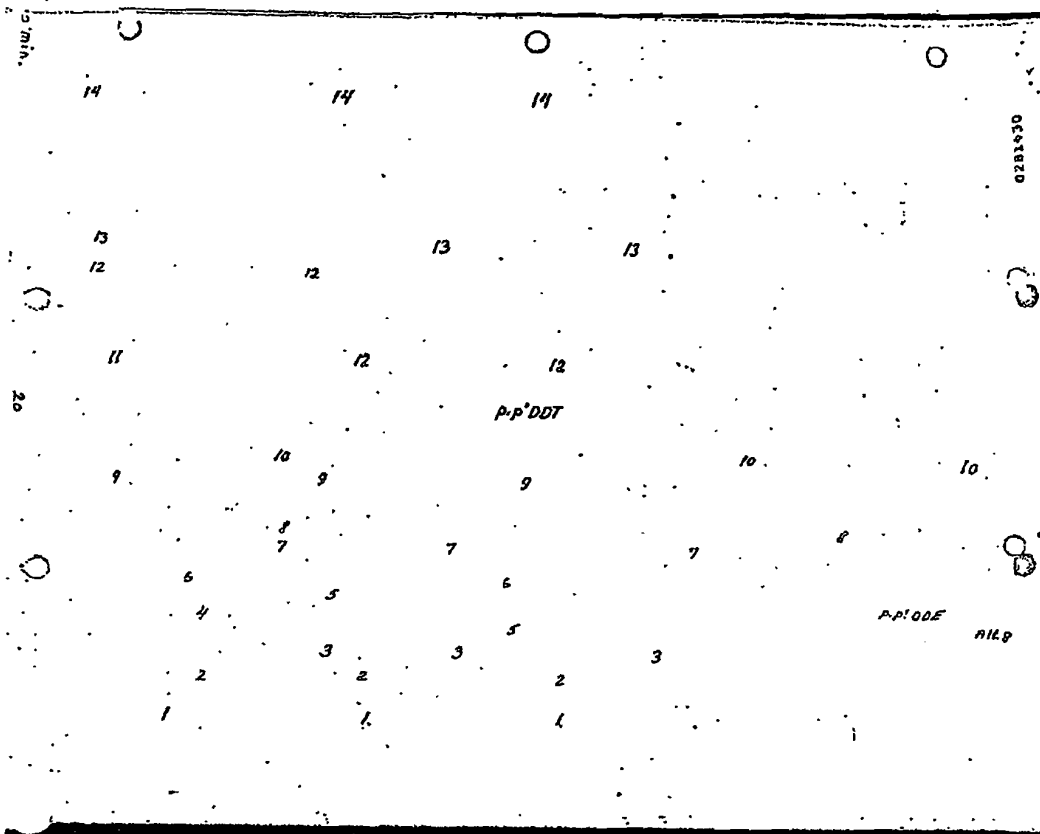
PHGNCR 2000096

MONSFOX00000097



PHGNCR 2000097

MONSFOX00000098



PHGNCR 2000098

MONSFOX00000099

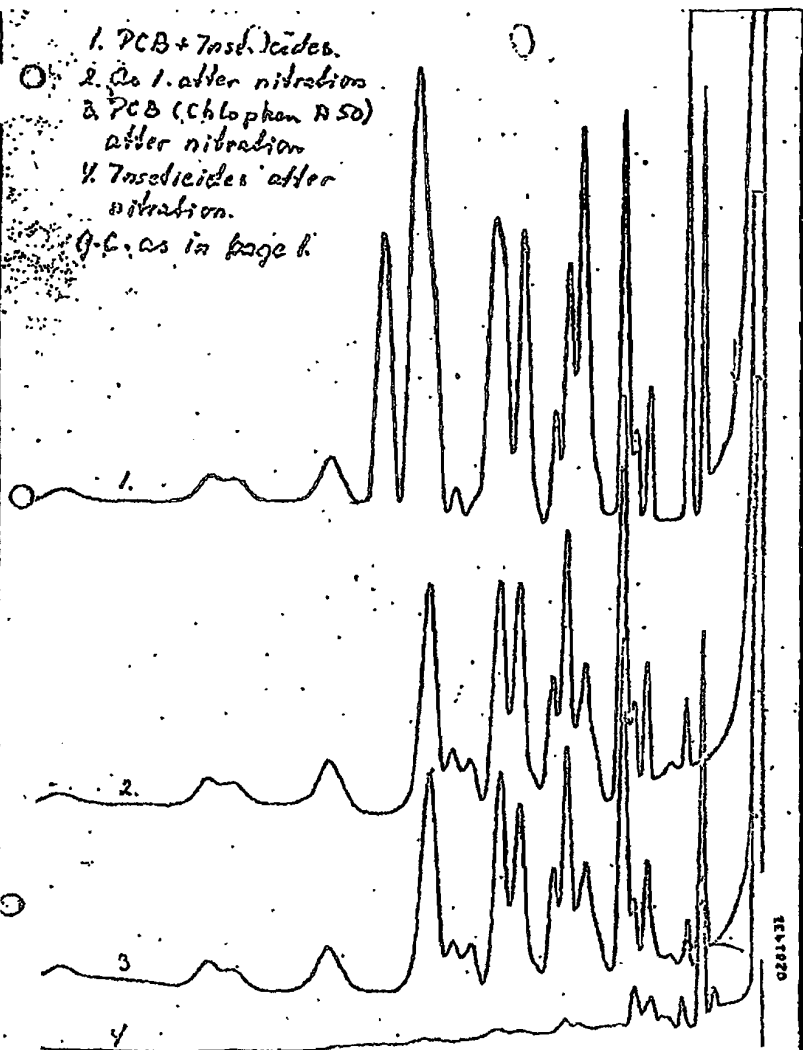
1. PCB + 7nsd.icides.

2. Co 1. after nitration

3. PCB (Chlophan B 50)
after nitration

4. 7nsdicides after
nitration.

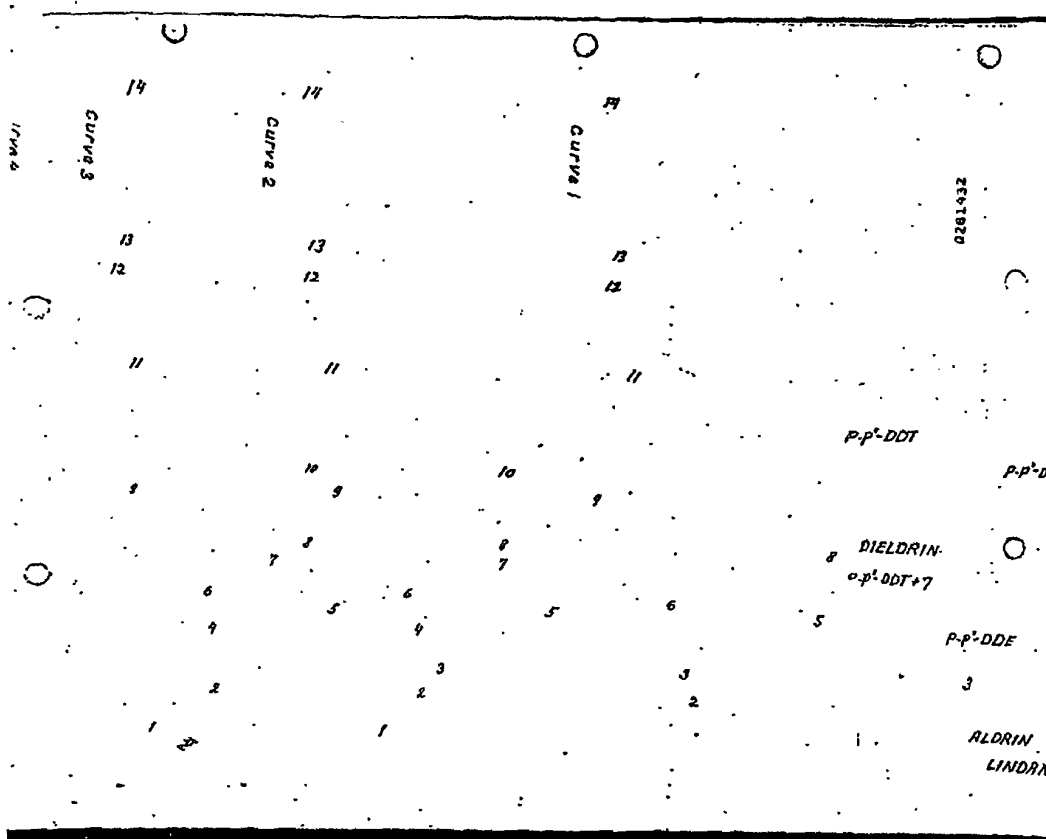
Q-C. as in page 6.



PHGNCR 2000099

MONSFOX00000100

WATER_PCB-SD0000054860



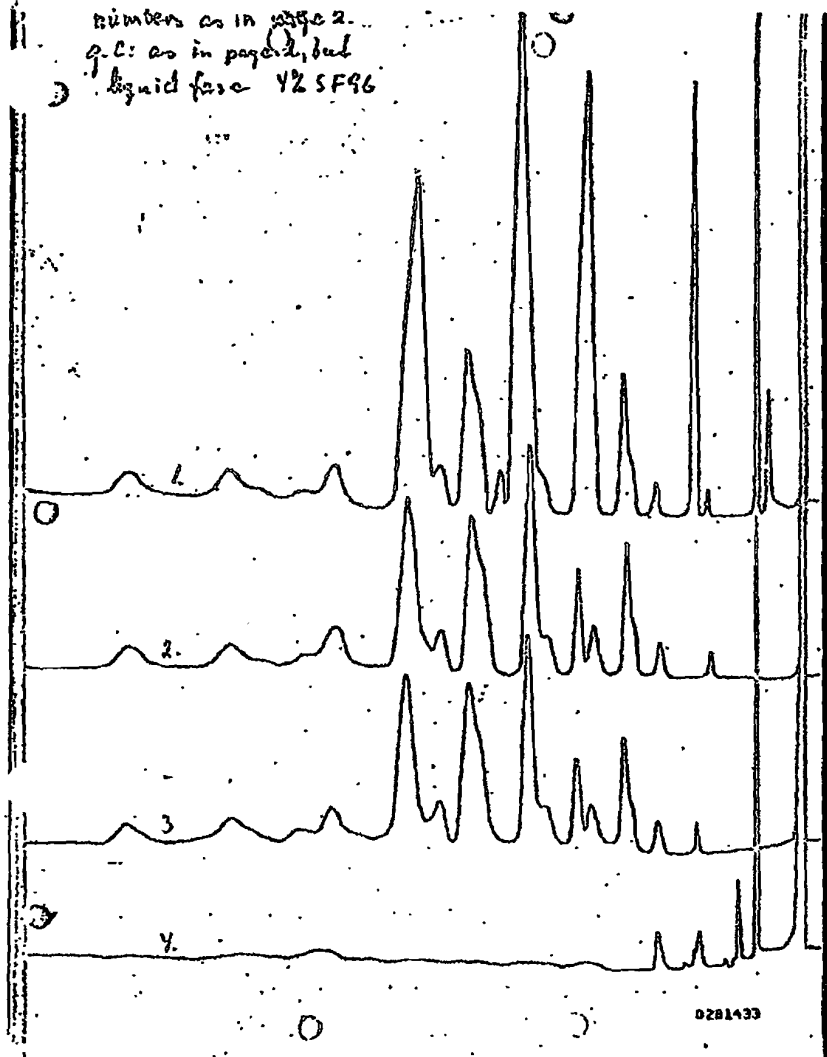
FIGNCR 2008100

MONSFOX00000101

numbers as in page 2.

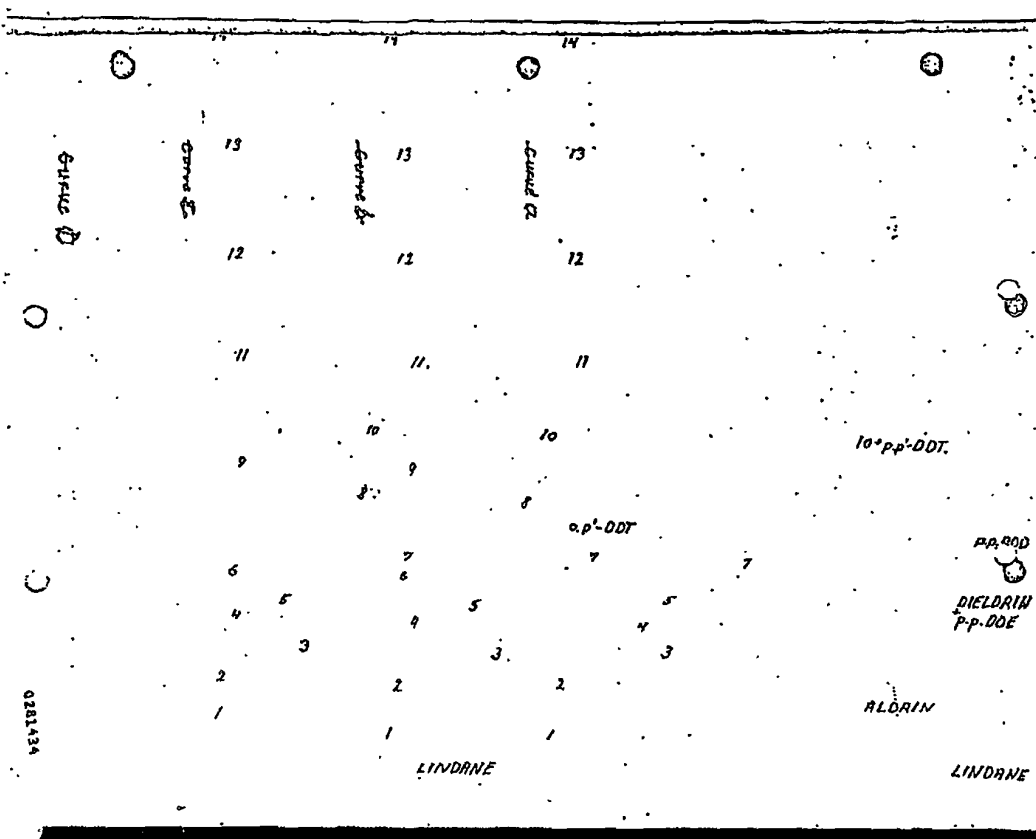
q.c. as in page 2, but

liquid phase YZ SF96



PHGNCR 2000101

MONSFOX00000102



PHGNCR 2000102

MONSFOX00000103

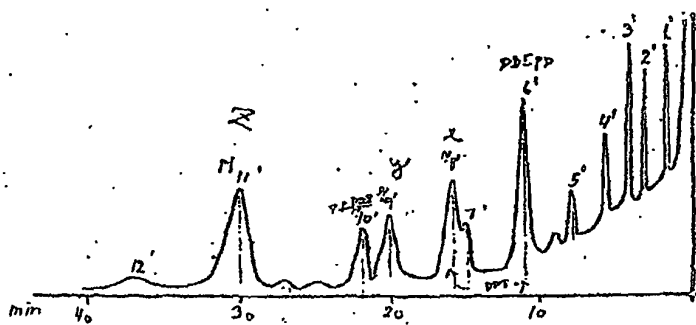


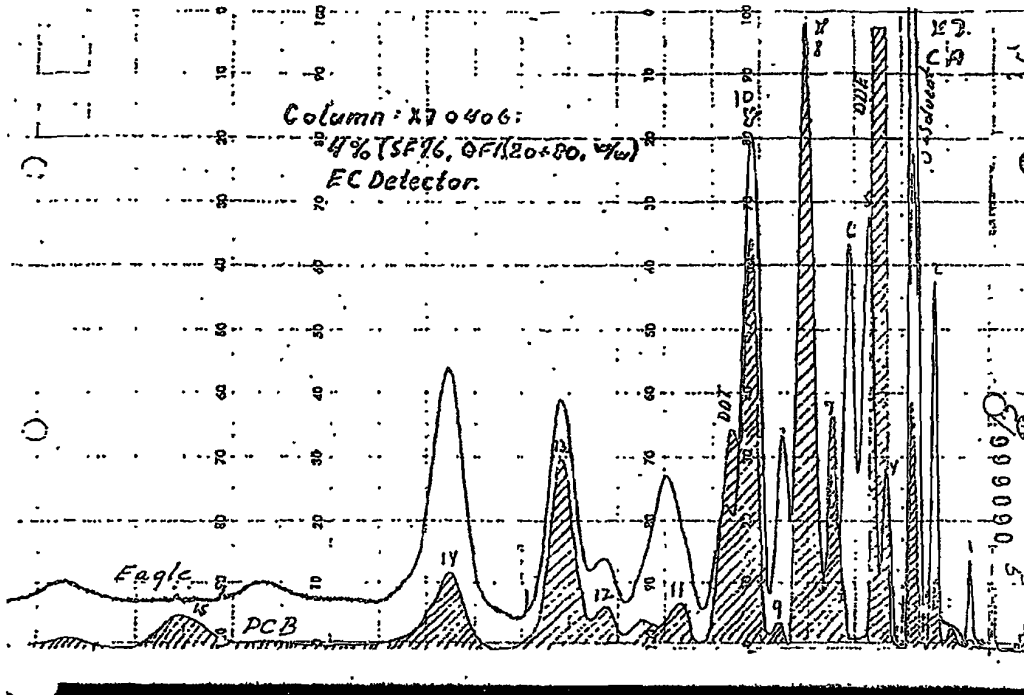
Fig 3. GC-MS chromatogram from eagle sample. Peak numbers represent the points of scanning.

028135

PHGNCR 2000103

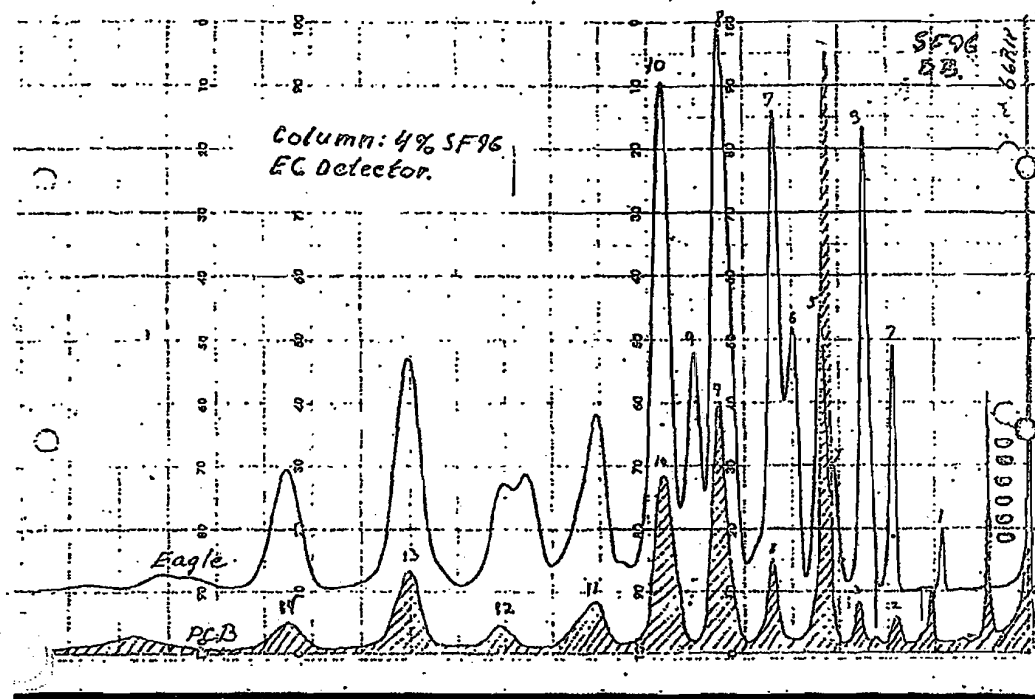
MONSFOX00000104

0281436



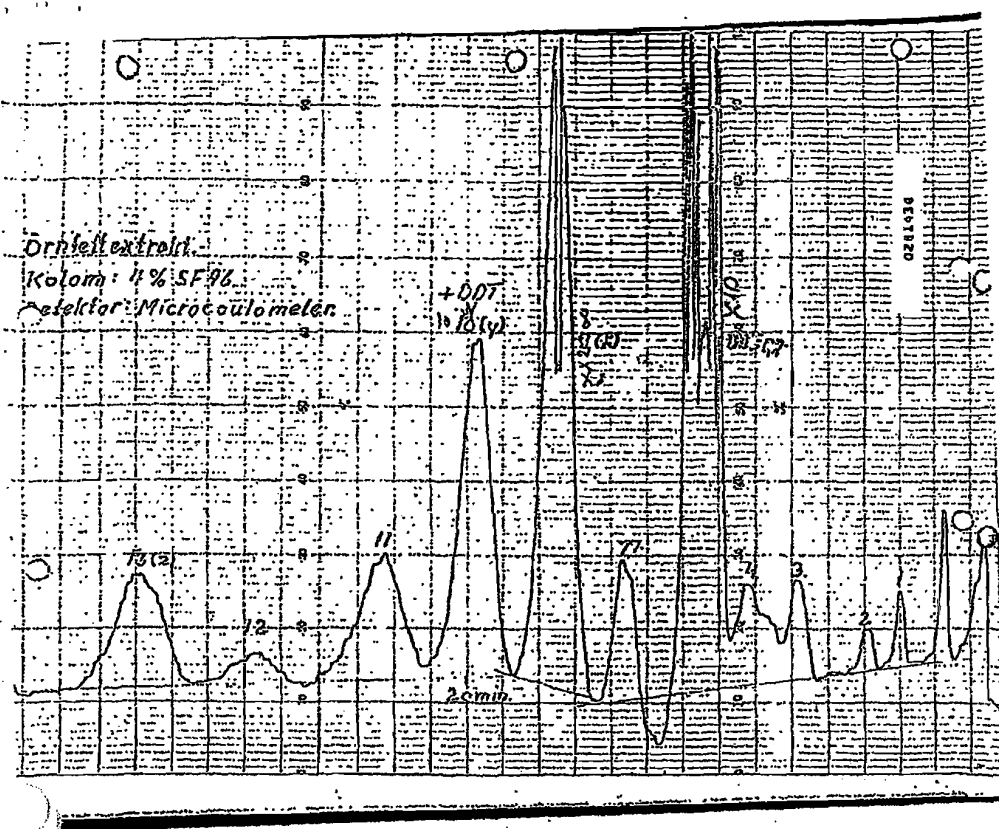
PHGNCR 2000104

MONSFOX00000105

c1ccc(cc1)-c2ccccc2

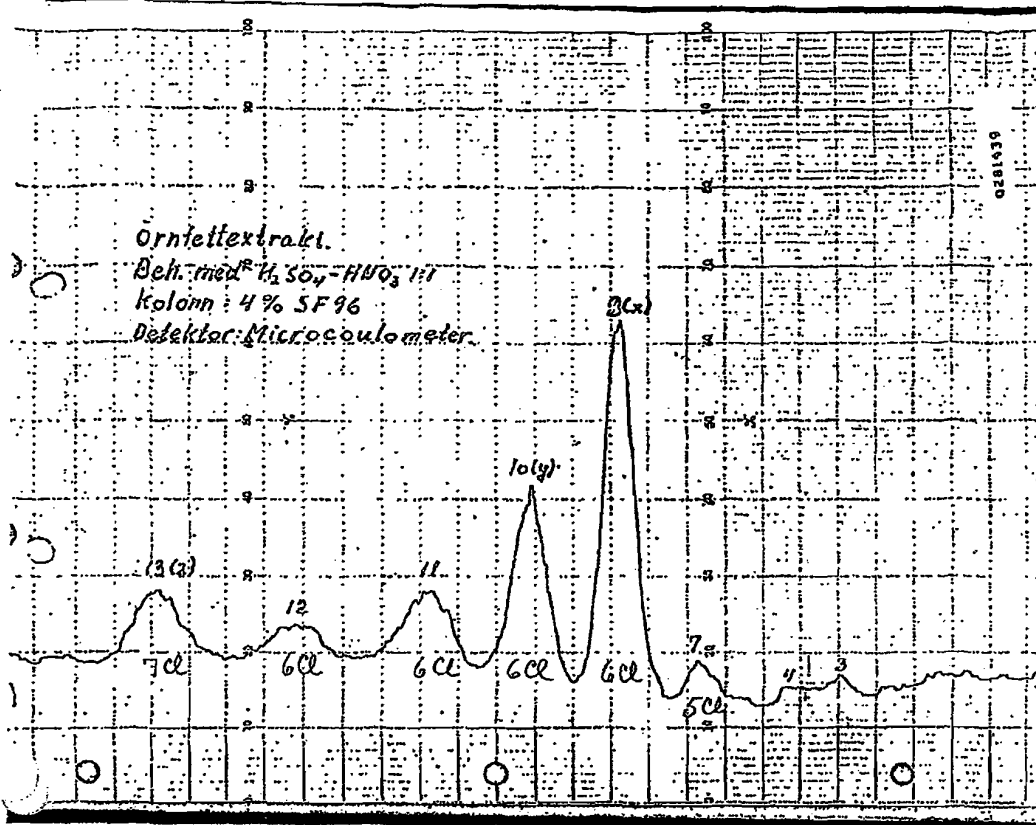
PHGNCR 2000105

MONSFOX00000106



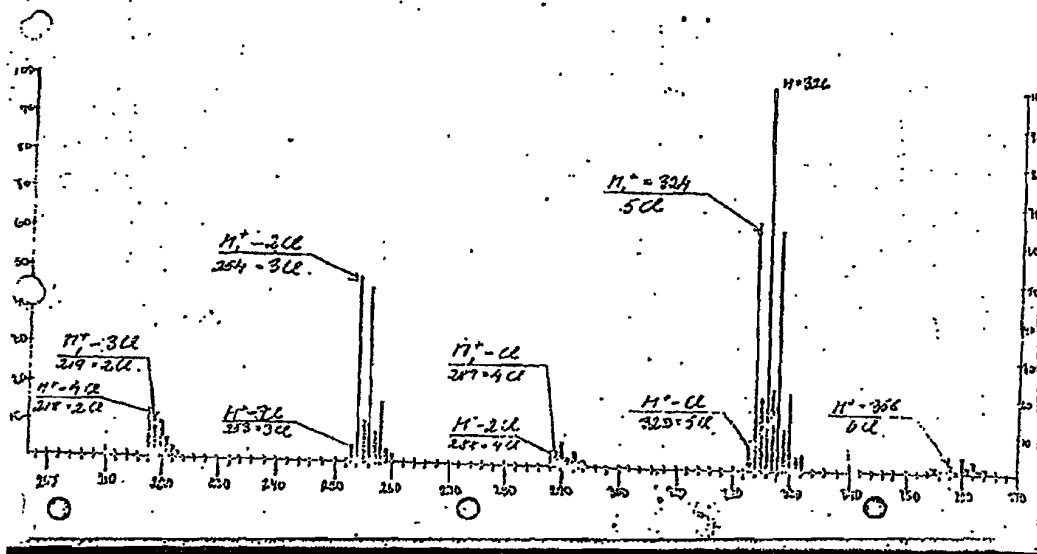
PHGNCR 2000106

MONSFOX00000107



Glophen ASD
 19pp nr. 7.

0281440

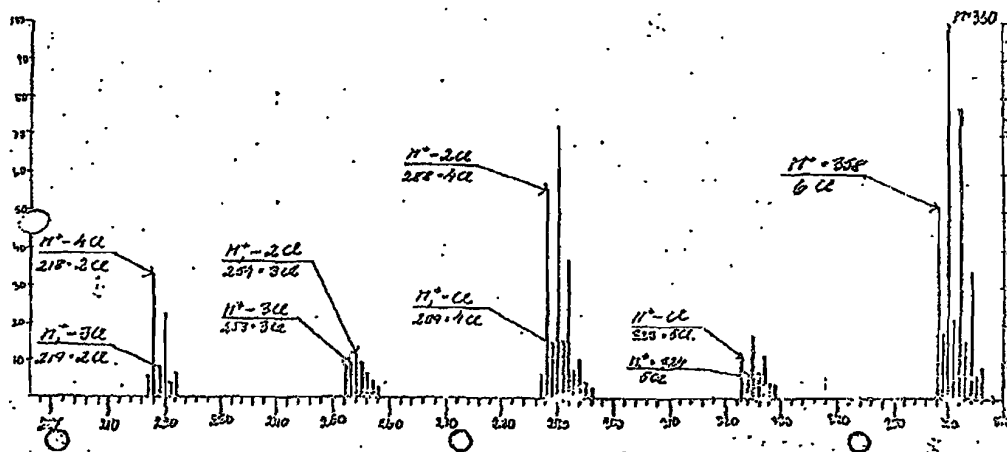


PHGNCR 2000108

MONSFOX00000109

Clophen A50
topp nr 8

0281441

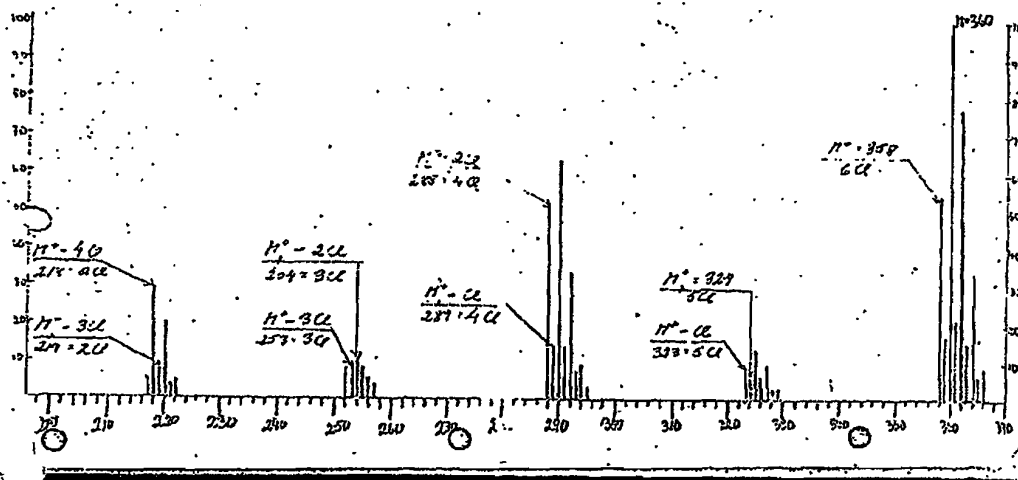


PHGNCR 2000109

MONSFOX00000110

Clapham A50
 10pp. nr 10

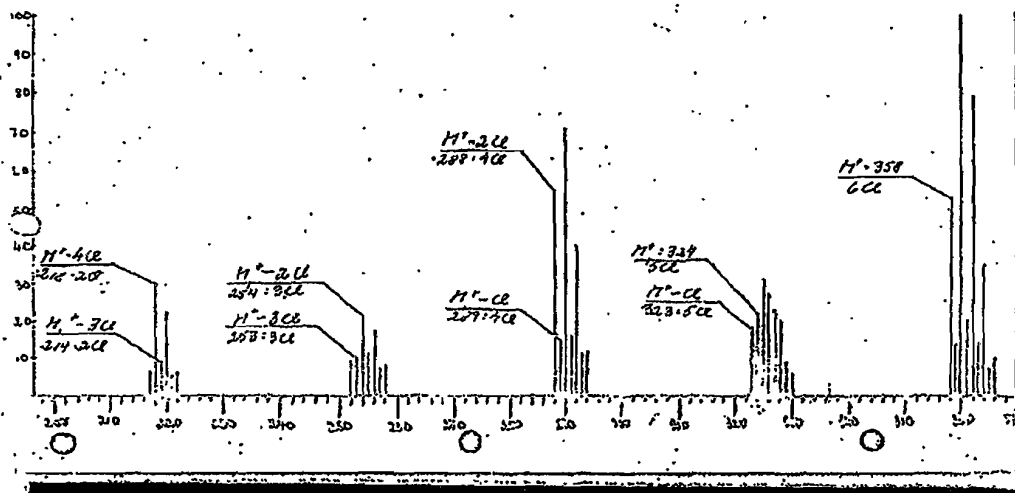
02181920



PHGNCR 2000110

MONSFOX00000111

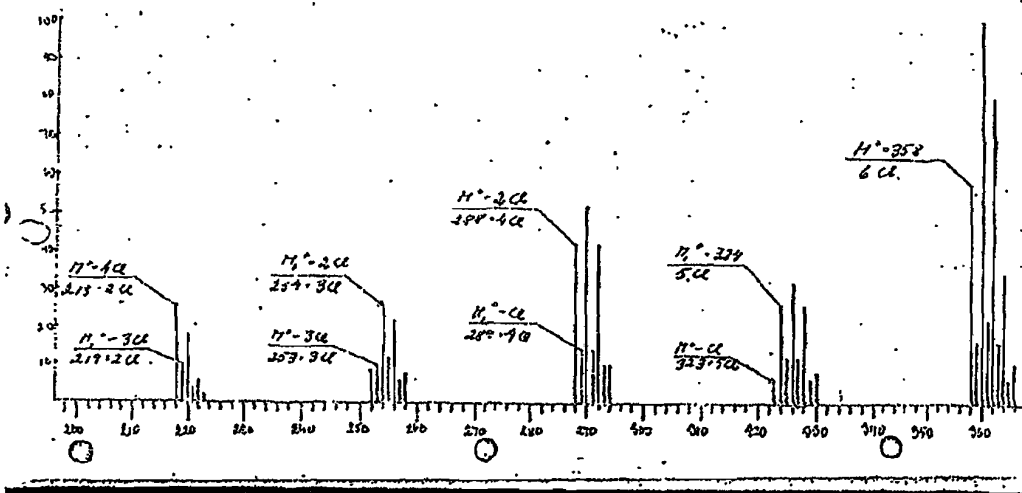
040107



MONSFOX00000112

Clepton ASD
 10pp or 12

0281949



FIGNCR 2000112

MONSFOX00000113