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ASSESSMENT OF THE BIOLOGICAL PERSISTENCE OF POLYCHLORINATED BIPHENYLS

The research I will review today will focus upon one aspect of Monsanto's efforts to understand the environmental impact and behavior of our polychlorinated biphenyl or PCB products.

This research was initiated early in 1969 after development of the necessary PCB analytical methodology and subsequent confirmation of Dr. Soren Jensen's identification of PCB residues in fish and birds in Sweden.

At this point in time, PCB residue data from Monsanto and external environmental monitoring programs indicated that at the previous rate of use and release of these products that some PCB homologs were beginning to reach detectable levels in fish, birds and mammals. Conversely, these data indicated to us that with the exception of localized, controllable contamination, PCB homologs with less than five chlorine atoms per molecule had not accumulated to detectable levels; even though it was known that significantly greater amounts of the PCB homologs with less than 5 chlorine atoms per molecule had been manufactured and used over the years.

Now, before discussing our biological studies, I would like to review for you the gross homolog composition of our Aroclor products and then in a very brief fashion, try and illustrate to you the complexity of these materials and hence the complexity of the problem.

In the first slide (#1), is shown the most recent data on the weight % composition of four of our Aroclor products as a function of each detectable PCB homolog. The first column on the left lists the homolog in question and the subsequent columns under each product show the weight % distribution of each PCB homolog in each product.

As most of you probably know, with the exception of Aroclor 1016, the last two digits of each product number refer to the degree of chlorination. For example, Aroclor 1221 contains 21% chlorine by weight, and so on.

Aroclor 1016 is a special case in that while it contains about 41% chlorine by weight, its penta, hexa, and heptachloro biphenyl content has been significantly reduced with respect to Aroclor 1242, a product produced by direct chlorination, containing 42% by weight chlorine. Please note that the penta, hexa, and heptachloro biphenyl homologs in Aroclor 1016 have been reduced by factors of about 8, 10, and 10, respectively, with reference to Aroclor 1242.

As you can also see, the chief constituents of Aroclor 1221 are the mono- and dichloro biphenyls, while Aroclor 1016 and Aroclor 1242 contain predominantly di-, tri-, and tetrachloro biphenyls, and Aroclor 1254, tetra, penta, and hexachloro biphenyls.

As I indicated earlier, this table represents our most recent efforts at determining the homolog distribution of our PCB products and as such a number of you may have seen estimations of the homolog content of these products which are significantly different. The analytical methodology used is currently in a very dynamic state and our understanding of the contents of these products increases as the methodology is improved. At this point in time, we regard these numbers as the most accurate ones currently available.

In the next slide (#2) are shown examples of low resolution - packed column electron capture chromatograms of Aroclor 1221, Aroclor 1242, Aroclor 1254, and Aroclor 1260. This is what these products look like to a residue analyst using the most commonly employed detection system.

From these chromatograms, it can be readily seen that we are dealing with multi-component products, which of course, increases the complexity of assessing every aspect of this problem - relative to a well defined single component system such as DDT.

I should mention at this point, that the PCB residues generally found in wildlife are most similar to Aroclor 1254 and Aroclor 1260 chromatograms.

The next slide (#3) demonstrates that in reality, these materials are even more complex than is generally realized. In the upper right portion of this slide is again shown a low resolution electron capture gas chromatogram of Aroclor 1242 under the optimum conditions normally employed by residue analysts. Under these conditions, Aroclor 1242 would appear to be a 15 component system.

In the lower portion of this slide is a flame ionization gas chromatogram of the same material using a high resolution S.C.O.T. column. If one carefully inspects this chromatogram, our simple 15 component product has now been resolved into 55 different components.

These facts simply indicate that all PCB products cannot be lumped together in terms of either their environmental impact or persistence.

The type of biological studies which we have carried out to date are shown in the next slide (#5). For discussion purposes, they can be conveniently divided into two categories: "Primary Bacterial Degradation Studies", an area in which research on PCBs is just beginning, and "Residue Accumulation Studies". Most of our bacterial degradation work has been centered around the fairly well known semi-continuous activated sludge degradation test.

Our residue accumulation studies have been fairly extensive and have involved the exposure of better than 2100 fish, chickens, rats and dogs to the various Aroclor products; resulting in the collection of over 1200 samples of which approximately 500 pooled samples were eventually analyzed for PCB residues.

The prime objective of these studies is given on Slide #6.

Slide #7. The semi-continuous activated sludge test procedure we used to evaluate the primary bacterial degradation rates of the Aroclor products is the test method recommended by the Soap and Detergents Association for the evaluation of the biodegradability of linear alkyl benzene sulfonate type surfactants [JAOCS 42, 986 (1965) & 46, 432 (1969)].

> Primary Biodegradation - Minimum alternation of the chemical structure of the material in question to an extent that characteristic properties of the original material are no longer evident.

This procedure employs sludge from a sewage treatment plant as the source of microorganisms to which a specific amount of the material being evaluated and a synthetic sewage mixture are fed on a periodic basis in a specially designed aeration chamber. The next slide (#8) graphically illustrates what the aeration chamber looks like. It is simply a large glass cylinder with provisions for aeration, auxiliary stirring, a siphon for periodic removal of the supernatant and a septum for introduction of the test material.

The mixed liquor (sludge + water) obtained from the sewage treatment plant is initially adjusted with tap water to a suspended solids concentration of about 2500 mg/l, and 1500 ml of this mixture is then charged to the aeration chamber.

The mechanical cycle employed is shown in the next slide (#9). Each cycle is initiated by the addition of the synthetic sewage and 1 mg of the PCB product being tested.

Since the PCBs are quite water insoluble, they are fed to the unit via injection of 200 uls of a concentrated ethanol solution. In this manner, homogenous dispersion of the PCBs on the bacterial sludge is obtained.

After about one hour of aeration an aliquot of the mixed liquor is withdrawn from the chamber and analyzed for PCBs via UV spectrophotometry and/or electron capture gas chromatography. Aeration is continued for about 48 hours and a second sample is withdrawn for analysis. At this point, the aeration is stopped and the sludge allowed to settle, the sludge volume and pH are then checked to insure that the unit is operating satisfactorily. Two-thirds of the supernatant is withdrawn and replaced with tap water; aeration is then resumed. The cycle is re-initiated by the addition of the synthetic sewage and Aroclor in question. This cycle is continuously repeated until a steady state and consistent degradation rates are obtained.

The per cent degradation rate is calculated as shown in the equation on the slide from the amounts found in the samples analyzed during each cycle.

Degradation testing of the Aroclor products shown in the next slide (#10) have been carried out over an eight month period in our laboratories. In this slide, we have shown graphically the results observed to date. Here we have plotted the mean per cent degradation

rates for biphenyl, Aroclor 1221, MCS 1043, a research material containing 30% by weight chlorine, Aroclor 1016 (41% chlorine), Aroclor 1242, and Aroclor 1254 versus the weight per cent chlorine present in each. The actual mean per cent degradation rates and 95% confidence limits for each material are shown in the lower left portion of the slide. These data were all obtained by UV spectrophotometry which in essence follows the decrease in the aromatic ring content and is indicative of bacterial ring cleavage. The important point to note here is that as the degree of chlorination decreases the degradation rate increases.

In order to give you a feeling for the degradation rates observed with other materials, Aroclor 1221 degrades at about the same rate as a non-linear ABS surfactant.

We have also used this technique to study p,p'-DDT and have at this point in time noted no significant primary degradation.

The next slide (#11) shows the changes in homolog distribution observed for Aroclor 1242 via electron capture gas chromatographic analyses. The upper chromatogram shows the character of the residue one hour after addition. The numbers above each peak indicate the dominant homolog or homologs present in each. The lower chromatogram is of the residue after 72 hours of exposure to the bacterial sludge. It can be readily seen by comparing the two chromatograms that all the dichloro biphenyls, most of the trichloro biphenyls, and a significant portion of the tetrachloro biphenyls are degraded in 48 hours under these test conditions.

The conclusions which we draw from this preliminary data are shown in the next slide (#12).

Next, I will discuss our "Aroclor Residue Studies" (Slide #13).

Our white leghorn chicken studies (Slide #14) have consisted of a 90 day oral exposure of Aroclor 1242, Aroclor 1254, and Aroclor 1260 at 1, 10, and 100 ppm feed levels and a repeat 90 day study of Aroclor 1242 at the 2, 4, and 8 ppm feed levels. 336 Chickens were employed from which a total of 521 tissue, chick, and egg samples were collected. Of these 112 pooled samples were analyzed for PCB residues.

In the next slide (#15) are shown the results of the 90 day oral exposure of white leghorn chickens to Aroclor 1242. On the left side, we have shown the oral exposure levels which were 1, 10, and 100 ppm, the theoretical residue in ppm, which would have been found in the lipid if the chickens had retained all of the Aroclor 1242 which they orally injested. As you can see, these levels are ~125, 1250, and 12,500 ppm. Next is shown the actual average level in ppm found in the lipid of the muscle, fat, and liver samples and then the levels found after 30 days on a PCB free diet. The important points to note are that ~90% of all the Aroclor 1242 consumed is directly excreted and/or metabolized and that after 30 days on a PCB free diet 35%, 44%, and 57% of the PCBs retained after 90 days of continuous exposure at the 1, 10, and 100 ppm levels was excreted and/or metabolized.

On the right hand side of this slide is shown the homolog distribution of the product fed and that of the residues isolated from the tissues after 90 days of exposure and 30 days on a PCB free diet. The numbers across the top simply refer to the number of chlorine atoms per biphenyl molecule. As you can see, Aroclor 1242 contains dominant amounts of the di- through pentachlorobiphenyls and a minor amount of hexachlorobiphenyl. After 90 days of exposure the dichlorobiphenyl was no longer observable and the dominant components were the tri- through pentachloro biphenyl homologs. After 30 days on a PCB free recovery diet, the hexachloro biphenyl is now a dominant component because of continued excretion and/or metabolism of the lower chlorinated homologs.

In the next slide (#16) are shown the results for the 90 day oral exposure of Aroclor 1254 in white leghorn chickens at the 1, 10, and 100 ppm exposure levels. The theoretical residues are the same as before and we have again shown the actual levels found in the tissues after 90 days of continuous exposure and 30 days on a PCB free diet. In this instance, ∿70-72% of the Aroclor 1254 ingested was directly excreted and/or metabolized and after 30 days on a PCB free diet, ∿45% of the residues retained were excreted and/or metabolized.

The homolog distribution of the product and residues is shown on the right of the slide, the product Aroclor 1254 contains minor amounts of the tri- and heptachloro homologs and dominant amounts of the tetra-, penta-, and hexachloro biphenyls. The residue after 90 days of exposure did not contain detectable amounts of the trichloro biphenyls and the tetrachloro biphenyls were no longer a dominant component. The dominant homologs were penta- and hexachloro biphenyls. After 30 days on a PCB free diet, the tetrachloro biphenyls were now not detectable, the pentachloro biphenyls were a minor component, and the hexachloro biphenyls the dominant component.

In the next slide (#17) are the results for Aroclor 1260. Again, the oral exposure level and theoretical residue levels are the same and the PCB residues found in the tissues after 90 days of continuous exposure and 30 days on a PCB free recovery diet are shown. After 90 days of exposure, 57-52% of all Aroclor 1260 consumed was directly excreted and/or metabolized and after 30 days on a PCB free diet ∿40% of the residues retained were excreted and/or metabolized.

As shown on the right, Aroclor 1260 contains dominant amounts of penta-, hexa-, and heptachloro biphenyls and a minor amount of octachloro biphenyl. The residues after 90 days of exposure and 30 days on a PCB free recovery diet contain minor amounts of the penta- and octachloro homologs. In both cases, the dominant homologs were the hexa- and heptachloro biphenyls.

Our albino rat work (Slide #18) has consisted of 30 day oral, 2 year chronic oral and a 3 generation rat reproduction exposure study with Aroclor 1242, Aroclor 1254, and Aroclor 1260 and a 90 day subacute

oral exposure of Aroclor 1221. Approximately 1400 animals were used in these exposure studies from which about 400 muscle, liver, and fat samples were collected. Two hundred of these samples were analyzed for PCB residues.

In the next slide (#19) are shown the results of our two year chronic oral exposure study of Aroclor 1242 in albino rats. The oral exposure levels were 1, 10, and 100 ppm and the theoretical residues were ∿800, 8000, 80,000 ppm, respectively. The actual residues found in the tissue lipid are shown after 3, 12, and 24 months of exposure. Comparison of the residues found after 24 months to the theoretical residue levels indicates that 99% of the Aroclor 1242 fed was directly excreted and/or metabolized at all exposure levels.

As is shown on the right, Aroclor 1242 contains dominant amounts of the di- through pentachloro biphenyl homologs and a minor amount of the hexa-. The residues after two years did not contain a significant amount of the dichloro biphenyls and the dominant components were the tri-, tetra-, and pentachloro biphenyl homologs.

In the next slide (#20) are shown the results of the two year chronic oral exposure of albino rats to Aroclor 1254. The exposure and theoretical residue levels are the same as with Aroclor 1242. The residues found after 3, 12, and 24 months of exposure are also shown. Comparison of the residues after two years to the amount ingested demonstrates that 95-95% of the Aroclor 1254 consumed is directly excreted and/or metabolized.

The homolog distribution of Aroclor 1254 and the residues are shown on the right. Aroclor 1254 contains minor amounts of the tri- and heptachloro biphenyl homologs and dominant amounts of the tetra-, penta-, and hexachloro biphenyl homologs. The residues did not contain detectable levels of the trichloro homologs and the tetrachloro biphenyls were no longer a dominant component. The dominant homologs were the penta- and hexachloro biphenyls.

The next slide (#21) shows the the results for the two year exposure of Aroclor 1260 in albino rats. Again, the exposure and theoretical residue levels are the same and the residues found in the tissues after 3, 12, and 24 months of exposure are shown. In this case 93-95% of all Aroclor 1260 ingested was directly excreted and/or metabolized.

The dominant homologs in Aroclor 1260 and the residues isolated from the tissues were similar in all cases.

In order to demonstrate the relationship between residue storage levels and the degree of chlorination of the product fed (slide #22). I have plotted the average ppm PCB found in the lipid vs the weight per cent chlorine in the product fed. These data were taken from our 90 day subacute albino rat studies with Aroclor 1221, Aroclor 1242, Aroclor 1254, and Aroclor 1260 at an exposure level of 100 ppm. As you can see, the residue storage levels decrease exponentially as the weight per cent chlorine decreases, simply demonstrating the relationship between the higher homolog content of an Aroclor product and the tissue storage level.

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Our beagle dog studies (Slide #23) have consisted of a two year chronic exposure of Aroclor 1242, Aroclor 1254, and Aroclor 1260 and a 90 day subacute study of Aroclor 1221. In these studies, 108 beagle dogs were used, resulting in the collection of 263 samples, and the analyses of 146 for PCB residues.

The next slide (#24) shows the results of one two year study of Aroclor 1242 at exposure levels of 1, 10, and 100 ppm. In this study, the theoretical residue levels are ~500, 5000, and 50,000 ppm respectively. We have also shown on this slide the residue levels found after two years of exposure and after 30 and 60 day periods on PCB free recovery diets. The beagle dogs directly excreted and/or metabolized 99.6% of the Aroclor 1242 consumed and after 60 days on PCB free diets, 50-60% of PCB residue retained after two years of exposure was excreted and/or metabolized.

Aroclor 1242 contains dominant amounts of the di-, tri-, tetra-, and pentachloro homologs and a minor amount of the hexa- homolog. The residue found after two years of exposure contained no detectable levels of the dichloro homologs and dominant levels of the tri-, tetra-, hepta-, and octachloro biphenyls. The pentachloro homolog, although dominant in the product fed, was not a dominant component of the residue. After 30 days on a PCB free recovery diet, the di-, tri-, and tetrachloro biphenyls were not detectable components of the residue. At this the dominant components were the hexa-, hepta-, and octachloro biphenyls. The trend toward excretion and/or metabolism of the lower chlorinated homologs continued to the extent that after 60 days on the recovery diet the hexachloro biphenyl was no longer a dominant component, and the hepta- and octachloro biphenyls became the dominant constituents in the residue.

In the next slide (#25) are the results for the two year exposure of beagle dogs to Aroclor 1254. The oral exposure and theoretical residue levels are the same and the PCB residue levels found in the tissues after two years of exposure and after 30 and 60 day periods on PCB free recovery diets are again shown.

In this instance, the dogs excreted and/or metabolized 98-99% of all Aroclor 1254 consumed over a two year period. After 60 days on a PCB free recovery diet, 30-40% of the residues retained were excreted and/or metabolized. The product fed, Aroclor 1254, contains minor amounts of the tri- and heptachloro biphenyls and dominant amounts of the tetra-, penta-, and hexachloro homologs. After two years of exposure, the residue retained from the product did not contain detectable levels of the tri- or tetrachloro biphenyls, the penta-, and hexachloro biphenyls remained dominant components, and the heptachloro biphenyls became dominant constituents.

After 30 days on the PCB free recovery diet, the pentachloro homologs became a minor component of the residue, the hexa- and heptachloro biphenyls remained dominant components and the octachloro homologs became a minor component. After 60 days on the PCB free recovery diet, the pentachloro biphenyls were excreted and/or metabolized to the extent that the octachloro homologs became a dominant constituent of the residue.

The next slide (#26) shows the results for the two year oral exposure residue study of Aroclor 1260 in beagle dogs. The exposure and theoretical residue levels are the same as those for the Aroclor 1242 and Aroclor 1254 studies. Next is shown the residues which accumulated after two years of continuous exposure and the residues retained after 30 and 60 day recovery periods on PCB free diets. With this Aroclor 98-99% of the amount consumed over two years was directly excreted and/or metabolized. After 60 days on the recovery diet ~13% of the retained residues were excreted and/or metabolized.

The homolog distribution of Aroclor 1260 is as shown, dominant amounts of the penta-, hexa-, and heptachloro biphenyls with a minor amount of the octachloro homologs. After two years, the PCB residue contains no detectable level of the pentachloro homologs, a minor amount of the heptachloro biphenyls, and dominant amounts of the hexachloro and octachloro homologs. After 30 days on the recovery diet, the dominant homologs are now the hexa-, hepta-, and octachloro biphenyls becoming a dominant component of the residue via loss of some of the hexachloro biphenyls. After 60 days on the recovery diet, the hexachloro biphenyls are no longer dominant components of the residue and it is now mainly the hepta- and octachloro biphenyls.

The next slide (#27) illustrates the residue fall off as a function of Aroclor and recovery period. These data are from the two year beagle dog studies and the exposure level is 1 ppm. In this graph, I have plotted the average ppm PCB found in the lipid for Aroclor 1260, Aroclor 1254, and Aroclor 1242 after two years of continuous oral exposure and then after 1 month and 2 month recovery periods on PCB free diets. This plot simply demonstrates that the PCB residues retained from Aroclor 1242 fall off more quickly than those retained from Aroclor 1254 and Aroclor 1260.

The table in the next slide (#28) shows the relative ability of fowl, small mammals, and large mammals to retain orally ingested PCBs. These data are for Aroclor 1242 at the exposure levels and periods shown. The concentration factor is calculated by dividing the maximum PCB level found in the lipid by the exposure level. As you can see from the factors, chickens retain PCBs to a greater extent than do rats or dogs.

Our fish residue work is not very extensive at this point - primarily because we have had problems in finding consulting laboratories capable of carrying out dynamic low level fish exposure studies, and secondly, because government laboratories such as those in Duluth, Minnesota, Columbia, Missouri, and Gulf Breeze, Florida were, and are, still in better positions to carry out and evaluate these types of studies.

We have done some very preliminary 21 day dynamic exposures of catfish and bluegill fingerlings to some of our Aroclor products and it generally supports the conclusions which can be drawn from literature data.

The general conclusions which we draw from these residue studies are shown in the next three slides.

Slide #29 - Build-Up - Conclusions

Slide #30 - Fall-Off - Conclusions

Slide #31 - Alteration of Homolog Distribution - Conclusions

In summary, we feel that the results of our preliminary research support, what has and is being observed via residue analysis of environmental samples; that is to say, from a residue viewpoint, that the bulk of the PCB homologs released to the environment (PCBs with less than 5 chlorines) are subject to environmental degradation of one sort or another at measurable rates and as such have not accumulated.

E. S. Tucker

TYPICAL % COMPOSITION OF POLYCHLORINATED BIPHENYL PRODUCTS *

HOMOLOG # Cl/BIPHENYL	AROCLOR 1221	AROCLOR 1016	AROCLOR 1242	AROCLOR 1254
0	11	<0.1	<0.1	<0.1
1	51	1	1	<0.1
2	32	20	16	<0.5
3	4	57	49	1
4	2	21	25	21
5	<0.5	1	8	48
6	ND	<0.1	1	23
7	ND	ND	<0.1	6
8	ND	ND	ND	ND

*PER CENT (W/W) BY GC/MASS USING AREA CORRECTION FACTORS BY HOMOLOG RESPONSE

NONE DETECTED, <0.01% = ND

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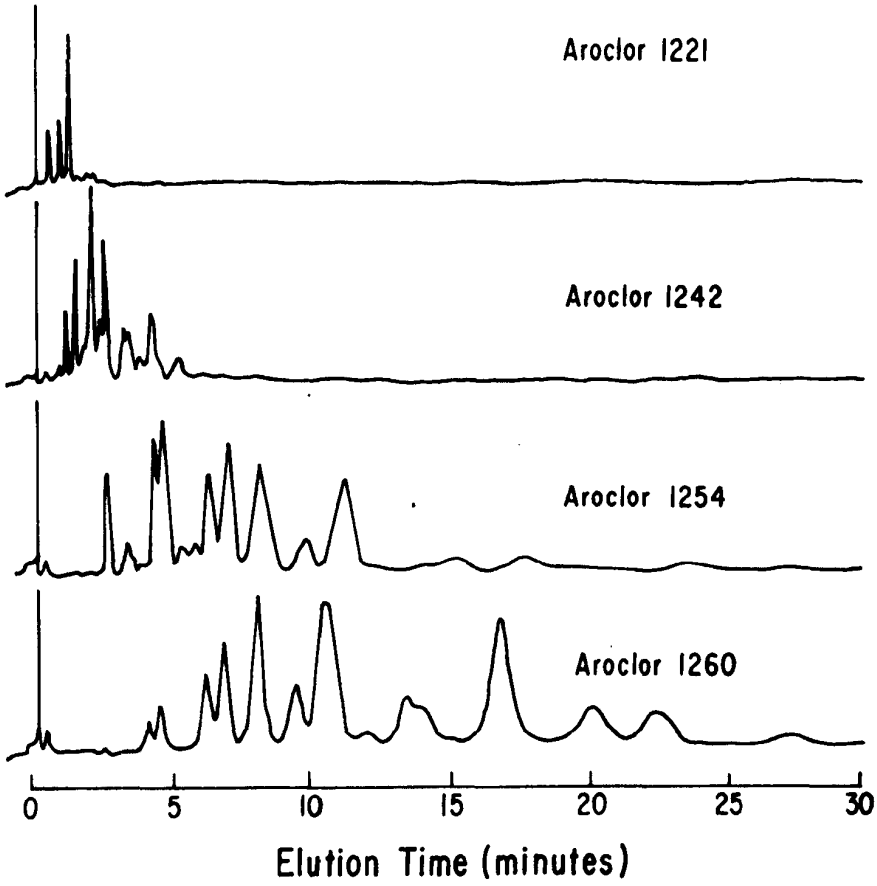
Electron Capture Gas Chromatograms
Aroclor 1221
Aroclor 1242
Aroclor 1254
Aroclor 1260
0
5
10
15
20
25
30
Elution Time (minutes)

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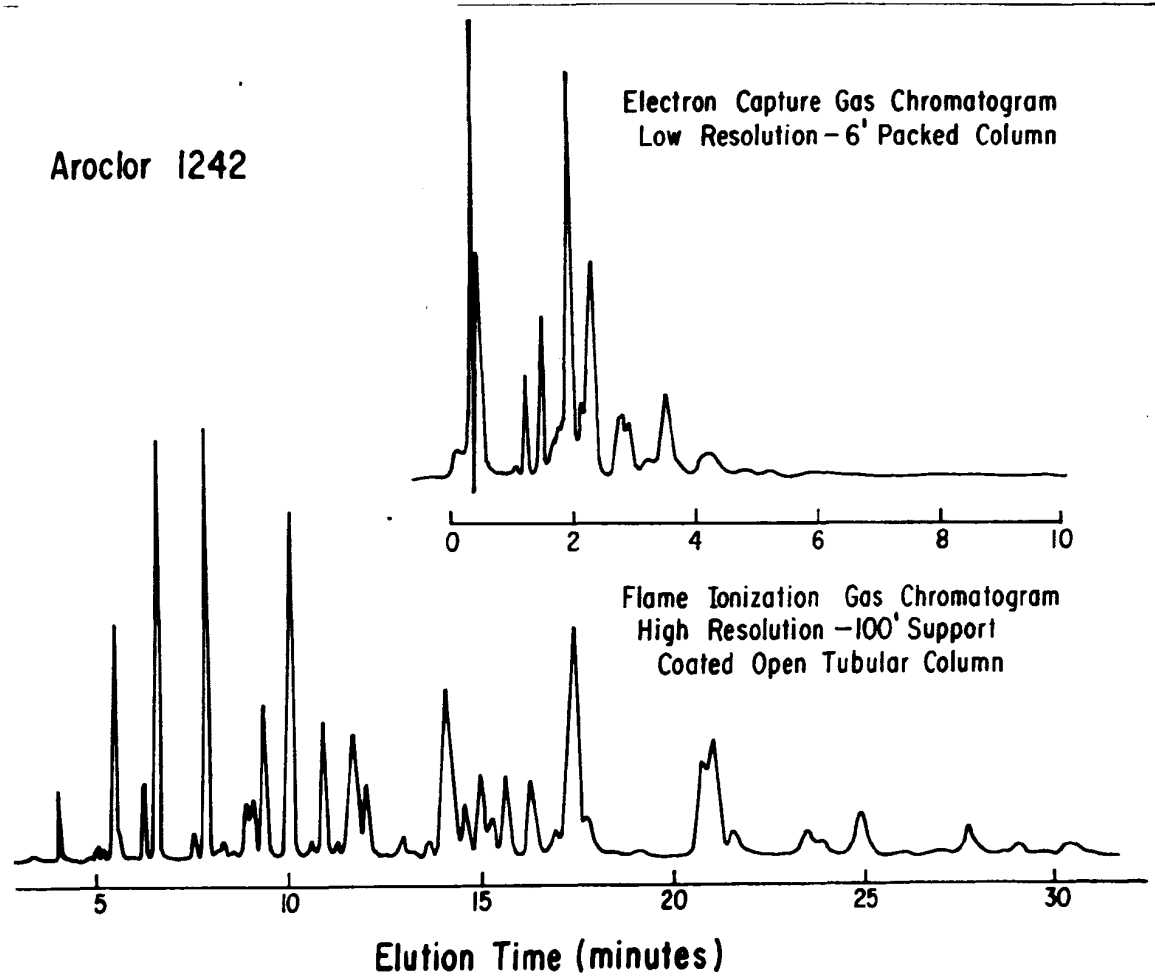
Aroclor 1242
Electron Capture Gas Chromatogram
Low Resolution – 6' Packed Column
0
2
4
6
8
10
Flame Ionization Gas Chromatogram
High Resolution – 100' Support
Coated Open Tubular Column
5
10
15
20
25
30
Elution Time (minutes)

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- PCB PRODUCTS ARE NOT A SINGLE ENTITY, BUT COMPLEX MULTI-COMPONENT MIXTURES

- PCB RESIDUES FOUND IN WILD LIFE ARE DOMINANTLY PENTA-, HEXA-, HEPTA-, AND OCTA-CHLORO BIPHENYLS

- PCB RESIDUE ARE MOST SIMILAR TO AROCLOR 1254 AND AROCLOR 1260 PRODUCTS

PRIMARY BACTERIAL DEGRADATION STUDIES

- SEMI-CONTINUOUS ACTIVATED SLUDGE DEGRADATION

RESIDUE ACCUMULATION STUDIES

- FISH - CATFISH AND BLUEGILLS
- BIRDS - WHITE LEGHORN CHICKENS
- MAMMALS - ALBINO RATS AND BEAGLE DOGS

ASSESSMENT OF THE PERSISTENCE OF POLYCHLORINATED BIPHENYLS IN BIOLOGICAL SYSTEMS

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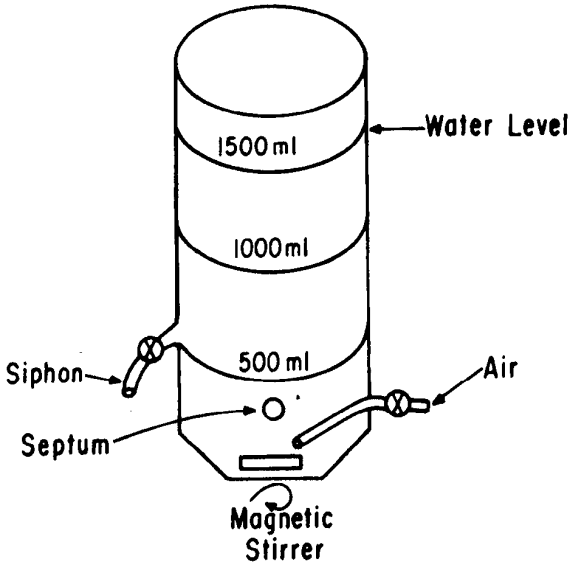
AROCLOR SEMI-CONTINUOUS ACTIVATED SLUDGE PRIMARY DEGRADATION STUDIES

MONS 069957

Semi-Continuous Activated Sludge Test Unit



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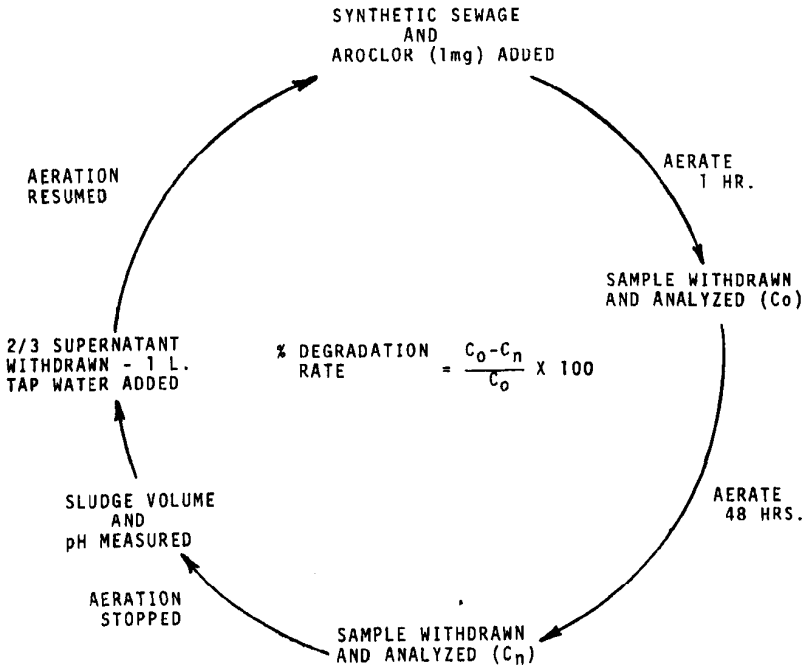
MECHANICAL CYCLE



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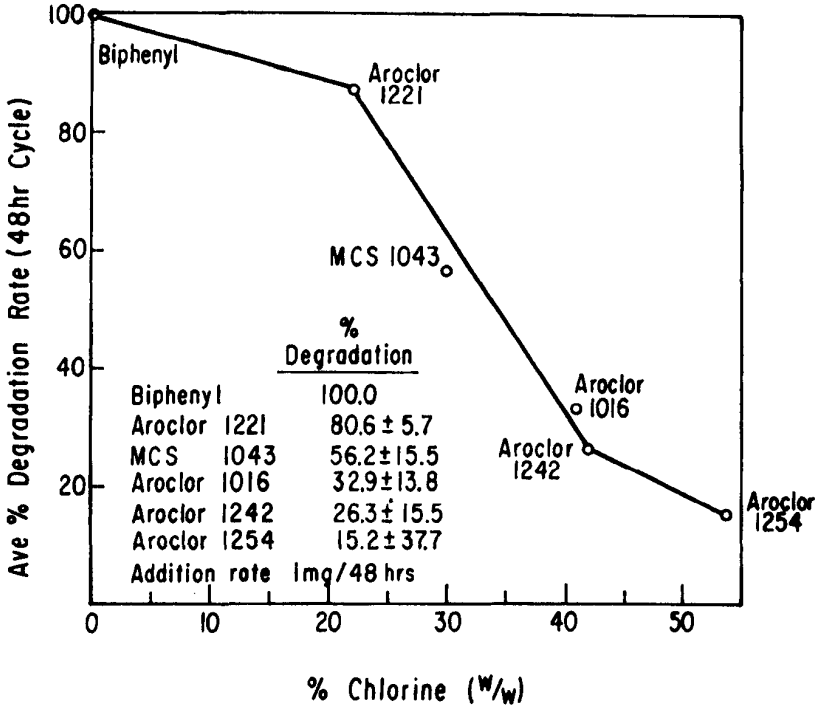
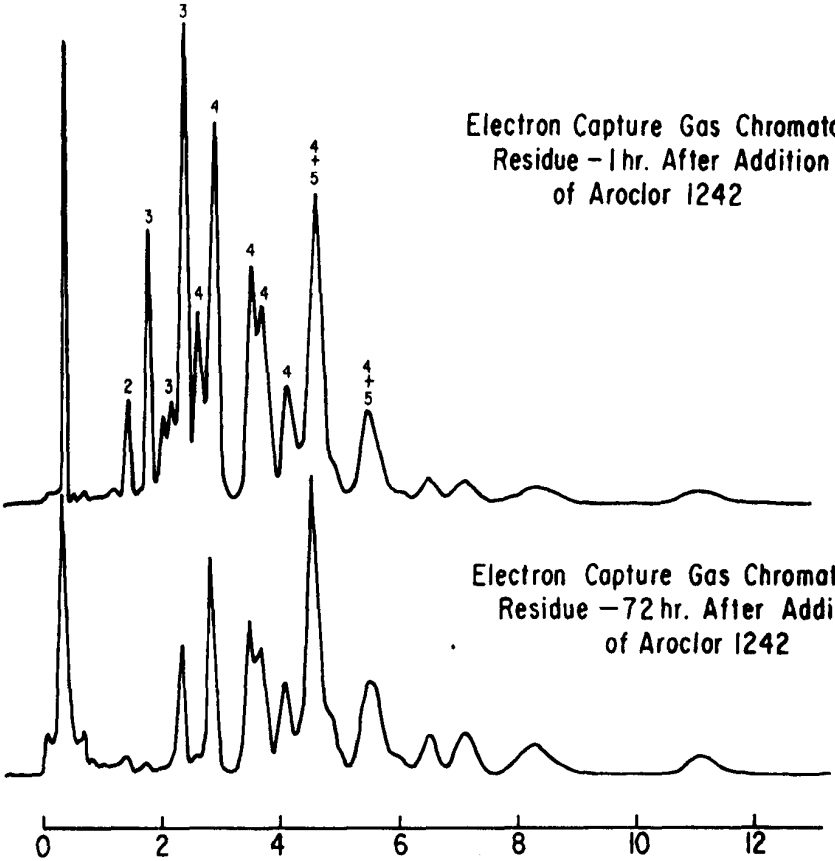
Semi-Continuous Activated Sludge Degradation of Polychlorinated Biphenyls



MONS 069960

Semi-Continuous Activated Sludge Degradation of Aroclor 1242



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BACTERIAL DEGRADATION STUDIES

CONCLUSIONS

- RATE OF PRIMARY DEGRADATION INCREASES AS THE DEGREE OF CHLORINATION OF THE AROCLOR PRODUCT DECREASES

BIPHENYL>AROCLOR 1221>MCS 1043 >AROCLOR 1016>AROCLOR 1242>AROCLOR 1254

- BIPHENYL, MONO-, DI-, TRI-, AND TETRACHLORO BIPHENYL HOMOLOGS UNDERGO PRIMARY BACTERIAL DEGRADATION

AROCLOR RESIDUE STUDIES

WHITE LEGHORN CHICKEN STUDIES

PRODUCTS STUDIED		90 DAY ORAL AROCLOR 1242 AROCLOR 1254 AROCLOR 1260	AROCLOR 1242
NUMBER OF CHICKENS	FEMALE	200	80
	MALE	40	16
NUMBER OF SAMPLES	MUSCLE	40	8
	LIVER	40	8
	FAT	40	8
	EGGS	∿250	∿60
	CHICKS	30	10
NUMBER OF SAMPLES ANALYZED	MUSCLE	14	5
	LIVER	12	4
	FAT	18	5
	EGGS	25	8
	CHICKS	11	10
TOTAL SAMPLES COLLECTED		400	121
TOTAL ANALYZED		80	32

ORAL EXPOSURE CARRIED OUT BY
INDUSTRIAL BIO-TEST LABORATORIES, INC.
NORTHBROOK, ILLINOIS

RESIDUE STUDY OF AROCLOR 1242 IN WHITE LEGHORN CHICKENS

90 DAY ORAL

	PPM PCBs			HOMOLOG DISTRIBUTION									
				1	2	3	4	5	6	7	8	9	10
ORAL EXPOSURE LEVEL	1	10	100										
†THEORETICAL RESIDUE	126	1260	12602	-	(2)	(3)	(4)	(5)	6	-	-	-	-
*RESIDUE, 12 WEEK EXPOSURE	14	136	1312	-	-	(3)	(4)	(5)	6	-	-	-	-
*RESIDUE, 30 DAY RECOVERY	9	77	749	-	-	(3)	(4)	(5)	(6)	-	-	-	-

†IF TOTAL PCB CONSUMED WERE RETAINED

*AVERAGE PPM IN LIPID - ALL TISSUES

( )ELECTRON CAPTURE PEAKS GREATER THAN 5% OF TOTAL

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RESIDUE STUDY OF AROCLOR 1254 IN WHITE LEGHORN CHICKENS

90 DAY ORAL

	PPM PCBs			HOMOLOG DISTRIBUTION									
				1	2	3	4	5	6	7	8	9	10
ORAL EXPOSURE LEVEL	1	10	100										
†THEORETICAL RESIDUE	126	1260	12602	-	-	3	(4)	(5)	(6)	7	-	-	-
*RESIDUE, 12 WEEK EXPOSURE	38	362	3506	-	-	-	4	(5)	(6)	7	-	-	-
*RESIDUE, 30 DAY RECOVERY	17	164	1580	-	-	-	-	5	(6)	7	-	-	-

†IF TOTAL PCB CONSUMED WERE RETAINED

*AVERAGE PPM IN LIPID - ALL TISSUES

( )ELECTRON CAPTURE PEAKS GREATER THAN 5% OF TOTAL

MONS 069966

RESIDUE STUDY OF AROCLOR 1260 IN WHITE LEGHORN CHICKENS

90 DAY ORAL

	PPM PCBs			HOMOLOG DISTRIBUTION									
				1	2	3	4	5	6	7	8	9	10
ORAL EXPOSURE LEVEL	1	10	100										
†THEORETICAL RESIDUE	126	1260	12602	-	-	-	-	(5)	(6)	(7)	8	-	-
*RESIDUE, 12 WEEK EXPOSURE	65	607	5909	-	-	-	-	5	(6)	(7)	8	-	-
*RESIDUE, 30 DAY RECOVERY	26	232	2363	-	-	-	-	5	(6)	(7)	8	-	-

†IF TOTAL PCB CONSUMED WERE RETAINED

*AVERAGE PPM IN LIPID - ALL TISSUES

( )ELECTRON CAPTURE PEAKS GREATER THAN 5% OF TOTAL

MDNS 069967

ALBINO RAT STUDIES

PRODUCTS STUDIED		30 DAY ORAL AROCLOR 1242 AROCLOR 1254 AROCLOR 1260	2 YEAR CHRONIC ORAL AROCLOR 1242 AROCLOR 1254 AROCLOR 1260	3 GENERATION RAT REPRODUCTION AROCLOR 1242 AROCLOR 1254 AROCLOR 1260	90 DAY SUBACUTE ORAL AROCLOR 1221
NUMBER OF RATS	FEMALE	115	500	~20	60
	MALE	115	500	~20	60
NUMBER OF SAMPLES	MUSCLE	18	33	-	7
	KIDNEY	18	-	-	-
	LIVER	18	33	-	7
	FAT	18	33	-	7
	PUPS	-	-	10	-
NUMBER OF SAMPLES ANALYZED	MUSCLE	18	33	-	7
	KIDNEY	18	-	-	-
	LIVER	18	33	-	7
	FAT	18	33	-	7
	PUPS	-	-	10	-
TOTAL SAMPLES COLLECTED		144	198	20	42
TOTAL ANALYZED		72	99	10	21

ORAL EXPOSURE CARRIED OUT BY
INDUSTRIAL BIO-TEST LABORATORIES, INC.
NORTHBROOK, ILLINOIS

MONS 069968

RESIDUE STUDY OF AROCLOR 1242 IN ALBINO RATS

TWO YEAR CHRONIC ORAL EXPOSURE

ORAL EXPOSURE LEVEL	PPM PCBs 1	10	100	HOMOLOG DISTRIBUTION 1	2	3	4	5	6	7	8	9	10
†THEORETICAL RESIDUE	803	8037	80373	-	(2)	(3)	(4)	(5)	6	-	-	-	-
*RESIDUE, 3 MONTH EXPOSURE	6	23	90	-	-	(3)	(4)	(5)	6	-	-	-	-
*RESIDUE, 12 MONTH EXPOSURE	9	37	155	-	-	(3)	(4)	(5)	6	-	-	-	-
*RESIDUE, 24 MONTH EXPOSURE	12	53	240	-	-	(3)	(4)	(5)	6	-	-	-	-

†IF TOTAL PCB CONSUMED WERE RETAINED

*AVERAGE PPM IN LIPID - ALL TISSUES

( )ELECTRON CAPTURE PEAKS GREATER THAN 5% OF TOTAL

MONS 069969

RESIDUE STUDY OF AROCLOR 1254 IN ALBINO RATS

TWO YEAR CHRONIC ORAL EXPOSURE

	PPM PCBs			HOMOLOG DISTRIBUTION									
				1	2	3	4	5	6	7	8	9	10
ORAL EXPOSURE LEVEL	1	10	100										
†THEORETICAL RESIDUE	803	8037	80373	-	-	3	(4)	(5)	(6)	7	-	-	-
*RESIDUE, 3 MONTH EXPOSURE	13	94	679	-	-	-	4	(5)	(6)	7	-	-	-
*RESIDUE, 12 MONTH EXPOSURE	24	189	1471	-	-	-	4	(5)	(6)	7	-	-	-
*RESIDUE, 24 MONTH EXPOSURE	42	355	3038	-	-	-	4	(5)	(6)	7	-	-	-

†IF TOTAL PCB CONSUMED WERE RETAINED

*AVERAGE PPM IN LIPID - ALL TISSUES

( )ELECTRON CAPTURE PEAKS GREATER THAN 5% OF TOTAL

MONS 069920

RESIDUE STUDY OF AROCLOR 1260 IN ALBINO RATS

TWO YEAR CHRONIC ORAL EXPOSURE

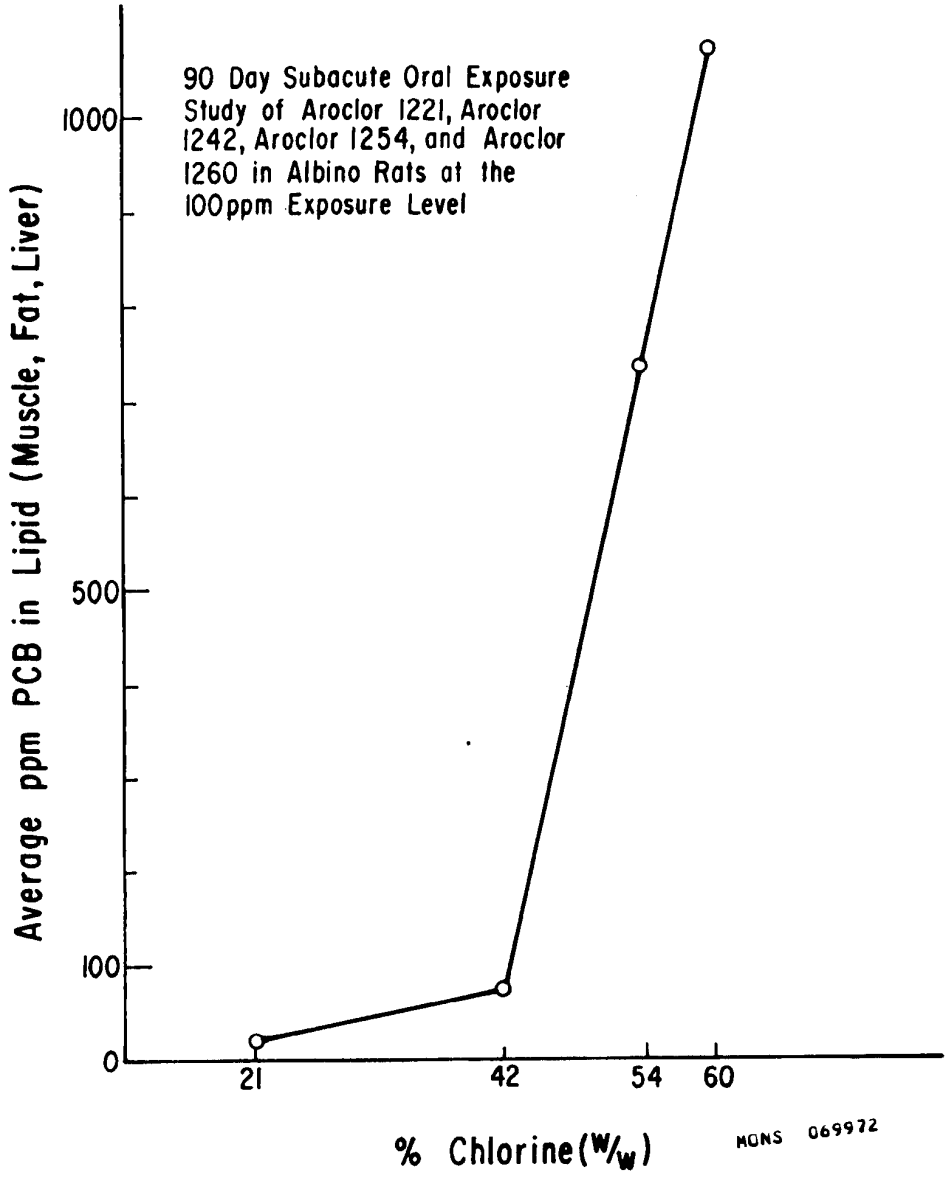
ORAL EXPOSURE LEVEL	PPM PCBs 1	10	100	HOMOLOG DISTRIBUTION 1	2	3	4	5	6	7	8	9	10
†THEORETICAL RESIDUE	803	8037	80373	-	-	-	-	(5)	(6)	(7)	8	-	-
*RESIDUE, 3 MONTH EXPOSURE	18	134	970	-	-	-	-	(5)	(6)	(7)	8	-	-
*RESIDUE, 12 MONTH EXPOSURE	35	270	2099	-	-	-	-	(5)	(6)	(7)	8	-	-
*RESIDUE, 24 MONTH EXPOSURE	59	507	4338	-	-	-	-	(5)	(6)	(7)	8	-	-

†IF TOTAL PCB CONSUMED WERE RETAINED

*AVERAGE PPM IN LIPID - ALL TISSUES

( )ELECTRON CAPTURE PEAKS GREATER THAN 5% OF TOTAL

MCNS 069921


90 Day Subacute Oral Exposure Study of Aroclor 1221, Aroclor 1242, Aroclor 1254, and Aroclor 1260 in Albino Rats at the 100ppm Exposure Level
Average ppm PCB in Lipid (Muscle, Fat, Liver)
1000
500
100
0
21
42
54
60
% Chlorine (W/W)
MONS 069972

BEAGLE DOG STUDIES

	2 YEAR CHRONIC ORAL	90 DAY SUBACUTE ORAL
PRODUCTS STUDIED	AROCLOR 1242 AROCLOR 1254 AROCLOR 1260	AROCLOR 1221
NUMBER OF DOGS) FEMALE	40	14
NUMBER OF DOGS) MALE	40	14
NUMBER OF SAMPLES) MUSCLE	54	7
NUMBER OF SAMPLES) LIVER	54	7
NUMBER OF SAMPLES) FAT	54	7
TOTAL SAMPLES COLLECTED	242	21
NUMBER OF SAMPLES ANALYZED) MUSCLE	42	7
NUMBER OF SAMPLES ANALYZED) LIVER	40	7
NUMBER OF SAMPLES ANALYZED) FAT	43	7
TOTAL ANALYZED	125	21

ORAL EXPOSURE CARRIED OUT BY
INDUSTRIAL BIO-TEST LABORATORIES, INC.
NORTHBROOK, ILLINOIS

RESIDUE STUDY OF AROCLOR 1242 IN BEAGLE DOGS

TWO YEAR CHRONIC ORAL EXPOSURE

ORAL EXPOSURE LEVEL	PPM PCBs			HOMOLOG DISTRIBUTION									
	1	10	100	1	2	3	4	5	6	7	8	9	10
†THEORETICAL RESIDUE	519	5186	51865	-	(2)	(3)	(4)	(5)	6	-	-	-	-
*RESIDUE, 2 YEAR EXPOSURE	2	6	12	-	-	(3)	(4)	5	6	(7)	(8)	-	-
*RESIDUE, 30 DAY RECOVERY	1	4	10	-	-	-	-	5	(6)	(7)	(8)	-	-
*RESIDUE, 60 DAY RECOVERY	0.8	3	6	-	-	-	-	5	6	(7)	(8)	-	-

†IF TOTAL PCB CONSUMED WERE RETAINED

*AVERAGE PPM IN LIPID - ALL TISSUES

( )ELECTRON CAPTURE PEAKS GREATER THAN 5% OF TOTAL

MONS 069674

RESIDUE STUDY OF AROCLOR 1254 IN BEAGLE DOGS

TWO YEAR CHRONIC ORAL EXPOSURE

ORAL EXPOSURE LEVEL	PPM PCBs 1	10	100	HOMOLOG DISTRIBUTION 1	2	3	4	5	6	7	8	9	10
†THEORETICAL RESIDUE	519	5186	51865	-	-	3	(4)	(5)	(6)	7	-	-	-
*RESIDUE, 2 YEAR EXPOSURE	7	30	132	-	-	-	-	(5)	(6)	(7)	8	-	-
*RESIDUE, 30 DAY RECOVERY	5	24	109	-	-	-	-	5	(6)	(7)	8	-	-
*RESIDUE, 60 DAY RECOVERY	4	20	88	-	-	-	-	5	(6)	(7)	(8)	-	-

†IF TOTAL PCB CONSUMED WERE RETAINED

*AVERAGE PPM IN LIPID - ALL TISSUES

( )ELECTRON CAPTURE PEAKS GREATER THAN 5% OF TOTAL

MONS 069975

RESIDUE STUDY OF AROCLOR 1260 IN BEAGLE DOGS

TWO YEAR CHRONIC ORAL EXPOSURE

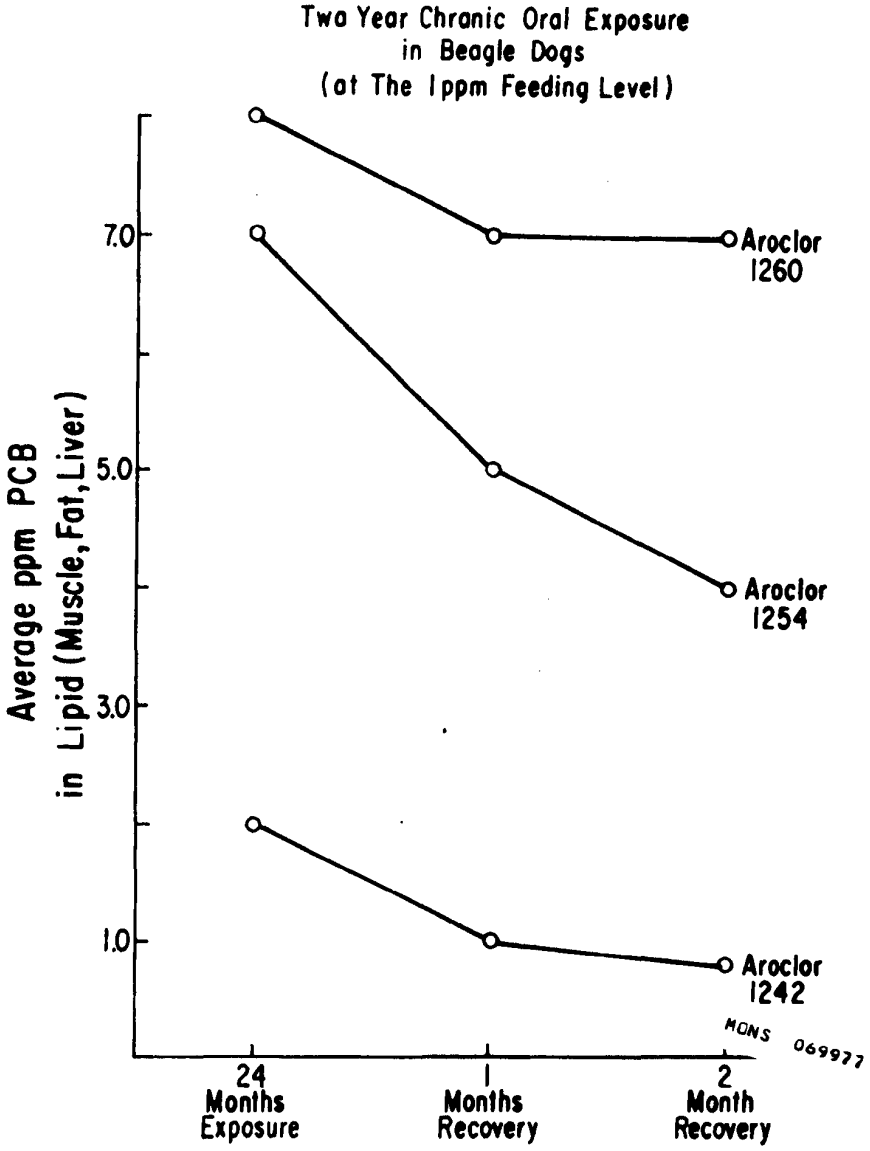
	PPM PCBs			HOMOLOG DISTRIBUTION									
				1	2	3	4	5	6	7	8	9	10
ORAL EXPOSURE LEVEL	1	10	100										
†THEORETICAL RESIDUE	519	5186	51865	-	-	-	-	(5)	(6)	(7)	8	-	-
*RESIDUE, 2 YEAR EXPOSURE	8	63	388	-	-	-	-	-	(6)	7	(8)	-	-
*RESIDUE, 30 DAY RECOVERY	7	60	363	-	-	-	-	-	(6)	(7)	(8)	-	-
*RESIDUE, 60 DAY RECOVERY	7	55	337	-	-	-	-	-	6	(7)	(8)	-	-

†IF TOTAL PCB CONSUMED WERE RETAINED

*AVERAGE PPM IN LIPID - ALL TISSUES

( )ELECTRON CAPTURE PEAKS GREATER THAN 5% OF TOTAL

MONS 069976


Two Year Chronic Oral Exposure
in Beagle Dogs
(at The 1ppm Feeding Level)
Average ppm PCB
in Lipid (Muscle, Fat, Liver)
7.0
5.0
3.0
1.0
Aroclor
1260
Aroclor
1254
Aroclor
1242
MONS 069977
24
Months
Exposure
1
Months
Recovery
2
Month
Recovery

	EXPOSURE LEVEL (PPM)	EXPOSURE PERIOD	CONCENTRATION FACTOR*
WHITE LEGHORN CHICKENS	1.0 (FEED)	12 WEEKS	20
ALBINO RATS	1.0 (FEED)	2 YEARS	9
BEAGLE DOGS	1.0 (FEED)	2 YEARS	2

*CONCENTRATION FACTORS DO NOT TAKE INTO ACCOUNT EXPOSURE PERIODS

CONCLUSIONS

RESIDUE BUILDUP STUDIES

- FOR ALL PRODUCTS THE PCB RESIDUE LEVEL INCREASED AS THE EXPOSURE LEVEL AND PERIOD INCREASED

- RELATIVE TO AROCLOR 1260, THE HIGHEST CHLORINATED PRODUCT STUDIED, THE PCB RESIDUE LEVEL DECREASED EXPONENTIALLY AS AS THE WEIGHT % CHLORINE OF THE PRODUCT DECREASED

- RELATIVE LEVELS OF PCB RESIDUES FOUND WERE -

  FISH>>>>>CHICKENS>RATS>DOGS

- RELATIVE PCB RESIDUE LEVELS IN TISSUES WERE -

CHICKENS	FAT = MUSCLE>LIVER
RATS	LIVER>FAT = MUSCLE>KIDNEY
DOGS	LIVER>FAT = MUSCLE

- FOR ALL PRODUCTS THE DOGS AND RATS EXCRETED AND/OR METABOLIZED 93 TO 99% OF THE TOTAL AROCLOR ORALLY INGESTED

- CHICKENS EXCRETED AND/OR METABOLIZED PRODUCTS INJESTED ORALLY AS FOLLOWS -

  90% OF AROCLOR 1242
  70% OF AROCLOR 1254
  50% OF AROCLOR 1260

CONCLUSIONS

RESIDUE FALL-OFF

- DOGS AND CHICKENS CONTINUED TO EXCRETE AND/OR METABOLIZE PCB RESIDUES, INCLUDING PENTA AND HEXA HOMOLOGS, RETAINED WHEN PLACED ON PCB FREE RECOVERY DIETS

  DOGS > CHICKENS

- RELATIVE RESIDUE FALL-OFF RATES SHOWED -

  DOGS: AROCLOR 1242>AROCLOR 1254 >AROCLOR 1260

  CHICKENS: AROCLOR 1242>AROCLOR 1254>AROCLOR 1260

CONCLUSIONS

RESIDUE STUDIES

ALTERATIONS OF AROCLOR HOMOLOG DISTRIBUTION

- ALTERATION OF THE HOMOLOG DISTRIBUTION OF ALL PRODUCTS WAS OBSERVED IN ALL STUDIES EXCEPT FISH

- LOWER CHLORINATED HOMOLOGS PRESENT IN ALL PRODUCTS WERE PREFERENTIALLY EXCRETED AND/OR METABOLIZED

- ALTERATION OF THE HOMOLOG DISTRIBUTION INCREASED AS THE WEIGHT % CHLORINE OF THE PRODUCTS DECREASED (INCLUDING PENTA AND HEXA HOMOLOGS)

  AROCLOR 1221>>AROCLOR 1242>AROCLOR 1254 >AROCLOR 1260

  DOGS>RATS>CHICKENS>>>>>FISH