

REVIEW

Environmental epigenetics

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Epigenetics investigates heritable changes in gene expression that occur without changes in DNA sequence. Several epigenetic mechanisms, including DNA methylation and histone modifications, can change genome function under exogenous influence. We review current evidence indicating that epigenetic alterations mediate effects caused by exposure to environmental toxicants. Results obtained from animal models indicate that *in utero* or early-life environmental exposures produce effects that can be inherited transgenerationally and are accompanied by epigenetic alterations. The search for human equivalents of the epigenetic mechanisms identified in animal models is under way. Recent investigations have identified a number of environmental toxicants that cause altered methylation of human repetitive elements or genes. Some exposures can alter epigenetic states and the same and/or similar epigenetic

alterations can be found in patients with the disease of concern. On the basis of current evidence, we propose possible models for the interplay between environmental exposures and the human epigenome. Several investigations have examined the relationship between exposure to environmental chemicals and epigenetics, and have identified toxicants that modify epigenetic states. Whether environmental exposures have transgenerational epigenetic effects in humans remains to be elucidated. In spite of the current limitations, available evidence supports the concept that epigenetics holds substantial potential for furthering our understanding of the molecular mechanisms of environmental toxicants, as well as for predicting health-related risks due to conditions of environmental exposure and individual susceptibility.

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Epigenetic mechanisms

Epigenetics is the study of heritable changes in gene expression that occur without changes in DNA sequence (Wolffe and Guschin, 2000). Epigenetic mechanisms are flexible genomic parameters that can change genome function under exogenous influence, and also provide a mechanism that allows for the stable propagation of gene activity states from one generation of cells to the next. There are at least two kinds of epigenetic information that can be inherited with chromosomes. The first is DNA methylation, and the second involves changes in chromatin proteins, usually due to modifications in histone tails.

DNA methylation

DNA methylation is a covalent modification, heritable by somatic cells after cell division. 5-Methyl-cytosine represents 2–5% of all cytosines in mammalian genomes and is found primarily on CpG dinucleotides (Millar *et al.*, 2003; Lister *et al.*, 2009; Rossella *et al.*, 2009).

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Cytosine methylation of CpG dinucleotides is found in close proximity to critically important *cis* elements within promoters and is often associated with a repressed chromatin state and inhibition of transcription (Orphanides and Reinberg, 2002). DNA methylation also has an important role in the maintenance of genome integrity by transcriptional silencing of repetitive DNA sequences and endogenous transposons (Bestor, 1998; Hedges and Deinerger, 2007).

Histone modifications

Histones can be modified by acetylation, methylation, phosphorylation, glycosylation, sumoylation and ADP ribosylation (Suganuma and Workman, 2008). The most common modifications are acetylation and methylation of lysine residues in the amino terminal of histone 3 (H3) and histone 4 (H4). Increased acetylation induces transcription activation, whereas decreased acetylation usually induces transcription repression. Methylation of histones is associated with either repression or activation of transcription, depending on the lysine residue position (Yan and Boyd, 2006).

Environmental health and genes: beyond gene–environment interactions

According to the WHO (World Health Organization), more than 13 million deaths annually are due to environmental causes and as much as 24% of disease is

caused by exposures that can be averted (Prüss-Üstün and Corvalán, 2006). The list of environmental threats to human health includes a large number of environmental pollutants. For instance, in the third National Report on Human Exposure to Environmental Chemicals by the Center for Disease Control and Prevention, 148 different environmental chemicals that can be detected in the blood and urine were found in a sample of the US population. The list of pollutants evaluated included metals, phytoestrogens, polycyclic aromatic hydrocarbons, dioxin-like chemicals, polychlorinated biphenyls, phthalates and several classes of pesticides (Department of Health and Human Services Centers for Disease Control and Prevention, 2005).

In Figure 1, we propose a categorization of how environmental exposures, with particular interest in environmental toxic chemicals, may interact with genetic and epigenetic mechanisms. In particular, we are interested in contrasting genetic vs epigenetic mechanisms in their possible interplays with environmental exposures.

Gene–environment interactions

The interplay between the environment and human genome has been traditionally presented under the framework of gene–environment interactions (Figure 1, path A; also indicated as genotype–environment or $G \times E$ interaction) (Ishibe and Kelsey, 1997; Kraft and Hunter, 2005; Dempfle *et al.*, 2008; Baccarelli, 2009; London and Romieu, 2009). Under this model, diseases result from interactions between the individual genetic make-up and environmental factors. Geneticists have always held true that the expression of a genetic trait in the phenotype is highly variable, largely depending on the environment to which the individual carrying the trait of concern is subjected. For instance, in patients with phenylketonuria, which is caused by mutations to a gene coding the liver enzyme phenylalanine hydroxylase, the amino acid phenylalanine does not get converted into tyrosine and reaches high levels in the blood and other tissues

(Scriver, 2007). The elevated phenylalanine levels affect brain development leading to mental retardation. However, a low-phenylalanine diet can keep the blood phenylalanine levels low and avoid the severe effects of phenylketonuria.

The same concept can be approached from the realm of environmental health: some individuals have low risk of developing a disease as a result of an environmental exposure, whereas others are much more susceptible. For example, individuals who carry genetic polymorphisms that make their cells less capable of responding to oxidative stress have been found in several investigations to be more susceptible to the cardiovascular and respiratory effects of air pollution, which produces health effects in humans, at least in part, through oxidative stress generation (Park *et al.*, 2006; Chahine *et al.*, 2007; Baccarelli *et al.*, 2008a).

A purely DNA sequence-based approach (naked DNA snapshot) is not sufficient to fully explain the risks of common diseases, which are modulated by other nongenetic or extragenetic mechanisms. In fact, growing evidence shows that the molecular influences of the environment extend well beyond the interaction with the DNA sequence. Several investigations, as we will discuss in the following sections, have shown that environmental toxicants modify epigenetic states.

Gene–environment vs epigene–environment

In gene–environment interactions (Figure 1, path A), the genetic polymorphisms that modify the effects of environmental exposures are transmitted transgenerationally according to Mendelian genetics, and the trait determining effect modifications is generally assumed to follow the same genetic model (dominant, codominant, recessive) as that of the levels of expression or function of the protein coded by the locus of concern. A second well-established area of interplay (Figure 1, path B) includes the direct effects of environmental exposures on the genome, for example, DNA damage and/or mutations

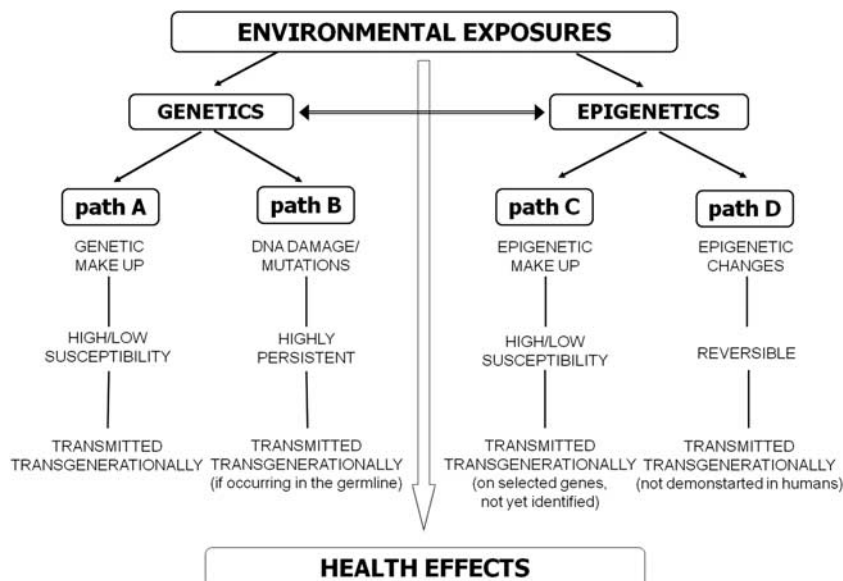


Figure 1 Gene–environment vs epigene–environment interplay: a model of possible genetic and epigenetic paths linking environmental exposures to health effects.

induced by environmental exposures. In environmental health, the recognition that exposures could produce DNA mutations represented a major landmark for risk assessment and prevention. Consequently, genotoxic agents have been categorized according to their capability to alter DNA sequence and thus increase disease risk (Siemiatycki *et al.*, 2004). Such information has been fundamental to determine environmental risks and shape current regulatory efforts for exposure reduction. In particular, potential carcinogenic agents have been carefully tested in *in vitro* and *in vivo* models of mutagenicity. In human subjects, some of these molecular events may represent early events along the pathways linking carcinogen exposure to cancer. For example, in our own study on the population exposed after the Seveso, Italy accident to high doses of dioxin (Pesatori *et al.*, 2008; Baccarelli *et al.*, 2008b), a powerful promoting carcinogen in animals, we showed an increased number of t(14;18) translocations detectable in phenotypically normal blood lymphocytes collected from healthy subjects (Baccarelli *et al.*, 2006). This effect may represent an early expansion of lymphocyte clones potentially related to the increased risk of non-Hodgkin's lymphoma among subjects exposed to high doses of dioxin (Steenland *et al.*, 2004). Environmentally induced DNA mutations can have a transgenerational effect only if occurring in the germ line. For instance, parental exposure to ionizing radiation has been shown to increase the frequency of germline mutations detectable in the next generation (Charles, 2001), and confer a predisposition to cancer (Dubrova *et al.*, 2000).

In principle, the effect-modification model should apply to epigene–environment interactions and to gene–environment interactions. Similar to the effect modifications shown or postulated for genetic polymorphisms (Figure 1, path A), epigenetic differences determining disease risk could make individuals less or more vulnerable to environmental insults (Figure 1, path C). However, to the best of our knowledge, a formal concept of epigene–environment interaction has not yet been developed and we are not aware of examples of epigene–environment interactions in environmental health or toxicology studies. In environmental studies, the flexibility of epigenetic states has generated a growing interest in evaluating the direct alterations that environmental exposures may produce on epigenetic states (Figure 1, path D), including changes in DNA methylation and histone modifications. Investigations that evaluated alterations in DNA methylation and histone modifications in response to environmental chemical exposures were reviewed by us in a recent article (Baccarelli and Bollati, 2009). In this review, we will discuss the biological basis for potential interplays with epigenetic states that might be activated in the presence of environmental exposures and determine health-related effects. We will also discuss whether available evidence suggests that epigenetics provides biological mechanisms for transgenerational environmental effects.

Epigenetic reprogramming in mammalian development

In mammals, DNA methylation is essential for embryogenesis, during which methylation patterns change

dynamically to adapt embryos to be fit for further differentiation (Reik *et al.*, 2001). Two main waves of genome-wide epigenetic reprogramming characterize mammalian development that occurs at the zygote stage and during primordial germ-cell formation (Shi and Wu, 2009).

The genome becomes demethylated during preimplantation to give rise to a totipotent zygote able to generate any cell type. Active DNA demethylation occurring in the paternal genome shortly after fertilization is independent of DNA replication. In contrast, the maternal genome remains highly methylated and undergoes passive DNA demethylation after embryo development (Mayer *et al.*, 2000; Santos *et al.*, 2002). After the first cell cycle, the maternal allele passively loses methylation through cell divisions up to the blastocyst stage (Shi and Wu, 2009). When implantation occurs, DNA methylation levels are then restored by *de novo* methylation that triggers cell lineage differentiation (Sassone-Corsi, 2002).

The second reprogramming event also occurs during embryogenesis, but only in the primordial germ cells in which DNA methylation patterns are erased at all single copy genes and some repetitive elements (Lees-Murdock and Walsh, 2008).

Similar to the pattern of asymmetric DNA methylation in parental genomes, histone H₃K₉ trimethylation and dimethylation exhibit asymmetric modifications in the parental pronuclei (Kurdستاني *et al.*, 2004; Santos *et al.*, 2005; Valls *et al.*, 2005; Wang *et al.*, 2007; Yoshida *et al.*, 2007).

The dilemma of epigenetic inheritance: can epigenetic marks survive reprogramming?

The cycles of erasure/reprogramming that occur during embryogenesis raise the question about how and how frequently epigenetic marks are inherited transgenerationally, and whether epigenetic inheritance occurs in humans and in animal models (Whitelaw and Whitelaw, 2008). Animal experiments provide us with a few examples suggesting that epigenetic marks that are established during the life of an organism can be passed on to the following generations (Probst *et al.*, 2009). These include epigenetic states of murine genes associated with distinctive phenotypes, such as the agouti locus in the viable yellow (A^{vy}/a) mice (determining fur color variation between yellow and dark brown), the Axin-fused (Axin^{fu}) allele (associated with a kinked tail) (Morgan *et al.*, 1999; Rakyán and Whitelaw, 2003) and the Cabp^{IAP} gene, which was identified using a bioinformatic approach and shows sequence homology to the rat CDK5 activator-binding protein (Druker *et al.*, 2004).

All these models derive their properties from metastable epialleles, which are alleles that are variably expressed because of epigenetic modifications that are established very early during development (Rakyán *et al.*, 2002). In metastable epialleles, the epigenetic state can be altered and the alteration shows transgenerational inheritance. Metastable alleles are most often determined by the presence of a retrotransposable element (or retrotransposon). In particular, the murine metastable epialleles A^{vy}, Axin^{fu} and Cabp^{IAP} are all associated with contraoriented insertions of a retrotransposable intracisternal A particle (IAP) sequence, a family of retro-

virus-like genetic elements coding for virus-like particles (Duhl *et al.*, 1994; Vasicek *et al.*, 1997; Druker *et al.*, 2004).

Epigenetic inheritance and metastable alleles in animal experiments: the viable yellow agouti A^{vy} mouse

The viable yellow agouti A^{vy} allele is the most extensively studied murine metastable epiallele. In viable yellow (A^{vy}/a) mice, transcription originating in an IAP retrotransposon inserted upstream of the agouti gene (A) causes ectopic expression of agouti protein, resulting in yellow fur, obesity, diabetes and increased susceptibility to tumors (Morgan *et al.*, 1999). A^{vy} mice show variable expressivity because they are epigenetic mosaics for activity of the retrotransposon: isogenic A^{vy} mice have coats that vary in a continuous spectrum from full yellow, through variegated yellow/agouti, to full agouti (pseudoagouti). The distribution of phenotypes among offspring is related to the phenotype of the dam; when an A^{vy} dam has the agouti phenotype, her offspring is more likely to be agouti. It has been shown that the offspring color is not the result of a maternally contributed environment, but rather genuinely inherited, and is associated with transmission of a silenced A^{vy} allele through the female germ line (Morgan *et al.*, 1999).

Is there enough evidence for epigenetic inheritance in humans?

Determining whether equivalents of metastable epialleles exist and are frequent in humans poses substantial challenges. There are just a few reports that have been used to suggest inheritance of epigenetic states in humans. For example, in several cases of familial colorectal cancer, the mismatch repair genes MLH1 and MSH2, which usually exhibit low or no methylation, have been found to be silenced because of promoter methylation, and this has been occasionally detected in successive generations (Suter *et al.*, 2004; Chan *et al.*, 2006; Hitchins *et al.*, 2007). However, these were reports of single families and it has been argued that the promoter methylation identified, even if detected in multiple family members, could be explained by somatic events that occurred after fertilization (Horsthemke, 2007). Whitelaw and Whitelaw (2008) have recently remarked that, in the light of current evidence, the notion that epigenetic marks can be directly inherited across generations in humans remains contentious.

Environmental influences on epigenetic states during early development in animal models

Modifications to the environment during early development can lead to permanent changes in the pattern of epigenetic modifications (Fauque *et al.*, 2007). Pregnant female rats exposed during time of sex determination to the endocrine disruptor vinclozolin have been shown to exhibit in their male offspring transgenerational disease state leading to spermatogenic defects, prostate disease, kidney disease, immune system abnormalities, hypercholesterolemia and an increased rate of tumor

development in the F1–F4 generation offspring (Anway *et al.*, 2005, 2006a,b; Anway and Skinner, 2006; Chang *et al.*, 2006). Both the F1 generation embryo and F2 generation germ line are directly exposed when an F0 generation pregnant mother is exposed. Therefore, only the F3 generation can provide the first unequivocal signs of transgenerational inheritance. The transgenerational disease states in the vinclozolin F1–F4 generation animals were found to be associated with a transgenerational alteration in the epigenetic programming of the male germ line (Anway *et al.*, 2005).

A second example of an epigenetic toxicant is represented by bisphenol A (BPA), a high-production-volume chemical used in the manufacture of polycarbonate plastic. *In utero* or neonatal exposure to BPA is associated with higher body weight, increased breast and prostate cancer, as well as altered reproductive function. Dolinoy *et al.* (2007a) showed that maternal BPA exposure shifted the coat color distribution of viable yellow agouti (A^{vy}) mouse offspring toward yellow by decreasing CpG (cytosine-guanine dinucleotide) methylation in the IAP sequence upstream of the Agouti gene (Waterland, 2009). In addition, CpG methylation was also decreased at the Cabp^{IAP} metastable locus. DNA methylation at the A^{vy} locus was similar in tissues from the three germ layers, suggesting that BPA affected epigenetic patterning during early stem-cell development. Moreover, maternal dietary supplementation with either methyl donors folic acid or the phytoestrogen genistein blunted the DNA-hypomethylating effect of BPA (Dolinoy *et al.*, 2007a). However, as this investigation was conducted only up to the F2 generation, it did not directly show inheritance of the epigenetic modifications induced by BPA exposure.

Translating environmental epigenetic effects from animal models to humans

Translating results obtained in mouse models to humans is not straightforward. The agouti model cannot be directly applied to humans, and neither is there a human equivalent of kinked tails. One common characteristic of the mouse models described above is that their metastable alleles all include an IAP retrotransposon. IAP sequences are endogenous retrovirus-like mobile elements, present at 1000 copies in the mouse genome. These elements transpose in a replicative manner through an RNA intermediate and its reverse transcription, and their transposition should therefore be tightly controlled by their transcription level (Dupressoir and Heidmann, 1997). In humans, more than one-third of DNA methylation occurs in retrotransposons (Kochanek *et al.*, 1993; Schmid, 1998), which represent a large portion of the human genome (Bernstein *et al.*, 2006). Among these sequences, Alu and LINE-1 retrotransposons are the most plentiful families representing ~30% of the human genome (Kazazian and Goodier, 2002; Grover *et al.*, 2004; Babushok and Kazazian, 2007) and are heavily methylated (Yang *et al.*, 2004). Owing to their high representation throughout the genome, Alu and LINE-1 have been proposed as surrogate markers for estimating the global DNA methylation level (Yang *et al.*, 2004; Weisenberger *et al.*, 2005), but growing evidence indicates that they could have specific and distinct

cellular roles (Wallace *et al.*, 2008). Hypomethylation of repetitive elements favors their activity as retrotransposable sequences and has been suggested to have deleterious effects on cells, initially through insertional mutations (Kazazian, 2004), and later by introducing genome instability through deletions and genomic rearrangements (Ostertag and Kazazian, 2001; Gilbert *et al.*, 2002; Wallace *et al.*, 2008).

In healthy populations, inter-individual methylation variations at Alu or LINE-1 elements from blood DNA have also been associated with risk factors for cancer, neurological and cardiovascular diseases (Bollati *et al.*, 2007; Rusiecki *et al.*, 2008; Baccarelli and Bollati, 2009; Baccarelli *et al.*, 2009). In addition, the finding of an age-associated decline in repetitive-element methylation in normal tissues of aging individuals suggests a possible role in various common human age-related diseases (Bollati *et al.*, 2009).

Epigenetics and environmental toxicants in humans

Exposure to air pollution, particularly to particulate matter (PM), has been associated with increased morbidity and mortality from cardiorespiratory disease, as well as with lung cancer risk (Samet *et al.*, 2000; Brook *et al.*, 2004; Peters, 2005; Vineis and Husgafvel-Pursiainen, 2005; Baccarelli *et al.*, 2008c). In a human study, we recently showed that promoter methylation of the iNOS (inducible Nitric Oxide Synthase) gene was lower in blood samples obtained from foundry workers with well-characterized exposure to PM with aerodynamic diameter $<10\mu\text{m}$ (PM₁₀) (Tarantini *et al.*, 2008). iNOS demethylation is expected to increase the expression and activity of the iNOS protein, which could in turn contribute to inflammation and oxidative stress generation, which are primary mechanisms linking inhalation of air pollutants to their acute health effects (Baccarelli *et al.*, 2007; Chahine *et al.*, 2007; Alexeeff *et al.*, 2008). In the same study, long-term exposure to PM₁₀ was negatively associated with methylation in both Alu and LINE-1 (Tarantini *et al.*, 2008). In a recent investigation, we showed that exposure to black carbon, a marker of particles from vehicular traffic, was also associated with decreased DNA methylation in LINE-1, measured in 1097 blood DNA samples obtained from the NAS (Normative Aging Study), an investigation of elderly men in the Boston area. As repetitive-element hypomethylation is believed to occur in patients with cancer (Ehrlich, 2002) and cardiovascular disease (Castro *et al.*, 2003); such changes may reproduce epigenetic processes related to disease development and represent mechanisms by which particulate air pollution affects human health (Baccarelli *et al.*, 2008c). Several *in vitro* studies have established an association between DNA methylation and environmental metals, which are components of PM, including nickel, cadmium, lead and particularly arsenic (McVeigh *et al.*, 2001; Bleich *et al.*, 2006; Wright and Baccarelli, 2007; Dolinoy *et al.*, 2007b).

In addition, in an animal study, Yauk *et al.* (2008) showed that sperm DNA of mice exposed to steel plant air was hypermethylated compared with control animals and this change persisted even after removal from environmental exposure. This finding calls for further

research to determine whether air pollutants produce DNA methylation changes that are transmitted transgenerationally.

In our laboratory, we also investigated whether DNA methylation changes are induced by low-benzene exposure in peripheral blood DNA of gasoline station attendants and traffic police officers. High-level exposure to benzene has been associated with increased risk of acute myelogenous leukemia (Snyder, 2002). In our study, airborne benzene exposure was associated with a significant reduction in LINE-1 and Alu methylation. Airborne benzene was also associated with hypermethylation in p15 and hypomethylation of the MAGE-1 cancer-antigen gene (Bollati *et al.*, 2007). These findings show that low-level benzene exposure may induce altered DNA methylation reproducing the aberrant epigenetic patterns found in malignant cells, which in most reports have been found to exhibit repetitive-element hypomethylation as well as either hypermethylation or hypomethylation of specific genes, depending on the gene function. In addition, benzene-associated demethylation of repetitive elements may help explain the epidemiological data linking benzene exposure to increased risk of multiple myeloma (Costantini *et al.*, 2008; Kirkeleit *et al.*, 2008), which also exhibits reduced methylation in Alu and LINE-1 repetitive elements (Bollati *et al.*, 2007).

Other exposures that are associated with increased risk of hematopoietic malignancies, such as persistent organic pollutants, have been associated with changes in repetitive-element DNA methylation. Rusiecki *et al.* (2008) evaluated the relationship between plasma persistent organic pollutant concentrations and blood global DNA methylation, estimated in Alu repeated elements, and in 70 Greenlandic Inuit, a population presenting some of the highest reported levels of persistent organic pollutants worldwide. In this study, a significant inverse linear relationship was found for DDT, DDE, β -BHC, oxychlordane, α -chlordane, mirex, several PCBs and the sum of all persistent organic pollutants (Rusiecki *et al.*, 2008). As most of the exposures investigated increase oxidative stress in human tissues, it is possible that the production of reactive oxygen species represents a unifying process to account for most of these findings across different chemicals (Wright and Baccarelli, 2007; Baccarelli and Bollati, 2009). Oxidative DNA damage can interfere with the ability of methyltransferases to interact with DNA (Valinluck *et al.*, 2004), thus resulting in a generalized altered methylation of cytosine residues at CpG sites (Turker and Bestor, 1997).

Final remarks

In the last few years, several investigations have examined the relationship between exposure to environmental chemicals and epigenetics, and identified several toxicants that modify epigenetic marks. Most of the studies conducted so far have been focused on DNA methylation, whereas only a few recent investigations have studied the effects of environmental chemicals on histone modifications (Baccarelli and Bollati, 2009). In animal models, environmental effects that might involve epigenetic mechanisms have been shown to be transmitted transgenerationally. Whether the transgenerational epigenetic effects of environmental exposures are

present in humans, or whether epigenetic inheritance exists at all in humans, remains to be elucidated.

Nonetheless, growing evidence indicates that epigenetics holds substantial potential for developing biological markers to predict which exposures would put exposed subjects at risk and which individuals will be more susceptible to develop disease. In human studies, this will require the use of laboratory methods with enhanced precision, sensitivity and coverage, so that epigenetic changes can be detected as early as possible and well ahead of disease diagnosis. For several exposures, it has been proven that chemicals can alter epigenetic marks and that the same or similar epigenetic alterations can be found in patients with the disease of concern and/or in diseased tissues. Future prospective investigations are required to determine whether exposed subjects develop epigenetic alterations over time and, in turn, whether such alterations increase the risk of disease.

Conflict of interest

The authors declare no conflict of interest.

References

- Alexeeff SE, Litonjua AA, Wright RO, Baccarelli A, Suh H, Sparrow D *et al.* (2008). Ozone exposure, antioxidant genes, and lung function in an elderly cohort: VA normative aging study. *Occup Environ Med* **65**: 736–742.
- Anway MD, Cupp AS, Uzumcu M, Skinner MK (2005). Epigenetic transgenerational actions of endocrine disruptors and male fertility. *Science* **308**: 1466–1469.
- Anway MD, Leathers C, Skinner MK (2006a). Endocrine disruptor vinclozolin induced epigenetic transgenerational adult-onset disease. *Endocrinology* **147**: 5515–5523.
- Anway MD, Memon MA, Uzumcu M, Skinner MK (2006b). Transgenerational effect of the endocrine disruptor vinclozolin on male spermatogenesis. *J Androl* **27**: 868–879.
- Anway MD, Skinner MK (2006). Epigenetic transgenerational actions of endocrine disruptors. *Endocrinology* **147**(6 Suppl): S43–S49.
- Babushok DV, Kazazian Jr HH (2007). Progress in understanding the biology of the human mutagen LINE-1. *Hum Mutat* **28**: 527–539.
- Baccarelli A (2009). Breathe deeply into your genes!—Genetic variants and air pollution effects (Editorial). *Am J Respir Crit Care Med* **179**: 431–432.
- Baccarelli A, Bollati V (2009). Epigenetics and environmental chemicals. *Curr Opin Pediatr* **21**: 243–251.
- Baccarelli A, Cassano PA, Litonjua A, Park SK, Suh H, Sparrow D *et al.* (2008a). Cardiac autonomic dysfunction: effects from particulate air pollution and protection by dietary methyl nutrients and metabolic polymorphisms. *Circulation* **117**: 1802–1809.
- Baccarelli A, Giacomini SM, Corbetta C, Landi MT, Bonzini M, Consonni D *et al.* (2008b). Neonatal thyroid function in Seveso 25 years after maternal exposure to dioxin. *PLoS Med* **5**: e161.
- Baccarelli A, Hirt C, Pesatori AC, Consonni D, Patterson Jr DG, Bertazzi PA *et al.* (2006). t(14;18) translocations in lymphocytes of healthy dioxin-exposed individuals from Seveso, Italy. *Carcinogenesis* **27**: 2001–2007.
- Baccarelli A, Martinelli I, Zanobetti A, Grillo P, Hou LF, Bertazzi PA *et al.* (2008c). Exposure to particulate air pollution and risk of deep vein thrombosis. *Arch Intern Med* **168**: 920–927.
- Baccarelli A, Wright RO, Bollati V, Tarantini L, Litonjua AA, Suh HH *et al.* (2009). Rapid DNA methylation changes after exposure to traffic particles. *Am J Respir Crit Care Med* **179**: 572–578.
- Baccarelli A, Zanobetti A, Martinelli I, Grillo P, Hou L, Lanzani G *et al.* (2007). Air pollution, smoking, and plasma homocysteine. *Environ Health Perspect* **115**: 176–181.
- Bernstein BE, Mikkelsen TS, Xie X, Kamal M, Huebert DJ, Cuff J *et al.* (2006). A bivalent chromatin structure marks key developmental genes in embryonic stem cells. *Cell* **125**: 315–326.
- Bestor TH (1998). The host defence function of genomic methylation patterns. *Novartis Found Symp* **214**: 187–195; discussion 195–189, 228–132.
- Bleich S, Lenz B, Ziegenbein M, Beutler S, Frieling H, Kornhuber J *et al.* (2006). Epigenetic DNA hypermethylation of the HEP gene promoter induces down-regulation of its mRNA expression in patients with alcohol dependence. *Alcohol Clin Exp Res* **30**: 587–591.
- Bollati V, Baccarelli A, Hou L, Bonzini M, Fustinoni S, Cavallo D *et al.* (2007). Changes in DNA methylation patterns in subjects exposed to low-dose benzene. *Cancer Res* **67**: 876–880.
- Bollati V, Schwartz J, Wright R, Litonjua A, Tarantini L, Suh H *et al.* (2009). Decline in genomic DNA methylation through aging in a cohort of elderly subjects. *Mech Ageing Dev* **130**: 234–239.
- Brook RD, Franklin B, Cascio W, Hong Y, Howard G, Lipsett M *et al.* (2004). Air pollution and cardiovascular disease: a statement for healthcare professionals from the Expert Panel on Population and Prevention Science of the American Heart Association. *Circulation* **109**: 2655–2671.
- Castro R, Rivera I, Ravasco P, Jakobs C, Blom HJ, Camilo ME *et al.* (2003). 5,10-Methylenetetrahydrofolate reductase 677C→T and 1298A→C mutations are genetic determinants of elevated homocysteine. *QJM* **96**: 297–303.
- Chahine T, Baccarelli A, Litonjua A, Wright RO, Suh H, Gold DR *et al.* (2007). Particulate air pollution, oxidative stress genes, and heart rate variability in an elderly cohort. *Environ Health Perspect* **115**: 1617–1622.
- Chan TL, Yuen ST, Kong CK, Chan YW, Chan AS, Ng WF *et al.* (2006). Heritable germline epimutation of MSH2 in a family with hereditary nonpolyposis colorectal cancer. *Nat Genet* **38**: 1178–1183.
- Chang HS, Anway MD, Rekow SS, Skinner MK (2006). Transgenerational epigenetic imprinting of the male germline by endocrine disruptor exposure during gonadal sex determination. *Endocrinology* **147**: 5524–5541.
- Charles M (2001). UNSCEAR report 2000: sources and effects of ionizing radiation. United Nations Scientific Committee on the Effects of Atomic Radiation. *J Radiol Prot* **21**: 83–86.
- Costantini AS, Benvenuti A, Vineis P, Kriebel D, Tumino R, Ramazzotti V *et al.* (2008). Risk of leukemia and multiple myeloma associated with exposure to benzene and other organic solvents: evidence from the Italian Multicenter Case-Control Study. *Am J Ind Med* **51**: 803–811.
- Dempfle A, Scherag A, Hein R, Beckmann L, Chang-Claude J, Schafer H (2008). Gene-environment interactions for complex traits: definitions, methodological requirements and challenges. *Eur J Hum Genet* **16**: 1164–1172.
- Department of Health and Human Services Centers for Disease Control and Prevention (2005). Third National Report on Human Exposure to Environmental Chemicals. National Center for Environmental Health, Atlanta, GA.
- Dolinoy DC, Huang D, Jirtle RL (2007a). Maternal nutrient supplementation counteracts bisphenol A-induced DNA hypomethylation in early development. *Proc Natl Acad Sci USA* **104**: 13056–13061.
- Dolinoy DC, Weidman JR, Jirtle RL (2007b). Epigenetic gene regulation: linking early developmental environment to adult disease. *Reprod Toxicol* **23**: 297–307.
- Druker R, Bruxner TJ, Lehrbach NJ, Whitelaw E (2004). Complex patterns of transcription at the insertion site

- of a retrotransposon in the mouse. *Nucleic Acids Res* **32**: 5800–5808.
- Dubrova YE, Plumb M, Gutierrez B, Boulton E, Jeffreys AJ (2000). Transgenerational mutation by radiation. *Nature* **405**: 37.
- Duhl DM, Vrieling H, Miller KA, Wolff GL, Barsh GS (1994). Neomorphic agouti mutations in obese yellow mice. *Nat Genet* **8**: 59–65.
- Dupressoir A, Heidmann T (1997). Expression of intracisternal A-particle retrotransposons in primary tumors of oncogene-expressing transgenic mice. *Oncogene* **14**: 2951–2958.
- Ehrlich M (2002). DNA methylation in cancer: too much, but also too little. *Oncogene* **21**: 5400–5413.
- Fauque P, Jouannet P, Lesaffre C, Ripoche MA, Dandolo L, Vaiman D *et al.* (2007). Assisted reproductive technology affects developmental kinetics, H19 imprinting control region methylation and H19 gene expression in individual mouse embryos. *BMC Dev Biol* **7**: 116.
- Gilbert N, Lutz-Prigge S, Moran JV (2002). Genomic deletions created upon LINE-1 retrotransposition. *Cell* **110**: 315–325.
- Grover D, Mukerji M, Bhatnagar P, Kannan K, Brahmachari SK (2004). Alu repeat analysis in the complete human genome: trends and variations with respect to genomic composition. *Bioinformatics* **20**: 813–817.
- Hedges DJ, Deininger PL (2007). Inviting instability: transposable elements, double-strand breaks, and the maintenance of genome integrity. *Mutat Res* **616**: 46–59.
- Hitchins MP, Wong JJ, Suthers G, Suter CM, Martin DI, Hawkins NJ *et al.* (2007). Inheritance of a cancer-associated MLH1 germ-line epimutation. *N Engl J Med* **356**: 697–705.
- Horsthemke B (2007). Rhythm is not enough. *Nat Genet* **39**: 1190–1191.
- Ishibe N, Kelsey KT (1997). Genetic susceptibility to environmental and occupational cancers. *Cancer Causes Control* **8**: 504–513.
- Kazazian Jr HH (2004). Mobile elements: drivers of genome evolution. *Science* **303**: 1626–1632.
- Kazazian Jr HH, Goodier JL (2002). LINE drive, retrotransposition and genome instability. *Cell* **110**: 277–280.
- Kirkeleit J, Riise T, Bratveit M, Moen BE (2008). Increased risk of acute myelogenous leukemia and multiple myeloma in a historical cohort of upstream petroleum workers exposed to crude oil. *Cancer Causes Control* **19**: 13–23.
- Kochanek S, Renz D, Doerfler W (1993). DNA methylation in the Alu sequences of diploid and haploid primary human cells. *EMBO J* **12**: 1141–1151.
- Kraft P, Hunter D (2005). Integrating epidemiology and genetic association: the challenge of gene-environment interaction. *Philos Trans R Soc Lond B Biol Sci* **360**: 1609–1616.
- Kurdistani SK, Tavazoie S, Grunstein M (2004). Mapping global histone acetylation patterns to gene expression. *Cell* **117**: 721–733.
- Lees-Murdock DJ, Walsh CP (2008). DNA methylation reprogramming in the germ line. *Epigenetics* **3**: 5–13.
- Lister R, Pelizzola M, Downen RH, Hawkins RD, Hon G, Tonti-Filippini J *et al.* (2009). Human DNA methylomes at base resolution show widespread epigenomic differences. *Nature* **462**: 315–322.
- London SJ, Romieu I (2009). Gene by environment interaction in asthma. *Annu Rev Public Health* **30**: 55–80.
- Mayer W, Niveleau A, Walter J, Fundele R, Haaf T (2000). Demethylation of the zygotic paternal genome. *Nature* **403**: 501–502.
- McVeigh GE, Allen PB, Morgan DR, Hanratty CG, Silke B (2001). Nitric oxide modulation of blood vessel tone identified by arterial waveform analysis. *Clin Sci (Lond)* **100**: 387–393.
- Millar D, Holliday R, Grigg G (2003). Five not four: history and significance of the fifth base. In: Beck S, Olek A (eds). *The Epigenome, Molecular Hide and Seek*. Wiley-VCH Verlag GmbH Co. KGaA: Weinheim, pp 3–20.
- Morgan HD, Sutherland HG, Martin DI, Whitelaw E (1999). Epigenetic inheritance at the agouti locus in the mouse. *Nat Genet* **23**: 314–318.
- Orphanides G, Reinberg D (2002). A unified theory of gene expression. *Cell* **108**: 439–451.
- Ostertag EM, Kazazian Jr HH (2001). Biology of mammalian L1 retrotransposons. *Annu Rev Genet* **35**: 501–538.
- Park SK, O'Neill MS, Wright RO, Hu H, Vokonas PS, Sparrow D *et al.* (2006). HFE genotype, particulate air pollution, and heart rate variability: a gene-environment interaction. *Circulation* **114**: 2798–2805.
- Pesatori AC, Baccarelli A, Consonni D, Lania A, Beck-Peccoz P, Bertazzi PA *et al.* (2008). Aryl hydrocarbon receptor-interacting protein and pituitary adenomas: a population-based study on subjects exposed to dioxin after the Seveso, Italy, accident. *Eur J Endocrinol* **159**: 699–703.
- Peters A (2005). Particulate matter and heart disease: evidence from epidemiological studies. *Toxicol Appl Pharmacol* **207** (2 Suppl): 477–482.
- Probst AV, Dunleavy E, Almouzni G (2009). Epigenetic inheritance during the cell cycle. *Nat Rev Mol Cell Biol* **10**: 192–206.
- Prüss-Ustün A, Corvalán C (2006). Preventing disease through healthy environments. Towards an estimate of the environmental burden of disease. Geneva, World Health Organization (WHO).
- Rakyan V, Whitelaw E (2003). Transgenerational epigenetic inheritance. *Curr Biol* **13**: R6.
- Rakyan VK, Blewitt ME, Druker R, Preis JL, Whitelaw E (2002). Metastable epialleles in mammals. *Trends Genet* **18**: 348–351.
- Reik W, Dean W, Walter J (2001). Epigenetic reprogramming in mammalian development. *Science* **293**: 1089–1093.
- Rossella F, Polledri E, Bollati V, Baccarelli A, Fustinoni S (2009). Development and validation of a gas chromatography/mass spectrometry method for the assessment of genomic DNA methylation. *Rapid Commun Mass Spectrom* **23**: 2637–2646.
- Rusiecki JA, Baccarelli A, Bollati V, Tarantini L, Moore LE, Bonfeld-Jorgensen EC (2008). Global DNA hypomethylation is associated with high serum-persistent organic pollutants in Greenlandic Inuit. *Environ Health Perspect* **116**: 1547–1552.
- Samet JM, Dominici F, Currier FC, Coursac I, Zeger SL (2000). Fine particulate air pollution and mortality in 20 U.S. cities, 1987–1994. *N Engl J Med* **343**: 1742–1749.
- Santos F, Hendrich B, Reik W, Dean W (2002). Dynamic reprogramming of DNA methylation in the early mouse embryo. *Dev Biol* **241**: 172–182.
- Santos F, Peters AH, Otte AP, Reik W, Dean W (2005). Dynamic chromatin modifications characterise the first cell cycle in mouse embryos. *Dev Biol* **280**: 225–236.
- Sassone-Corsi P (2002). Unique chromatin remodeling and transcriptional regulation in spermatogenesis. *Science* **296**: 2176–2178.
- Schmid CW (1998). Does SINE evolution preclude Alu function? *Nucleic Acids Res* **26**: 4541–4550.
- Scriver CR (2007). The PAH gene, phenylketonuria, and a paradigm shift. *Hum Mutat* **28**: 831–845.
- Shi L, Wu J (2009). Epigenetic regulation in mammalian preimplantation embryo development. *Reprod Biol Endocrinol* **7**: 59.
- Siemiatycki J, Richardson L, Straif K, Latreille B, Lakhani R, Campbell S *et al.* (2004). Listing occupational carcinogens. *Environ Health Perspect* **112**: 1447–1459.
- Snyder R (2002). Benzene and leukemia. *Crit Rev Toxicol* **32**: 155–210.
- Steenland K, Bertazzi P, Baccarelli A, Kogevinas M (2004). Dioxin revisited: developments since the 1997 IARC classification of dioxin as a human carcinogen. *Environ Health Perspect* **112**: 1265–1268.
- Suganuma T, Workman JL (2008). Crosstalk among histone modifications. *Cell* **135**: 604–607.

- Suter CM, Martin DI, Ward RL (2004). Germline epimutation of MLH1 in individuals with multiple cancers. *Nat Genet* **36**: 497–501.
- Tarantini L, Bonzini M, Apostoli P, Pegoraro V, Bollati V, Marinelli B *et al.* (2008). Effects of particulate matter on genomic DNA methylation content and iNOS promoter methylation. *Environ Health Perspect* (in press; doi:10.1289/ehp.11898) **117**: 217–222.
- Turker MS, Bestor TH (1997). Formation of methylation patterns in the mammalian genome. *Mutat Res* **386**: 119–130.
- Valinluck V, Tsai HH, Rogstad DK, Burdzy A, Bird A, Sowers LC (2004). Oxidative damage to methyl-CpG sequences inhibits the binding of the methyl-CpG binding domain (MBD) of methyl-CpG binding protein 2 (MeCP2). *Nucleic Acids Res* **32**: 4100–4108.
- Valls E, Sanchez-Molina S, Martinez-Balbas MA (2005). Role of histone modifications in marking and activating genes through mitosis. *J Biol Chem* **280**: 42592–42600.
- Vasicek TJ, Zeng L, Guan XJ, Zhang T, Costantini F, Tilghman SM (1997). Two dominant mutations in the mouse fused gene are the result of transposon insertions. *Genetics* **147**: 777–786.
- Vineis P, Husgafvel-Pursiainen K (2005). Air pollution and cancer: biomarker studies in human populations. *Carcinogenesis* **26**: 1846–1855.
- Wallace NA, Belancio VP, Deininger PL (2008). L1 mobile element expression causes multiple types of toxicity. *Gene* **419**: 75–81.
- Wang F, Kou Z, Zhang Y, Gao S (2007). Dynamic reprogramming of histone acetylation and methylation in the first cell cycle of cloned mouse embryos. *Biol Reprod* **77**: 1007–1016.
- Waterland RA (2009). Is epigenetics an important link between early life events and adult disease? *Horm Res* **71**(Suppl 1): 13–16.
- Weisenberger DJ, Campan M, Long TI, Kim M, Woods C, Fiala E *et al.* (2005). Analysis of repetitive element DNA methylation by MethyLight. *Nucleic Acids Res* **33**: 6823–6836.
- Whitelaw NC, Whitelaw E (2008). Transgenerational epigenetic inheritance in health and disease. *Curr Opin Genet Dev* **18**: 273–279.
- Wolffe AP, Guschin D (2000). Review: chromatin structural features and targets that regulate transcription. *J Struct Biol* **129**: 102–122.
- Wright RO, Baccarelli A (2007). Metals and neurotoxicology. *J Nutr* **137**: 2809–2813.
- Yan C, Boyd DD (2006). Histone H3 acetylation and H3 K4 methylation define distinct chromatin regions permissive for transgene expression. *Mol Cell Biol* **26**: 6357–6371.
- Yang AS, Estecio MR, Doshi K, Kondo Y, Tajara EH, Issa JP (2004). A simple method for estimating global DNA methylation using bisulfite PCR of repetitive DNA elements. *Nucleic Acids Res* **32**: e38.
- Yauk C, Polyzos A, Rowan-Carroll A, Somers CM, Godschalk RW, Van Schooten FJ *et al.* (2008). Germ-line mutations, DNA damage, and global hypermethylation in mice exposed to particulate air pollution in an urban/industrial location. *Proc Natl Acad Sci USA* **105**: 605–610.
- Yoshida N, Brahmajoshiyula M, Shoji S, Amanai M, Perry AC (2007). Epigenetic discrimination by mouse metaphase II oocytes mediates asymmetric chromatin remodeling independently of meiotic exit. *Dev Biol* **301**: 464–477.