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IN REPLY REFER TO:

FISH AND WILDLIFE SERVICE
BUREAU OF SPORT FISHERIES AND WILDLIFE

South Dakota Cooperative
Wildlife Research Unit
South Dakota State University
Brookings, South Dakota 57006

January 8, 1970

Mr. Jack T. Garrett
Manager, Industrial Hygiene
Monsanto Company
800 N. Lindbergh Boulevard
St. Louis, Missouri 63166

Dear Mr. Garrett:

In the past you have kindly supplied us with samples of Aroclor 1254. I'm sending a copy of a manuscript accepted for the next April issue of The Journal of Wildlife Management. It contains some of the results of our research on the effect of this compound on the ring-necked pheasant. As other results become available, I'll pass them along to you.

Sincerely,

Robert B. Dahlgren
Assistant Leader

RBD:cs

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STLCOPCB4052753

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EFFECTS OF POLYCHLORINATED BIPHENYLS ON PHEASANT REPRODUCTION, BEHAVIOR, AND SURVIVAL

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Abstract: Cock pheasants (Phasianus colchicus) were given single capsules weekly containing either 0 or 25 mg polychlorinated biphenyls (Aroclor 1254) for 17 weeks in the spring of 1970. Hen pheasants mated to these cock groups were given single capsules weekly containing 0, 12.5, or 50 mg PCB. Among groups receiving PCB, egg production was depressed significantly ($P<0.05$) and the number of eggs which were pipped but not hatched was increased significantly ($P<0.05$). Even though weekly weight changes of hens, fertility and hatchability of eggs, mortality of adults, and eggshell thickness did not appear to be affected by PCB, total reproductive success was depressed. Behavior, on a visual cliff, of chicks hatched from hens given 50 mg weekly was significantly different ($P<0.01$); more of them jumped to the visually deep side or made no choice of sides. Mortality of chicks from hatching to 6 weeks of age was significantly greater ($P<0.01$) in the offspring of hens receiving 50 mg PCB weekly, and surviving chicks of this group were significantly lighter ($P<0.05$) in weight than other groups at 6 weeks of age.

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[1] Supported jointly by the South Dakota Department of Game, Fish and Parks, Bureau of Sport Fisheries and Wildlife, South Dakota State University and The Wildlife Management Institute.

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Polychlorinated biphenyls (PCB) have been detected in wildlife in Europe (Jensen 1966, Holden and Marsden 1967, Holmes et al. 1967, Jensen et al. 1969, Koeman et al. 1969) and in the United States (Risebrough et al. 1968, Anderson et al. 1969, Bagley et al. 1970, Risebrough 1970). This chlorinated hydrocarbon was found as a residue in tissues of both sea birds and land-dwelling species.

The induction of steroid enzymes in the liver by PCB has been demonstrated in both laboratory rats (Street et al. 1969) and in pigeons (Risebrough et al. 1968), and since the capacity of PCB to induce these enzymes is as potent as or more potent than DDT, they have been implicated in eggshell thinning. Sax (1963) indicated that the liver of man could be injured by PCB; liver damage has been shown for guinea pigs, laboratory rats and domestic rabbits by Miller (1944). McCune et al. (1962) and Flick et al. (1965) found that PCB fed to young chickens produced edema and pathological changes in internal organs. The objective of our study was to evaluate the effects of PCB on reproduction, survival and behavior of pheasants.

We are grateful to W. L. Tucker, Statistician, Agricultural Experiment Station, South Dakota State University, for statistical advice. This study was conducted using funds supplied by the Bureau of Sport Fisheries and Wildlife.

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METHODS

We used Aroclor 1254 supplied by the Monsanto Co., St. Louis, Missouri. Hens were purchased in the winter of 1970 from the South Dakota Pheasant Co., Canton, and cocks were raised by us from birds previously obtained from the same source. Two groups of five cocks each were given weekly either a 00 size gelatin capsule containing corn oil or one containing corn oil in 9:1 dilution with 25 mg PCB. Each cock group was mated with three groups of five hens each given weekly a single corn oil placebo capsule, 12.5 mg PCB, or 50 mg PCB. Hens and cocks were maintained in separate cages. Cocks were dosed for 17 weeks from February 13 to June 5 and hens from February 20 to June 12.

General care, diet, and maintenance was the same as that described for a dieldrin experiment by Baxter et al. (1969), except that hens in this study were given a tablespoonful of oyster shells in their food cups once each week.

Eggs were gathered from March 7 to June 19 and set weekly in a forced draft incubator. Within 36 hours of hatching, chicks were taken from the incubator, placed directly on a visual cliff (Tallarico and Farrell 1964), and given 5 minutes to jump to the visually deep or shallow side. Chicks from this study and those hatched from a concurrent dieldrin reproduction study were placed together in brooders until they were 6 weeks of age, at which time each bird was weighed to the nearest gram.

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Adults were weighed weekly to the nearest gram. Eggshells of hatched eggs from all 15 weekly egg groups were measured, without the membrane, to the nearest 0.01 mm using a micrometer at three points around the midline perpendicular to the long axis, as described by Dahlgren and Linder (1970). Unhatched eggs from weekly egg groups 8-15 were broken, dried for 24 hours, and eggshells including the membrane were measured.

Fertility of unhatched eggs was based on a visual inspection for evidence of embryonic development.

RESULTS AND DISCUSSION

Effects of PCB on Condition and Reproduction

Only 2 hens died among the 30 hens and 10 cocks under study; both of the hens were receiving 50 mg PCB weekly. Analysis of weekly changes of weight of cocks and hens tested by analysis of variance showed no effect ($P>0.05$) from the administration of PCB. Flick et al. (1965), who fed 200 and 400 ppm and McCune et al. (1962), who fed at a dietary level of 100-800 ppm, found that the growth of chicks of the domestic chicken was retarded by Aroclor 1242.

The averages of eggs laid per hen per day (Table 1) among hen treatment groups were significantly different. Using an orthogonal test, rate of egg production among hens given 12.5 mg PCB weekly was not different ($P>0.05$) than the control group, but the 50 mg group was different ($P<0.01$) from the other groups.

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M. A. Haegele at the Denver Wildlife Research Center (personal communication) found that a single oral dose of 500 mg/kg of Aroclor 1254 given to Japanese quail (Coturnix coturnix) reduced egg production about 50 percent for 7 days post-treatment, but egg production was near normal 8-9 days after treatment.

The differences among treatment groups in egg fertility were significant ($P<0.05$); however, a significant ($P<0.01$) cock-hen interaction showed these results to be of no biological meaning.

Hatchability was highest in control groups and lowest in groups in which hens had received 50 mg PCB, but the differences tested by chi square were not significant ($P>0.05$). Significant differences ($P<0.05$) in the numbers of eggs which were pipped but not hatched were found between groups. Apparently the administration of PCB adversely affected the viability of the embryo at the time of hatching. McLaughlin et al. (1963) injected both 10 and 25 mg of Aroclor 1242 into chicken eggs and found only 0-5 percent hatchability, growth retardation, edema, and beak deformities in embryos.

No significant differences ($P>0.05$) in eggshell thickness using analysis of variance were noted among treatment groups (Table 2). Eggshells from hatched eggs without membranes laid by control hens averaged 0.26 mm $\pm$ 0.02 standard deviation, and those from all hen groups receiving PCB averaged 0.23 $\pm$ 0.02. Eggshells from unhatched eggs averaged 0.32 mm $\pm$ 0.02 for control hens and 0.31 mm $\pm$ 0.02 for hens receiving PCB. Dahlgren and Linder (1970) found the eggshell thickness of pheasants to be unaffected by

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weekly administration of 17 capsules containing 6 mg dieldrin, a chlorinated hydrocarbon as is the PCB compound.

Effects on Chick Behavior

When chicks were placed on a visual cliff for up to 5 minutes, significant differences ($P<0.01$) among treatment groups using chi-square analysis were found in their behavior (Table 3). Among offspring of hens on 50 mg PCB weekly, 7 jumped to the visually deep side, 11 jumped to the shallow side, and 11 made no choice in the allotted time. This was in sharp contrast to the behavior of all other groups combined, where 15 jumped deep, 132 shallow and 39 made no choice. Similar behavior changes in pheasant chicks hatched from parents given dieldrin were reported by Baxter et al. (1969).

Effects on Chick Weight and Survival

The weights of chicks at 6 weeks of age (Table 4) were tested by analysis of variance. There were no significant differences due to the PCB level given to cocks and no significant cock-hen interactions ($P>0.05$). However, chicks hatched from hens given 50 mg PCB were significantly lighter in weight ($P<0.05$) than those hatched from control hens and hens given 12.5 mg.

Chick survival was determined for treatment groups during the first 6 weeks of age while chicks were kept in brooders (Table 4). The survival of chicks was not related to whether cock parents received PCB, but chi-square analysis showed that significantly more ($P<0.01$) deaths occurred among chicks hatched from hens receiving 50 mg PCB. No difference ($P>0.05$) was found between mortality of chicks hatched from control hens and those hatched from hens given 12.5 mg PCB.

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Apparently 50 mg PCB given weekly to breeding hens were enough to cause a decrease in the weight of their chicks at 6 weeks of age and to cause an increase in chick mortality between hatching and 6 weeks of age. Administration of PCB decreased egg production and caused an increase in the number of eggs which were pipped but not hatched among treated groups. Behavioral changes on a visual cliff of newly-hatched chicks were also effected by PCB. However, fertility and hatchability of eggs, eggshell thickness, and weekly weights and survival of adult pheasants given PCB were unaffected.

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Table 1. Reproductive statistics from control pheasants and those given PCB.

Treatment Group	No. Eggs Per Hen Per Day	No. Eggs Set in Incubator	Fertile Eggs		Fertile Eggs Hatched		Fertile Eggs Pipped But Not Hatched	
			No.	Percent	No.	Percent	No.	Percent
0-0[a]	0.621	277	101	36	74	73	6	6
0-12.5	0.459	220	128	58	87	68	29	23
0-50	0.292	136	50	37	32	64	8	16
25-0	0.335	168	91	54	73	80	9	10
25-12.5	0.362	179	71	40	50	70	18	25
25-50	0.198	98	63	64	41	65	10	16

[a] The first number is the weekly PCB level given to cocks, the second that for hens.

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Table 2. Measurements of eggshells laid by control pheasants and those given PCB.

Treatment Group	Weekly Egg Groups 1-7		Weekly Egg Groups 8-15			
	No. Hatched Eggs	Average Thickness Without Membrane (mm)	No. Hatched Eggs	Average Thickness Without Membrane (mm)	No. Unhatched Eggs	Average Thickness With Membrane[b] (mm)
0-0[a]	41	0.25	30	0.24	63	0.32
0-12.5	42	0.24	45	0.23	28	0.31
0-50	19	0.24	12	0.24	37	0.31
25-0	30	0.27	41	0.27	27	0.33
25-12.5	17	0.23	30	0.23	28	0.31
25-50	35	0.23	5	0.24	13	0.30

[a] The first number is the weekly PCB level in mg given to cocks, the second that for hens.

[b] Unhatched eggs were measured only from weekly egg groups 8-15.

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Table 3. Behavior on the visual cliff of chicks hatched from control pheasants and those given PCB.

Treatment Group	No. Which Jumped to Visually Deep Side	No. Which Jumped to Visually Shallow Side	No. Not Jumping Within 5 Minutes
0-0[a]	2	35	8
0-12.5	3	42	8
0-50	3	6	5
25-0	5	33	15
25-12.5	5	22	7
25-50	4	5	6

[a] The first number is the weekly PCB level in mg given to cocks, the second that for hens.

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Table 4. Weights and survival of offspring from control pheasants and those given PCB, 1970.

Treatment Group	Weight At 6 Weeks (g)	No. of Chicks to Brooder	No. of Chicks Alive After 6 Weeks	Percent Survival
0-0[a]	396	73	49	67
0-12.5	378	84	49	58
0-50	303	29	3	10
25-0	389	72	50	69
25-12.5	403	50	30	60
25-50	344	40	9	22

[a] The first number is the weekly PCB level in mg given to cocks, the second that for hens.

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