

Pittsburgh PCB 12x30 activated carbon is found to be the most suitable of the commercially available carbons tested for personnel sampling of vinyl chloride, vinylidene chloride, and methyl chloride. The carbon is desorbed with CS₂ at dry ice temperature or with a thermal desorption technique.

Monitoring personnel exposure to vinyl chloride, vinylidene chloride and methyl chloride in an industrial work environment

L. W. SEVERS and L. K. SKORY
Dow Chemical U.S.A., Midland, Michigan 48640

Introduction

Late in 1973, information became available linking angiosarcoma of the liver (a rare form of cancer) with occupational exposures to high levels of vinyl chloride. Reacting to this, OSHA issued an Emergency Temporary Standard for vinyl chloride exposure, limiting them to a maximum of 50 ppm. Subsequently, a reduction of this standard was proposed and became effective whereby the permissible eight hour time-weighted average exposure may not exceed 1 ppm, with a ceiling of 5 ppm averaged over no less than a 15 minute sampling period. A program was undertaken to develop a suitable method for long and short term personnel sampling and analysis of vinyl chloride. Methods for long and short term exposure were developed in this investigation because of their relative similarity to vinyl chloride.

Breakthrough Studies

Carbon tubes gave significantly differing results

than sophisticated chlorohydrocarbon analyzers thus indicating that existing adsorbents were not adequate to quantitatively define vinyl chloride exposures to workers.

Available carbons were obtained and studied for breakthrough time by passing 200 ppm (parts per million by volume in air) vinyl chloride through a 30 ml bed of carbon at a flowrate of approximately 2 lpm (liters per minute) representing typical conditions that existed a year ago. Comparison of the parts per million breakthrough versus time is shown in Table I.^{2,3} Dow Experimental Carbon (XF-4175L), Pittsburgh MSC-V 14X40 mesh and Columbia Carbon showed the best breakthrough characteristics; however, the Dow Experimental Carbon and the MSC-V are not commercially available at this time.

Replicate samples of Columbia Carbon 13870 were tested against those of Pittsburgh



L. W. Severs is a Senior Research Scientist in the Health, Safety and Environment Department at Dow Chemical Company. He has been with Dow since 1967, joined the Health, Safety and Environment Department in 1973, and is currently responsible for the development and implementation of health and safety programs for Dow Chemical Company. He is also a member of the American Society of Safety Engineers and the American Society of Safety Engineers.



Lyman K. Skory graduated from Michigan State University in 1949 with a B.S. degree in Chemistry and from Pennsylvania State University with an M.S. degree in Organic Chemistry and joined The Dow Chemical Company, Midland, Michigan, in 1951. He is currently a Group Leader in the Dow Industrial Hygiene Laboratory, Midland. He works closely with OSHA and NIOSH in health standards programs and with Product Stewardship for Dow customers.

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TABLE I
Comparison in Parts per Million of Commercial Carbons;
30 ML Bed Volume, 200 ppm Vinyl Chloride, 1.97 lpm Flowrate

TIME (HOURS)	WESTVACO NUCHAR WYH 8x30 MESH	FISHER SCIENTIFIC COCOANUT CARBON 6x14 MESH	COLUMBIA CARBON SMALL MESH	PITTSBURGH BPL 4x10 MESH	DARCO 4x12 MESH	PITTSBURGH MSC.V 14x40 MESH	WITCO GRADE 360 10x20 MESH	DOW EXPERIMENTAL CARBON XF-4175L
0.5	< 0.05	< 0.05	< 0.05	< 0.05	< 0.02	< 0.1	< 2	< 0.1
1.0	< 0.05	< 0.05	< 0.05	0.24	< 0.02	< 0.1	2	< 0.1
1.5	0.08	< 0.05	< 0.05	0.86	0.02	< 0.1	4	< 0.1
2.0	0.53	0.21	< 0.05	1.77	0.12	< 0.1	4	< 0.1
2.5	5.22	0.95	< 0.05	—	0.51	< 0.1	12	< 0.1
3.0	26.30	5.45	0.05	3.15	11.80	< 0.1	40	< 0.1

PCB 12X30 Activated Carbon. The bed size was reduced to 1 gram of carbon contained in a stainless steel tube (14 cm x 0.25" O.D. x 0.028" wall) fitted with Swagelok caps. The test concentrations was reduced to 50 ppm vinyl chloride and the pump flowrate was reduced to 0.2 lpm. Table II shows the Pittsburgh PCB carbon to be the most suitable for eight hour time-weighted average personnel sampling. The 1 gram slugs of PCB carbon were also tested at 25 ppm with flowrates of 0.5 and 1.0 lpm. Table III gives the results of these tests. Based on these data, the following sampling conditions were recommended for the 1 gram slug of Pittsburgh PCB 12X30 Activated Carbon: (1) 0.2 lpm for 8 hours, (2) 0.5 lpm for 10 to 60 minutes, and (3) 1.0 lpm for 10 minutes.

Further breakthrough testing was conducted with different sized tubes for short time periods. The NIOSH method for vinyl chloride in air uses a 150 mg divided charcoal tube. This charcoal tube was tested with 1 ppm and 5 ppm vinyl chloride at a flowrate of 1 lpm. Figure 1 shows breakthrough within the first minute and approximately 12% breakthrough after 15 minutes. The flowrate for this size tube was reduced to 0.2 lpm and the experiment was repeated at the 5 ppm level.

Figure 2 shows that the breakthrough under these conditions is nearly 3% after 15 minutes. The NIOSH method also states that 5 liters of 200 ppm vinyl chloride can be collected at 0.05 lpm without significant amounts of vinyl chloride being found on the back section. Duplicate test runs under these conditions showed that substantial breakthrough occurs well before 5 liters of sample has been collected (Figure 2).

A direct comparison of the carbon in the NIOSH recommended tubes and PCB 12X30 was made by repacking the NIOSH tube with 150 mg of PCB carbon and repeating the experiment. Figure 1 shows that the two carbons have similar breakthrough characteristics under these conditions. However, this may be due to carbon bed size than type of carbon. The tubes could be used in a 5 ppm atmosphere at 0.05 lpm for 15 minute sampling; however, only 0.75 liter is collected. Even with a GC sensitivity of 1 µg/sample, a less than value of 0.52 ppm is obtained, which is above the action level (0.5 ppm) and much higher than the desired 0.1 ppm sensitivity. We, therefore, concluded that the 150 mg NIOSH recommended tube was inadequate for vinyl chloride sampling.

Two larger tubes of carbon were tested under similar conditions. A 600 mg charcoal

TABLE II
Percent Breakthrough for Columbia and
Pittsburgh PCB Activated Carbons at 50 ppm
Vinyl Chloride and 0.2 lpm Flowrate

TIME (HOURS)	COLUMBIA CARBON TS-570	PITTSBURGH PCB 12x30
1.0	< 0.3	< 0.3
2.5	< 0.3	< 0.3
12.0	< 0.3	< 0.3
24.0	< 0.3	< 0.3
46.0	< 0.3	< 0.3
72	—	< 0.3

TABLE III
Percent Breakthrough for One Gram of
Pittsburgh PCB 12X30 at 25 ppm Vinyl
Chloride at 0.5 and 1 lpm Flowrates

TIME (HOURS)	0.5 LPM	1.0 LPM
1.0	0.06	—
1.25	—	—
1.5	—	0.4
1.8	0.06	0.8
2.1	0.06	—
2.4	0.31	—
2.7	2.80	—
3.0	13.01	—

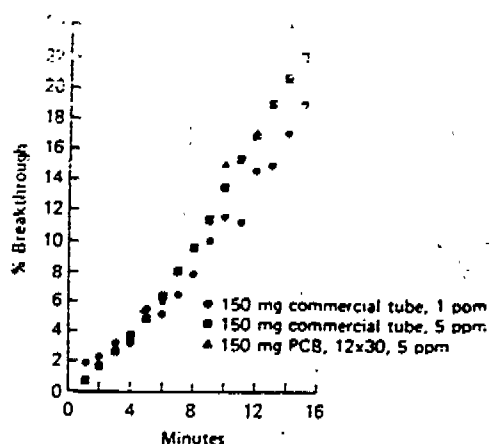


Figure 1-Vinyl chloride breakthrough, 150 mg tube at 1 lpm.

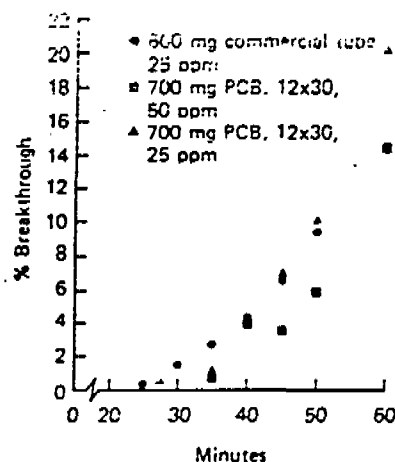


Figure 3-Vinyl chloride breakthrough, 600 and 700 mg tubes at 1 lpm.

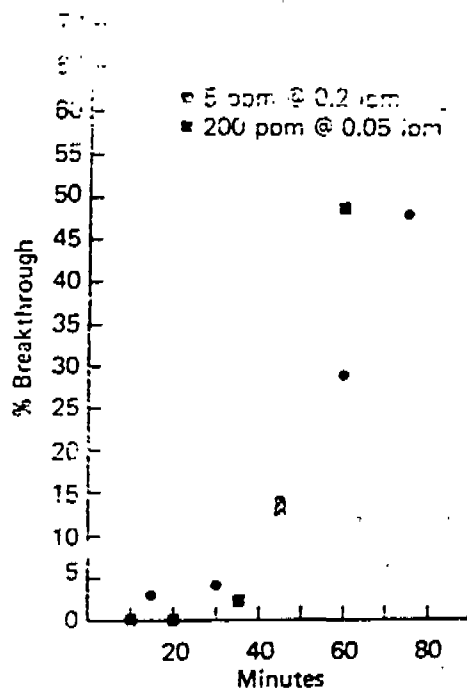


Figure 2-Vinyl chloride breakthrough, 150 mg tube at reduced flowrates.

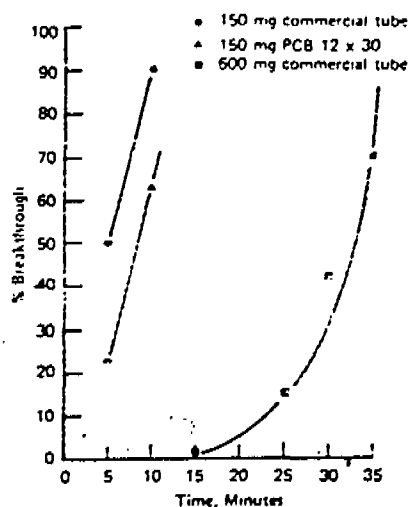


Figure 4-Methyl chloride breakthrough, 100 ppm at 0.2 lpm for 150 and 600 mg tubes.

tube is manufactured by the makers of the 150 mg OOSH tube. A tube was also made from a disposable glass transfer pipette containing approximately 700 mg of the Pittsburgh PCB

12X30 Activated Carbon. Figure 3 shows the results for testing these tubes at 25 and 50 ppm vinyl chloride at 1 lpm. Either tube could be used for the 15 minute sampling and at

reduced flowrates (0.2 or 0.1 lpm) for extended periods of time (7-8 hours) at these levels.

Two items of interest are evident from Figures 1 and 3: (1) the breakthrough time and percentages are not directly proportional to those seen for the 150 mg tube, indicating additional factors in adsorption (possibly tube dimension and/or a change of carbon), and (2) the breakthrough curves for 25 and 50 ppm are almost identical, as are those for 1 and 5 ppm on the 150 mg tube. These data indicate that the percentage breakthrough for a given flowrate and tube size is independent of the concentration for at least the 1-50 ppm range.

Similar breakthrough testing was performed for methyl chloride and vinylidene chloride. Methyl chloride was tested at 100 ppm (TLV) for various flowrates and tube sizes. Figure 4 shows 150 mg tubes are not suitable for methyl chloride sampling at the current allowable levels, although the PCB carbon appears to have better retention capability. The 600 mg tube might be used for short term samples. Reducing the flowrate below 0.2 lpm would increase the time of sampling. However, by using the 1 gram slug of PCB 12X30 and reducing the flowrate to 0.1 lpm, sampling times of up to 80-90 minutes are feasible (Figure 5).

This figure also shows the increase in sampling time possible by decreasing the flowrate from 0.2 to 0.1 lpm. A sample test of 200 ppm at 0.2 lpm flowrate is also known for comparison. If methyl chloride and vinyl chloride are to be sampled in the same area, the methyl chloride will be the limiting factor in choosing a sampling period and flowrate. Only by increasing the amount of carbon and/or greatly reducing the flowrate can eight hour samples be obtained.

(Methyl chloride breakthrough was also determined for a 1 gram slug of PCB 12X30 at the 50 ppm level. The breakthrough curves are for this test shown in Figure 5 for 50 ppm. This figure demonstrates that percentage breakthrough for a given flowrate and tube size is independent of concentration.)

Breakthrough characteristics for vinylidene chloride are much less severe than for methyl chloride and vinyl chloride throughout testing. Figure 6 shows that the 150 mg size tube may be used for short term samples (10-15 minutes) at 1 lpm. However, the PCB 12X30 carbon begins to show definite ad-

vantages over the carbon used in the commercial tube. Reducing the flowrate will, of course, allow for longer sample times, but not long enough for eight hour samples. The 600 mg size tube has very good retention capability for vinylidene chloride. A test of 31 ppm vinylidene chloride samples at 1 lpm showed less than 0.08% breakthrough after 75 minutes.

Thus, with a reduction in flowrate to 0.1 or 0.2 lpm, eight hour sampling is quite possible. (These data suggest that the type of carbon has been changed in this size tube because a similar test run a year ago showed substantial breakthrough after 60 minutes. The 150 mg and 600 mg tubes were SKC Lot 104 and Lot 105, respectively. SKC has confirmed a

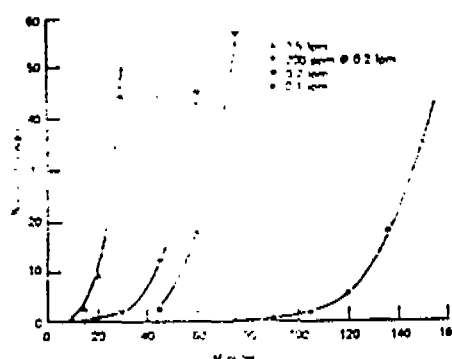


Figure 5—Methyl chloride breakthrough, 100 ppm at various flows for 1 gram Pittsburgh PCB 12X30 activated carbon.

change of carbon between the two lots. This change in carbon was recommended to the supplier after NIOSH had received our recommendations.) One gram of the PCB 12X30 carbon in a stainless steel slug has been run under these conditions for eight hours with no detectable breakthrough.

Carbon Disulfide Desorption

A desorption technique for vinyl chloride was devised that would ensure the integrity of the sample. An experiment in one of our laboratories indicated a 98% recovery was obtained when the carbon was slowly added to the CS_2 cooled in a dry ice/acetone slurry.² The following procedure has since proven to be satisfactory: (1) cool 10 ml of CS_2 to dry ice temperature; (2) slowly add the carbon to the cold CS_2 ; (3) agitate for 30 minutes, keeping cold; (4)

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refrigerate until analysis, analyzing from a wet ice bath. It was found that by thus minimizing the heat generation during desorption, replicate samples have given a recovery range of 93-101%.

This same desorption technique has been applied to samples of methyl chloride. Over a period of time, 25 known samples have shown a recovery range of 70-97% with an average of $83\% \pm 8\%$ (one standard deviation). The wide range of recovery values has not yet been accounted for.

Desorption of vinylidene chloride samples is accomplished by the same procedure. A wet ice bath can be used, unless one or more of the low boiling gases are to be determined also.

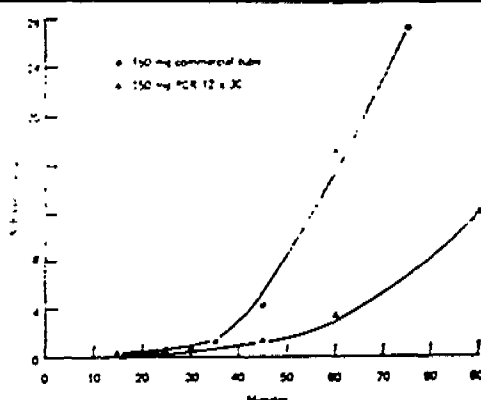


Figure 6—Vinylidene chloride breakthrough, 20 ppm at 1 ppm for 150 mg tube.

Recoveries for vinylidene chloride in CS_2 have a range of 95-100%.

Standard Preparation

The technique was adapted from a procedure used at Dow for many years.⁵ Vinyl chloride is condensed in a tared, narrow neck ampule in dry ice. The ampule is flame sealed and weighed to constant weight. A small Swagelok fitting is carefully placed over the neck of the ampule and the whole assembly gently lowered into a measured volume of CS_2 . The ampule is then broken by shaking the bottle. No bubbles are seen during this operation, indicating that the vinyl chloride is completely dissolving in CS_2 .

Methyl chloride standards can be made by following the same procedure.

Standards of vinylidene chloride are routinely prepared by injecting a measured volume of vinylidene chloride into a known volume of CS_2 with a microliter syringe.

There was some question about standards made by injecting a known volume of gas into a known volume of CS_2 in a sealed serum vial at room temperature and wet ice temperature so the above technique was utilized. Condensing the vinyl chloride in dry ice and using a 10 ml glass syringe cooled to dry ice temperature to prepare a standard proved difficult to duplicate.

Sample Storage

The flood of inquiries from industrial hygienists, both from industry and government regarding personnel samples taken at one location and forwarding to a laboratory at another location for analysis, set the priority for determining sample storage characteristics. The backlog of samples taken in our own company and limitations of available manpower and instrumentation further substantiated the need to know how much absorbed vapor was being lost during storage or shipment of caroon tubes.

Sample of 10 liters of vinyl chloride at 1 ppm on the PCB 12X30 activated carbon were stored in polyethylene tubing for one to three weeks; 10 to 20% loss was noted with increased storage time. The effects of four weeks storage at room temperature were checked with both the stainless steel and glass tubes, and no apparent sample degradation was observed.

However, it was found that if samples were removed from their tubes and placed in small vials sealed with Polyseal caps, up to 40% of the initially loaded vinyl chloride was lost after one week of storage in the freezer. Samples, therefore, should be kept sealed in the original sample tubes until desorbed.

The storage effects on vinylidene chloride in glass tubes proved to be within the error ($\pm 5\%$) of the analytical method. Effects on methyl chloride samples have not yet been determined.

Thermal Desorption

Thermal desorption was investigated as an alternate to CS_2 desorption. Thermal desorption, if perfected, offers several advantages over carbon desorption, such as reduced health hazards.

The experimental apparatus consists of a wettest, dry test meter for measuring gas volumes, a muffle furnace and a Saran[®] film bag to collect the expelled vapors. The inlet and outlet tubes are of stainless steel as are the sample tubes. Prepurified nitrogen at a flowrate of 500-800 ml/minute is used as the purge gas. The oven temperature was initially set at 225°C, however, the low and erratic recoveries obtained were eliminated by raising the temperature to 430°C.

The desorption procedure is as follows. Place a 10 liter Saran film bag at the exit end of the outlet tube; connect the sample tube in the line and start the nitrogen flow. Purge the system with approximately 1 liter of nitrogen before placing the tube in the furnace. After placing the tube in the furnace, collect the expelled vapors until the desired volume is obtained. Divert the nitrogen flow from the collection bag and remove the bag for analysis.

For vinyl chloride and vinylidene chloride, the recoveries were 92% $\pm$ 3.5% (one standard deviation) for 50 and 100 ppm samples, respectively. For methyl chloride, the recovery was 80% $\pm$ 5% (one standard deviation). This was true for 50 and 100 ppm samples, respectively. However, for short term samples, e.g., 10 liters collected at 1 lpm, it was found that more nitrogen was needed. At 25 ppm, 4 liters of nitrogen gave 90% recovery. At the 10 ppm level, 10 liters of nitrogen had to be used to obtain 93% recovery and at 1 ppm, 12 liters of nitrogen were needed to obtain this recovery. Fifteen replicate samples of 1 ppm vinyl chloride were analyzed by this method. The average recovery was 92% $\pm$ 3.5% (one standard deviation). Subsequent series of tests have shown a recovery factor of 80% $\pm$ 5%. Attempts were made to find the cause of the decrease. Neither the use of glass tubing nor deactivated and conditioned stainless steel tubing increased the recovery factor to its original value. Replacement of the inlet/outlet tubing and the nitrogen supply has had the same negative effect. Although this new factor is lower than what was originally found, it has been reproducible and could, therefore, be effectively used as the recovery factor for this technique.

The recoveries of methyl chloride and vinylidene chloride have also been determined by this technique. A recovery of 75% $\pm$ 5% is obtained (one standard deviation) for 6 liters of both 50 and 100 ppm methyl chloride desorbed with 10 liters of nitrogen. Replicate samples of 10 ppm vinylidene chloride gave a

recovery of 88% $\pm$ 5% (one standard deviation) when desorbed with 10 liters of nitrogen.

Analytical Conditions

Analysis is performed by gas chromatography/flame ionization detection. No one column or set of conditions can cover all situations involving analysis for vinyl chloride. A variety of columns have been used depending upon the situation. Figures 7-11 show some of the columns that can be used under varying circumstances.

Breakthrough was determined by periodically injecting air samples drawn from a glass tee placed behind the carbon tube. The following GC conditions were employed.

Column: 6' x 1/8" stainless steel 20%
DC 200 on Chromosorb W,
80-100 Mesh

Column
Temperature: 40°C

Detector
Temperature: 250°C
Sample Size: 0.5 cc with GSV, or 1.0 ml
Pressure-Lok Syringe

Sample analysis was originally performed on this same column at 40°C. Both vinyl chloride and vinylidene chloride can be determined (Figure 7); however, light hydrocarbons may interfere with the vinyl chloride determination. Such interferences were eliminated by employing the following conditions (Figures 8 and 9):

Column: 20' x 1/8" stainless steel
Carbowax 4000 on Supelco-
port #6, 100 Mesh

Column
Temperature: 80°C
Injection
Port: Unheated, on-column injection
Sample Size: 2 microliters

Other parameters were the same as previously stated. Temperature and carrier flow are quite critical because the vinylidene chloride is not completely separate from the CS₂.

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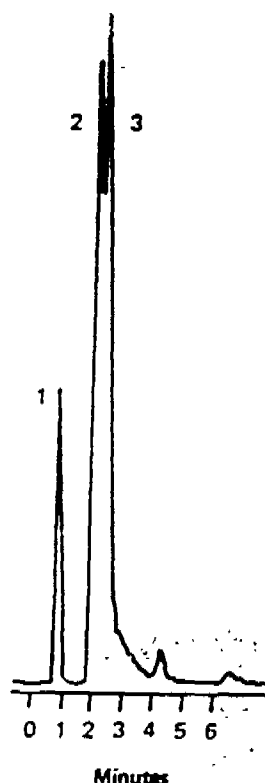


Figure 7—Chromatogram of components on DC 200 column; (1) vinyl chloride, (2) vinylidene chloride, (3) CS_2 .

small changes in either of these parameters results in loss of resolution.

These conditions have proved satisfactory except for samples containing methyl chloride, dichlorodifluoromethane and/or dimethyl ether in addition to vinyl chloride. For these situations satisfactory separations have been achieved under the following conditions (Figures 10 and 11).

Column: 5' x 1/8" stainless steel
Porapak QS, 80/100 Mesh
Carrier Flow: 40 ml/min N_2
Temperature: 70°C (methyl chloride, dichlorodifluoromethane, vinyl chloride)
115°C (methyl chloride, dimethyl ether, vinyl chloride)

Conclusions

When the vinyl chloride situation arose it was found that existing techniques for personnel sampling and analysis of vinyl chloride exposures were found to be inadequate because vinyl chloride vapor was easily lost from the sample at several different stages in the collection and analysis.

This study evaluated numerous adsorbing media and found that one specific coconut carbon, Pittsburgh PCB 12X30 Activated Carbon, is the most suitable of the commercially available carbons tested for personnel sampling of vinyl chloride and related compounds. One gram of this carbon packed in a stainless steel tube (14 cm x 0.25" O.D. x 0.028" wall) can be used satisfactorily to collect vinyl chloride, vinylidene chloride and methyl chloride according to the sampling conditions developed herein. The study developed improved techniques of desorbing the carbon using CS_2 at dry ice temperature. Thermal desorption was also investigated as an alternative to CS_2 desorption.

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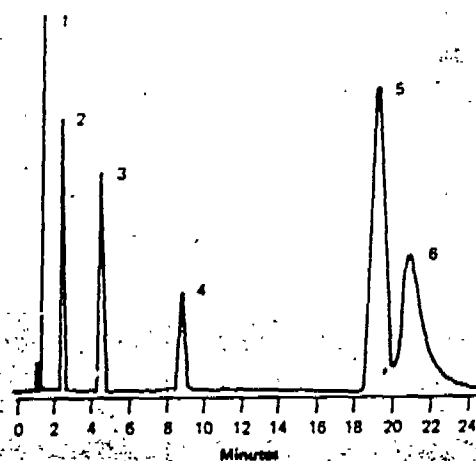


Figure 8—Chromatograms of components in air on Carbowax 4000 column; (1) isobutane, (2) vinyl chloride, (3) vinylidene chloride, (4) trans-1, 2-dichloroethylene, (5) methacrylonitrile, (6) acrylonitrile.

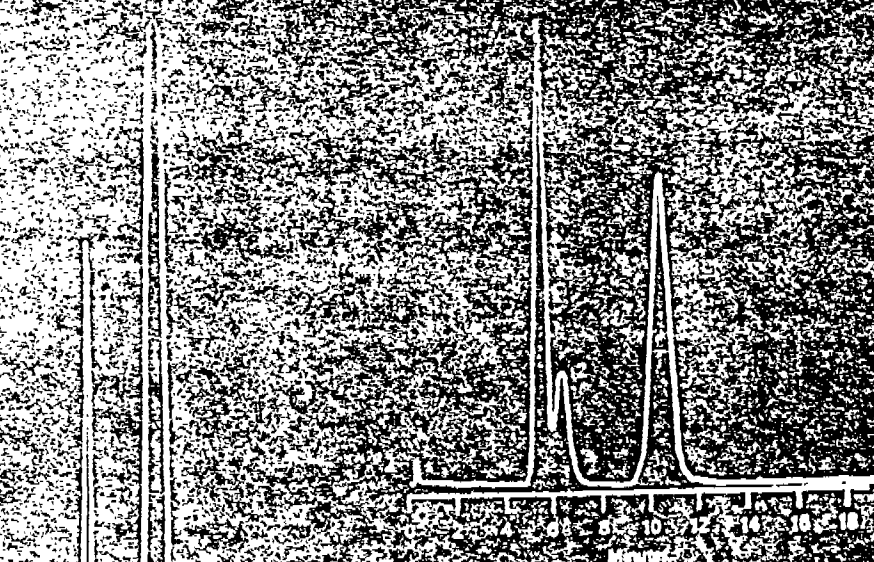


Figure 10. Chromatogram of methyl chloride and vinyl chloride in the presence of Freon 12 on Porapak Q5 column: (1) methyl chloride, (2) Freon 12, (3) vinyl chloride.

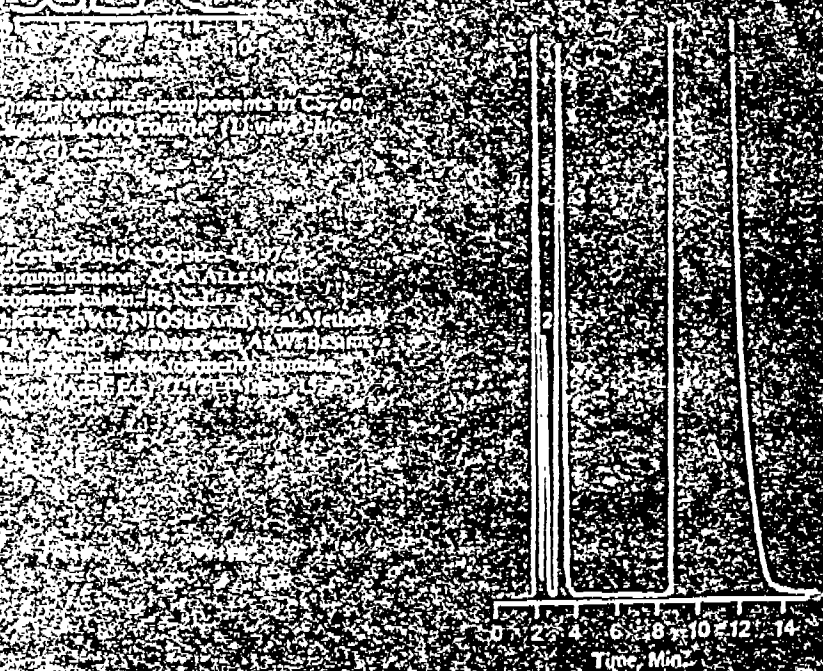


Figure 11. Chromatogram of methyl chloride and vinyl chloride in presence of dimethyl ether on Porapak Q5 column: (1) methyl chloride, (2) dimethyl ether, (3) vinyl chloride.

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- References:
1. J. Chromatogr., 1970, 40, 177-180.
 2. Private communication, A. S. Allen.
 3. Private communication, R. N. Hill.
 4. J. Chromatogr., 1971, 50, 100-105.
 5. J. Chromatogr., 1971, 50, 106-110.
 6. J. Chromatogr., 1971, 50, 111-115.
 7. J. Chromatogr., 1971, 50, 116-120.
 8. J. Chromatogr., 1971, 50, 121-125.
 9. J. Chromatogr., 1971, 50, 126-130.
 10. J. Chromatogr., 1971, 50, 131-135.
 11. J. Chromatogr., 1971, 50, 136-140.
 12. J. Chromatogr., 1971, 50, 141-145.
 13. J. Chromatogr., 1971, 50, 146-150.
 14. J. Chromatogr., 1971, 50, 151-155.
 15. J. Chromatogr., 1971, 50, 156-160.
 16. J. Chromatogr., 1971, 50, 161-165.
 17. J. Chromatogr., 1971, 50, 166-170.
 18. J. Chromatogr., 1971, 50, 171-175.
 19. J. Chromatogr., 1971, 50, 176-180.
 20. J. Chromatogr., 1971, 50, 181-185.
 21. J. Chromatogr., 1971, 50, 186-190.
 22. J. Chromatogr., 1971, 50, 191-195.
 23. J. Chromatogr., 1971, 50, 196-200.
 24. J. Chromatogr., 1971, 50, 201-205.
 25. J. Chromatogr., 1971, 50, 206-210.
 26. J. Chromatogr., 1971, 50, 211-215.
 27. J. Chromatogr., 1971, 50, 216-220.
 28. J. Chromatogr., 1971, 50, 221-225.
 29. J. Chromatogr., 1971, 50, 226-230.
 30. J. Chromatogr., 1971, 50, 231-235.
 31. J. Chromatogr., 1971, 50, 236-240.
 32. J. Chromatogr., 1971, 50, 241-245.
 33. J. Chromatogr., 1971, 50, 246-250.
 34. J. Chromatogr., 1971, 50, 251-255.
 35. J. Chromatogr., 1971, 50, 256-260.
 36. J. Chromatogr., 1971, 50, 261-265.
 37. J. Chromatogr., 1971, 50, 266-270.
 38. J. Chromatogr., 1971, 50, 271-275.
 39. J. Chromatogr., 1971, 50, 276-280.
 40. J. Chromatogr., 1971, 50, 281-285.
 41. J. Chromatogr., 1971, 50, 286-290.
 42. J. Chromatogr., 1971, 50, 291-295.
 43. J. Chromatogr., 1971, 50, 296-300.
 44. J. Chromatogr., 1971, 50, 301-305.
 45. J. Chromatogr., 1971, 50, 306-310.
 46. J. Chromatogr., 1971, 50, 311-315.
 47. J. Chromatogr., 1971, 50, 316-320.
 48. J. Chromatogr., 1971, 50, 321-325.
 49. J. Chromatogr., 1971, 50, 326-330.
 50. J. Chromatogr., 1971, 50, 331-335.
 51. J. Chromatogr., 1971, 50, 336-340.
 52. J. Chromatogr., 1971, 50, 341-345.
 53. J. Chromatogr., 1971, 50, 346-350.
 54. J. Chromatogr., 1971, 50, 351-355.
 55. J. Chromatogr., 1971, 50, 356-360.
 56. J. Chromatogr., 1971, 50, 361-365.
 57. J. Chromatogr., 1971, 50, 366-370.
 58. J. Chromatogr., 1971, 50, 371-375.
 59. J. Chromatogr., 1971, 50, 376-380.
 60. J. Chromatogr., 1971, 50, 381-385.
 61. J. Chromatogr., 1971, 50, 386-390.
 62. J. Chromatogr., 1971, 50, 391-395.
 63. J. Chromatogr., 1971, 50, 396-400.
 64. J. Chromatogr., 1971, 50, 401-405.
 65. J. Chromatogr., 1971, 50, 406-410.
 66. J. Chromatogr., 1971, 50, 411-415.
 67. J. Chromatogr., 1971, 50, 416-420.
 68. J. Chromatogr., 1971, 50, 421-425.
 69. J. Chromatogr., 1971, 50, 426-430.
 70. J. Chromatogr., 1971, 50, 431-435.
 71. J. Chromatogr., 1971, 50, 436-440.
 72. J. Chromatogr., 1971, 50, 441-445.
 73. J. Chromatogr., 1971, 50, 446-450.
 74. J. Chromatogr., 1971, 50, 451-455.
 75. J. Chromatogr., 1971, 50, 456-460.
 76. J. Chromatogr., 1971, 50, 461-465.
 77. J. Chromatogr., 1971, 50, 466-470.
 78. J. Chromatogr., 1971, 50, 471-475.
 79. J. Chromatogr., 1971, 50, 476-480.
 80. J. Chromatogr., 1971, 50, 481-485.
 81. J. Chromatogr., 1971, 50, 486-490.
 82. J. Chromatogr., 1971, 50, 491-495.
 83. J. Chromatogr., 1971, 50, 496-500.
 84. J. Chromatogr., 1971, 50, 501-505.
 85. J. Chromatogr., 1971, 50, 506-510.
 86. J. Chromatogr., 1971, 50, 511-515.
 87. J. Chromatogr., 1971, 50, 516-520.
 88. J. Chromatogr., 1971, 50, 521-525.
 89. J. Chromatogr., 1971, 50, 526-530.
 90. J. Chromatogr., 1971, 50, 531-535.
 91. J. Chromatogr., 1971, 50, 536-540.
 92. J. Chromatogr., 1971, 50, 541-545.
 93. J. Chromatogr., 1971, 50, 546-550.
 94. J. Chromatogr., 1971, 50, 551-555.
 95. J. Chromatogr., 1971, 50, 556-560.
 96. J. Chromatogr., 1971, 50, 561-565.
 97. J. Chromatogr., 1971, 50, 566-570.
 98. J. Chromatogr., 1971, 50, 571-575.
 99. J. Chromatogr., 1971, 50, 576-580.
 100. J. Chromatogr., 1971, 50, 581-585.
 101. J. Chromatogr., 1971, 50, 586-590.
 102. J. Chromatogr., 1971, 50, 591-595.
 103. J. Chromatogr., 1971, 50, 596-600.
 104. J. Chromatogr., 1971, 50, 601-605.
 105. J. Chromatogr., 1971, 50, 606-610.
 106. J. Chromatogr., 1971, 50, 611-615.
 107. J. Chromatogr., 1971, 50, 616-620.
 108. J. Chromatogr., 1971, 50, 621-625.
 109. J. Chromatogr., 1971, 50, 626-630.
 110. J. Chromatogr., 1971, 50, 631-635.
 111. J. Chromatogr., 1971, 50, 636-640.
 112. J. Chromatogr., 1971, 50, 641-645.
 113. J. Chromatogr., 1971, 50, 646-650.
 114. J. Chromatogr., 1971, 50, 651-655.
 115. J. Chromatogr., 1971, 50, 656-660.
 116. J. Chromatogr., 1971, 50, 661-665.
 117. J. Chromatogr., 1971, 50, 666-670.
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 183. J. Chromatogr., 1971, 50, 996-1000.